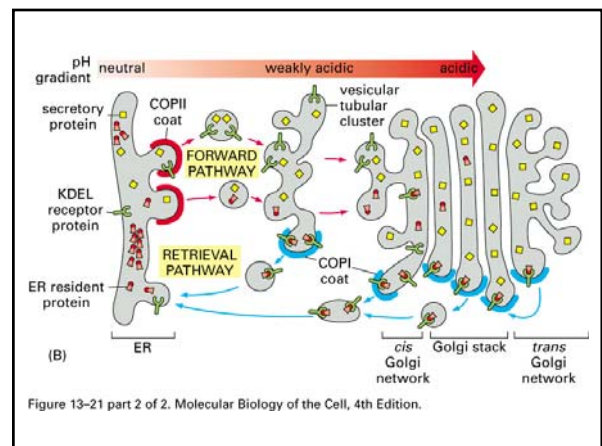
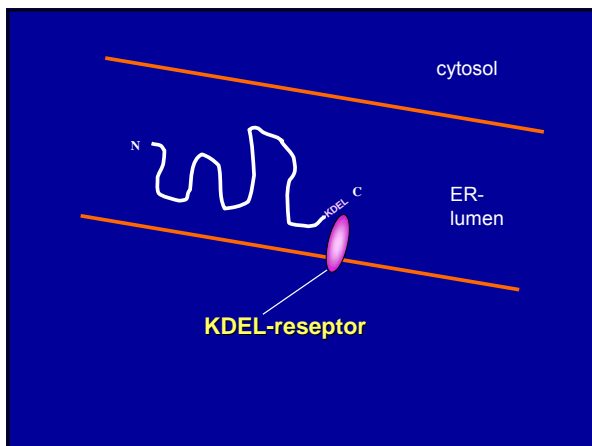
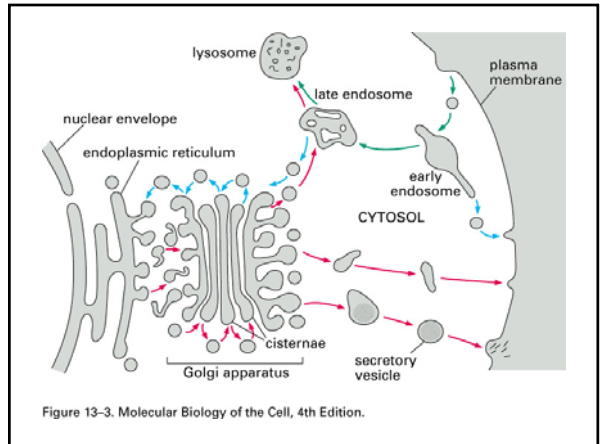
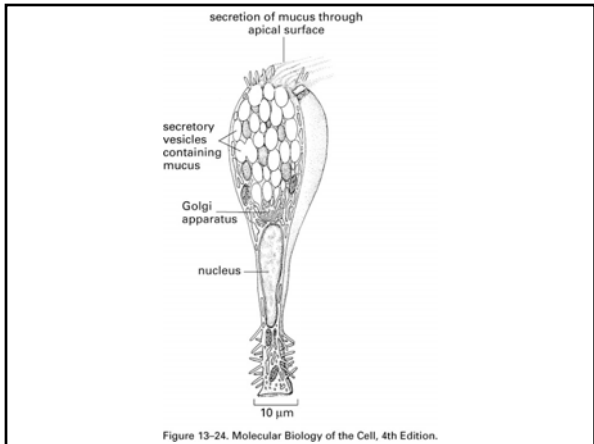
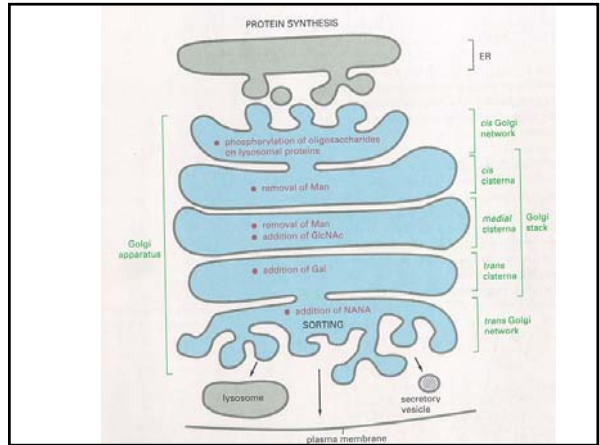
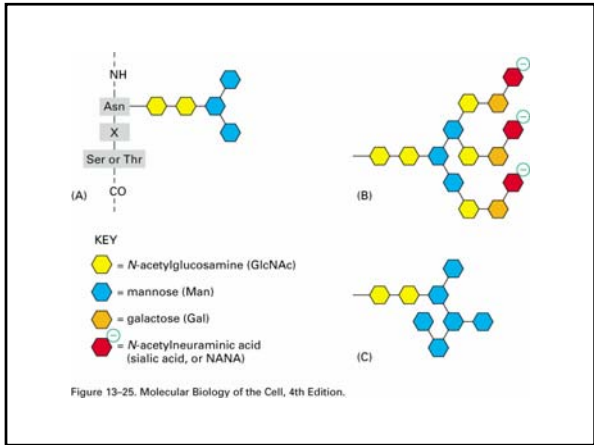
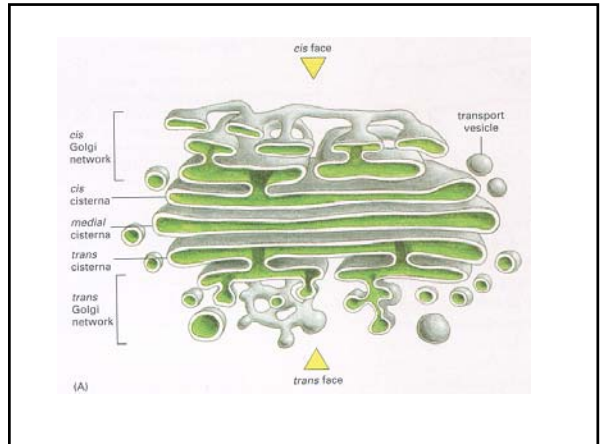
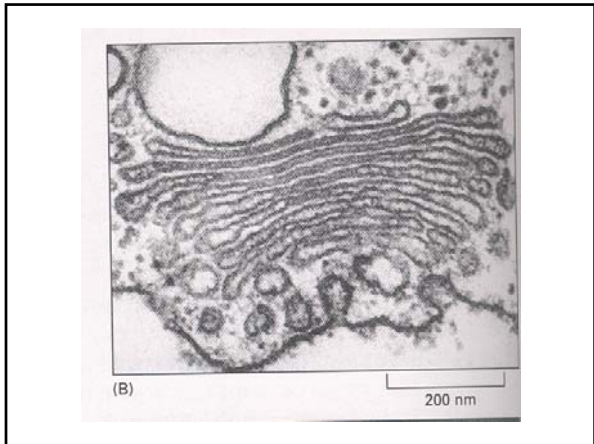
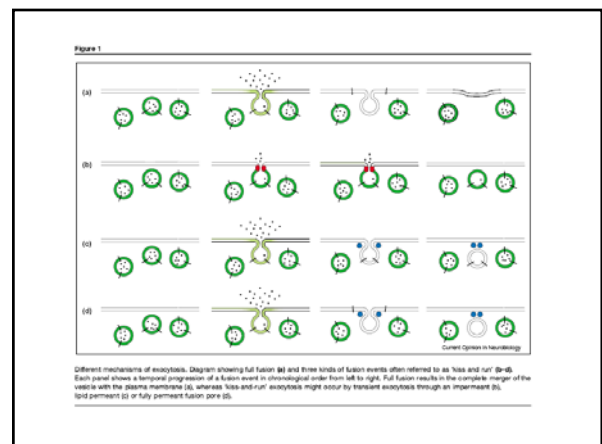
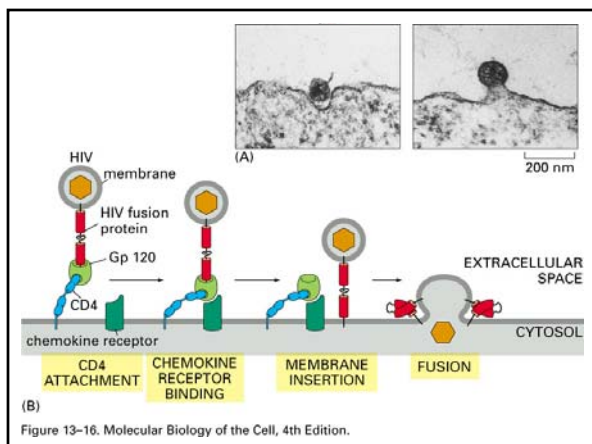
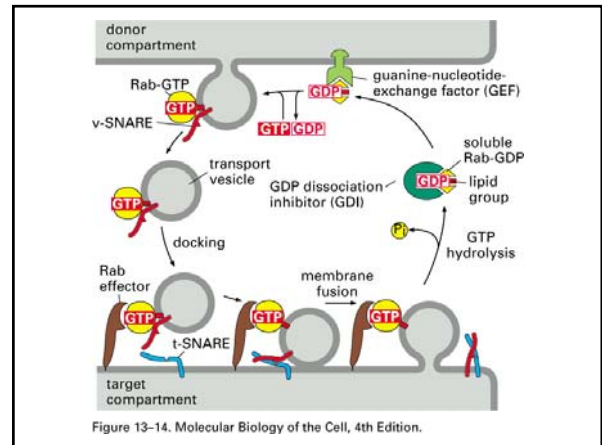
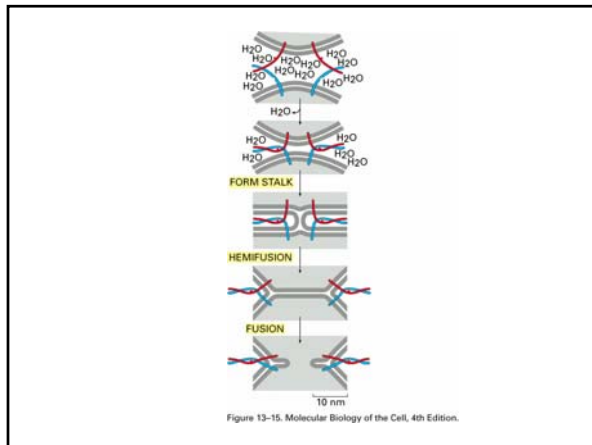
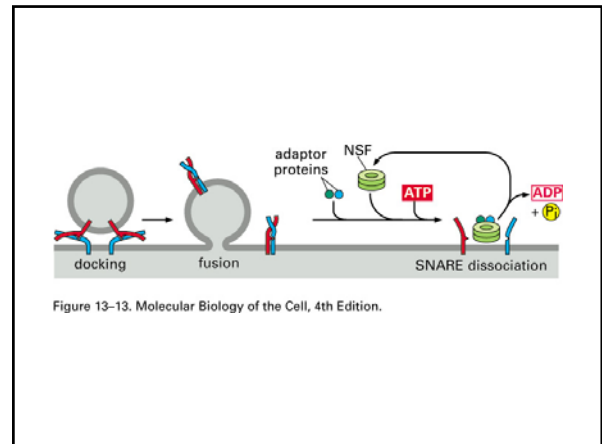
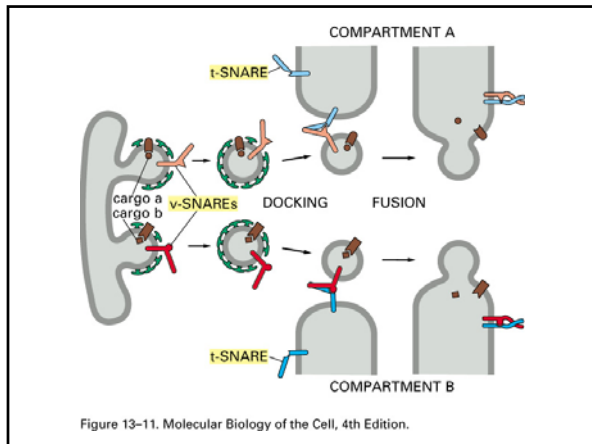


Retensjon i ER-lumen

- * Spesifikk sekvens: KDEL
- * C-terminal







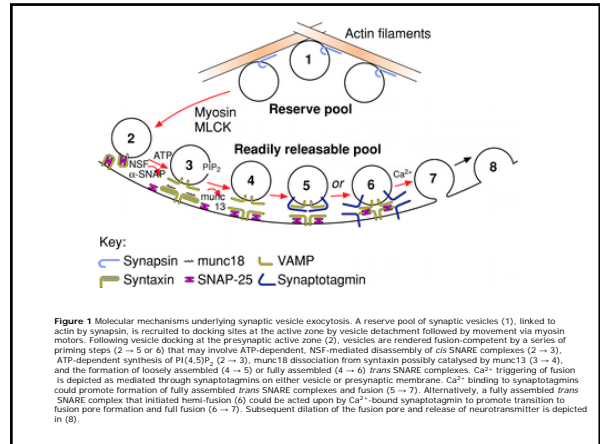
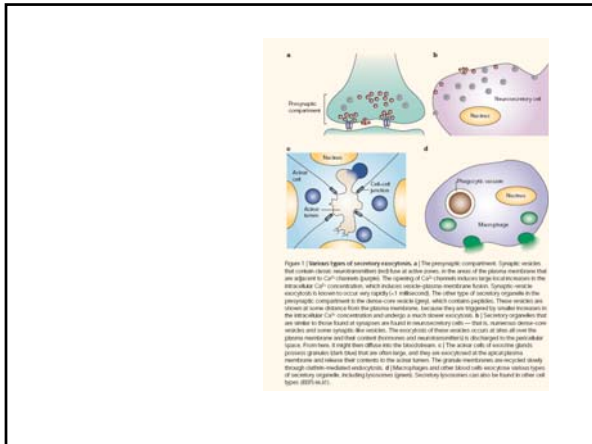
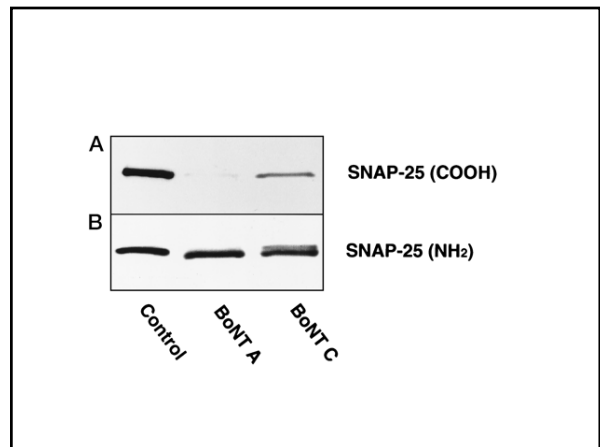
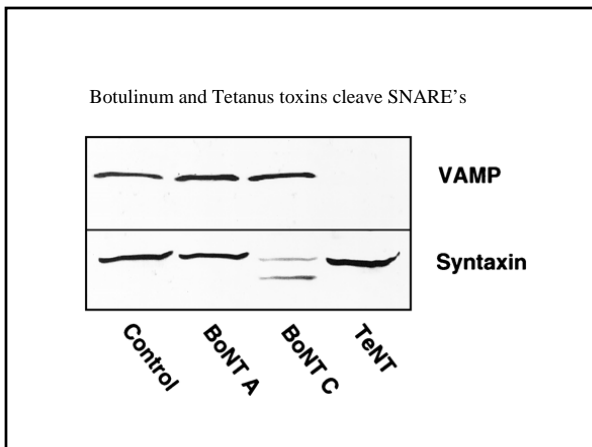
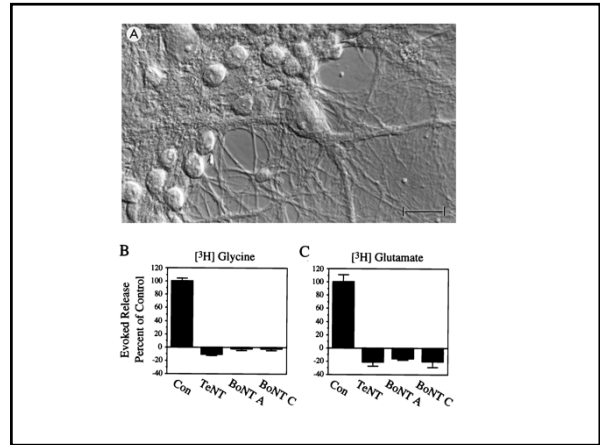
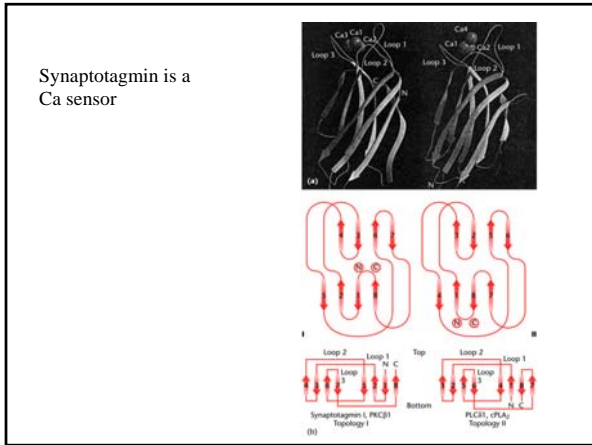
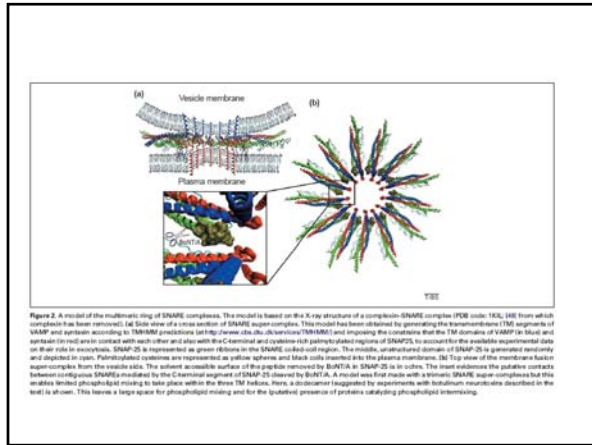
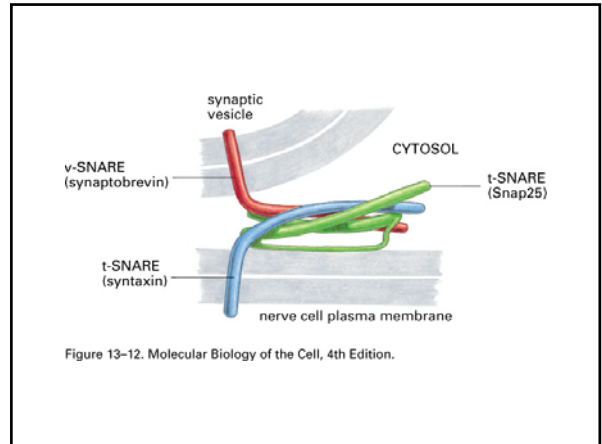
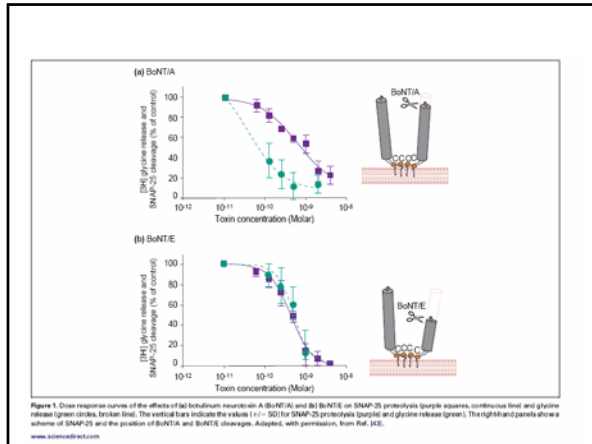


Figure 1 Molecular mechanisms underlying synaptic vesicle exocytosis. A reserve pool of synaptic vesicles (1), linked to actin by synapsin, is recruited to docking sites at the active zone by vesicle detachment followed by movement via myosin motors. Following vesicle docking at the presynaptic active zone (2), vesicles are rendered fusion-competent by a series of priming steps (2 → 5 or 6) that may involve ATP-dependent NSF-mediated disassembly of cis-SNARE complexes (2 → 3), ATP-dependent synthesis of PI(4,5)P₂ (2 → 3), munc18 dissociation from syntaxin possibly catalyzed by munc13 (3 → 4), and the formation of loosely assembled (4 → 5) or fully assembled (4 → 6) trans-SNARE complexes. Ca²⁺ triggering of fusion is depicted as mediated through synaptotagmins on either vesicle or presynaptic membrane. Ca²⁺ binding to synaptotagmins could promote formation of fully assembled trans-SNARE complexes and fusion (5 → 7). Alternatively, a fully assembled trans-SNARE complex that initiated hemifusion (6) could be acted upon by Ca²⁺-bound synaptotagmin to promote transition to fusion pore formation and full fusion (6 → 7). Subsequent dilation of the fusion pore and release of neurotransmitter is depicted in (8).





Regulert eksocytose

