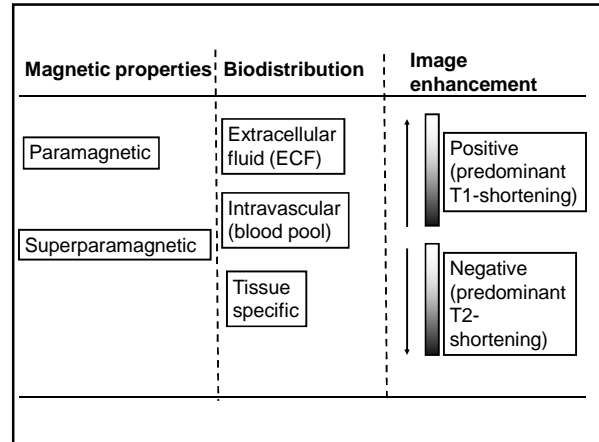


FYS-KJM 4740

MR-teori og medisinsk diagnostikk

Kap 11 (nytt)
MR kontrastmidler



MR Contrast Agents on the market or in clinical development*

Short Name	Generic Name	Trade Name	Enhancement Pattern (primary)
ECF agents:			
GdDTPA	gadopentetate dimeglumine	Magnevist	positive
GdDOTA	gadoterate meglumine	Dotarem	positive
GdDTPA-BMA	gadodiamide injection	Omniscan	positive
GdHP-DO3A	gadoterol injection	Proflance	positive
GdDTPA-BMEA	gadoversetamide	Optimark	positive
GdDO3A-butriol	gadobutrol	Gadovist	positive
GdBOPTA/Dimeg	gadobenate dimeglumine	MultiHance	positive
Organ specific agents:			
MnDPDP	mangafodipir trisodium	Testascan	positive
GdEOB-DTPA	gadovetic acid	Eovist	positive
GdBOPTA/Dimeg	gadobenate dimeglumine	MultiHance	positive
AMI-25	ferumoxides (SPIC)	Endorem/Feridex	negative
SHU 555A*	ferumoxan (SPIC)	Resovist	negative
AMI-227*	Ferumoxtran	Sinerem/Combidex	negative
SHU 555 C*	Ferucartran		Positive/negative
Blood pool agents:			
MS-325		Angiomark	positive
gadomer-17		----	positive
P792		----	positive

* adapted from Rinck et al. 'Magnetic Resonance in Medicine' (2001) ** also blood pool agents

MR Contrast Agents (MRCA)

- A contrast agent is nothing more than a catalyst that decreases the T_1 and/or T_2 of the tissue protons
- T_1 and T_2 relaxation are NOT independent processes
- T_1 cannot be reduced without reducing T_2
- T_2 can be reduced without reducing T_1

Background & Theory

Contrast Agent Relaxivity *in vitro*

MRCA effectiveness (ability to alter $1/T_1$ and $1/T_2$) *in vitro* - in a homogeneous medium - is a linear function of [CA]:

Commonly use 'relaxation rate' ($1/T_{1,2}$) due to linear relationship with [CA]

Background & Theory

Contrast Agent Relaxivity *in vitro*

In vitro MRCA relaxivity, r_1 and r_2 :

$$\frac{1}{T_{1,2}} = r_{1,2} \cdot [CA] + \left(\frac{1}{T_{1,2}} \right)_{pre}$$

- $r_1 = T_1$ relaxivity; $r_2 = T_2$ relaxivity
- $r_{1,2}$ is due to dipolar (short range) interactions
- $r_{1,2}$ must be specified at a given temp, field strength and medium

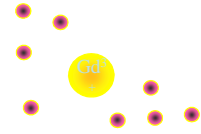
Contrast agents - basic principles

Two types of magnetic effects are used clinically to induce T_2 - and T_1 -shortening:

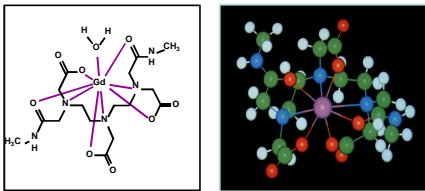
- Paramagnetism
- Superparamagnetism

Paramagnetic ions

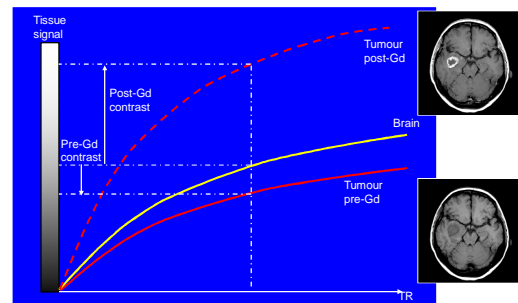
- Metal ions with unpaired electrons:
 - gadolinium (Gd^{3+})
 - iron (Fe^{2+}/Fe^{3+})
 - manganese (Mn^{2+})
- Magnetic moment of unpaired electrons is \gg magnetic moment of protons
- Dipolar magnetic interaction between electrons and water protons



Metal chelate – use of biocompatible ligand



Paramagnetic chelates enhance contrast in T1-w images



Paramagnetic ions

Typical in vitro relaxivity of small MW gadolinium chelates :

$$r_1 \cong 4 \text{ mM}^{-1}\text{s}^{-1}$$

$$r_2 \cong 4.5 \text{ mM}^{-1}\text{s}^{-1}$$

- Relaxivity is limited by rapid tumbling rate of the Gd-chelate
- T_1 relaxivity can be increased by selective binding to macromolecules (proteins) or by combining multiple Gd-ions in a rigid structure (polymers)
- Will also modify biodistribution from extracellular to intravascular.

Superparamagnetic Iron Oxide Particles;

- Iron oxide particles (nanoparticles) made up of several thousand magnetic ions in the form of magnetite or maghemite crystals
- Much larger magnetic moment than paramagnetic agents
- Developed as liver/spleen/lymph node specific (T_2) or blood pool (T_1, T_2) agents
- Imaging effect and biodistribution is size dependent

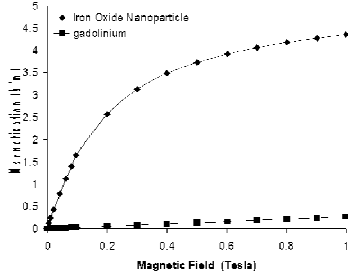
Typical in vitro relaxivity of iron oxide nanoparticles :

$$r_1 \cong 20 \text{ mM}^{-1}\text{s}^{-1}$$

$$r_2 \cong 40 \text{ mM}^{-1}\text{s}^{-1}$$

Background & Theory

Iron oxides have much larger magnetic moment than gadolinium and are therefore much more potent relaxation enhancers



Dipolar relaxivity is a complex function of CA properties

$$r_1 = A \frac{q}{T_{1m} + \tau_m} \quad r_2 = \frac{q}{\tau_m} \left[\frac{(\tau_m^{-2} + \tau_{2m}^{-2} - \tau_{1m}^{-2} + \Delta\omega_m^2)}{(\tau_m^{-4} + \tau_{2m}^{-4})^2 + \Delta\omega_m^2} \right]$$

Important dependence of 'effective' correlation rate of molecule:

$$1/\tau_c = 1/\tau_r + 1/\tau_s + 1/\tau_m$$

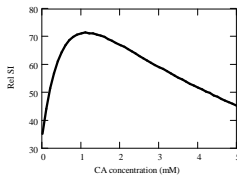
1/T_{1m} and 1/T_{2m} dependent on spectral density functions of the form:

$$\frac{\tau_c}{1 + \omega_I^2 \tau_c^2} \quad \frac{\tau_e}{1 + \omega_S^2 \tau_s^2}$$

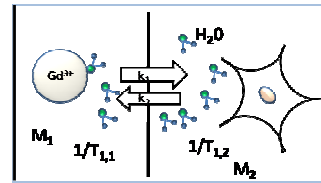
In vivo CA effects

e.g. spin echo sequence:

$$SI(C) = \rho (1 - \exp(-(R_1^0 + r_1 C)TR)) \exp(-(R_2^0 + r_2 C)TE)$$



Water exchange effects



$$1/\tau = k_1 + k_2$$

Fast exchange: $\frac{1}{\tau} \gg 1/T_{1,1} - 1/T_{1,2}$

Slow exchange: $\frac{1}{\tau} \ll 1/T_{1,1} - 1/T_{1,2}$

$$k_2 = k_1 \frac{\xi_1}{\lambda - \xi_1}$$

Water exchange influence T1-relaxation:
Bloch equations including water exchange terms:

$$\frac{dM_1}{dt} = \frac{M_0 - M_1}{T_{1,1}} - k_1 M_1 + k_2 M_2$$

$$\frac{dM_2}{dt} = \frac{M_0 - M_2}{T_{1,2}} - k_1 M_1 - k_2 M_2$$

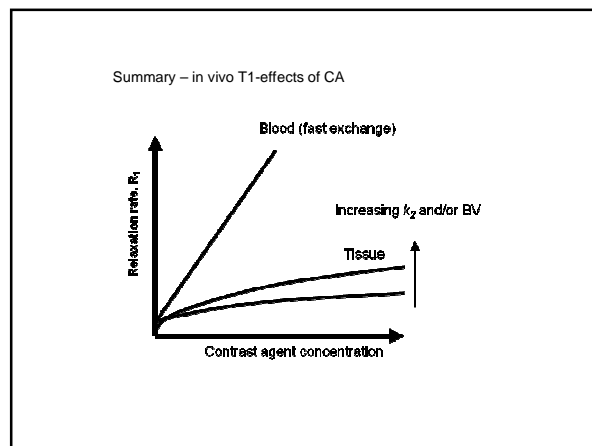
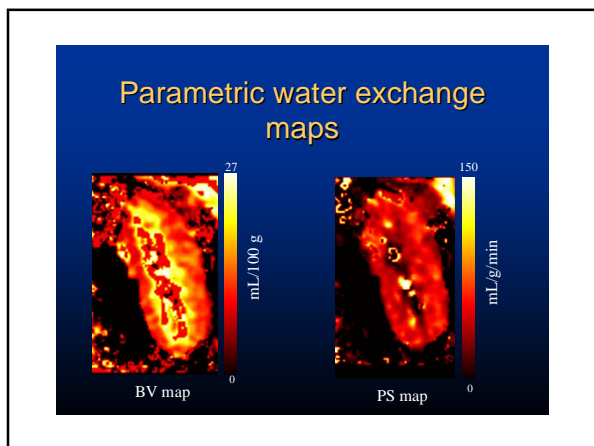
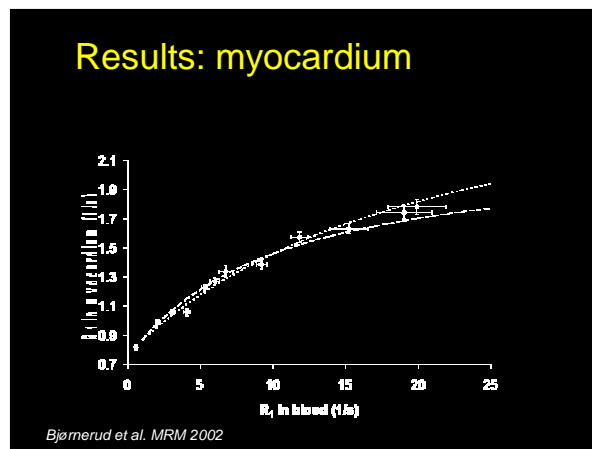
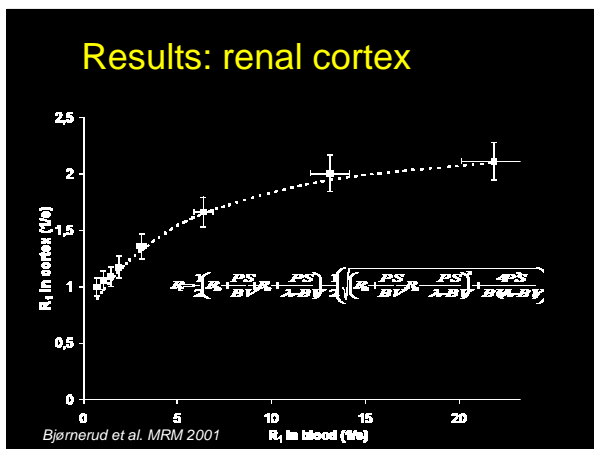
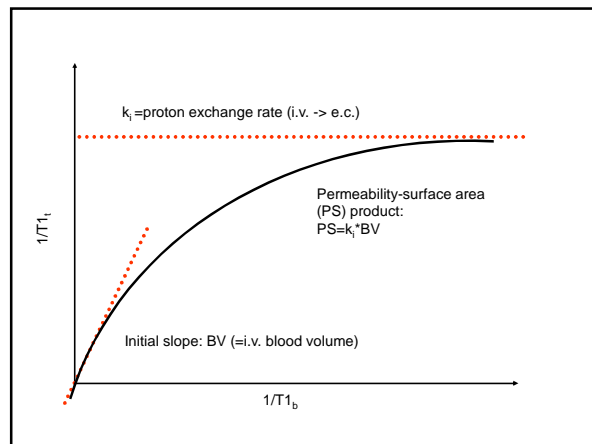
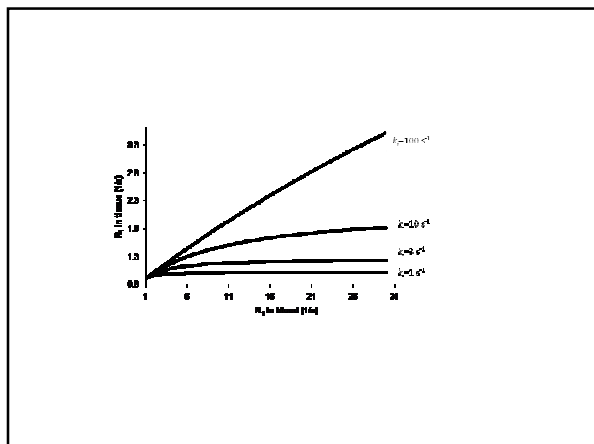
→ $M(t) = M_0 + c_1 \exp(-u_1 t) + c_2 \exp(-u_2 t)$

$$M(t) = M_0 + c_1 \exp(-u_1 t) + c_2 \exp(-u_2 t)$$

Mono-exp approximation:

↓ $M(t) = M_0 (1 - 2 \exp(-R_1 t))$

$$R_1 \approx u_2 = \frac{1}{2} (R_{1,1} + k_1 + R_{1,2} + k_2) - \frac{1}{2} \sqrt{(R_{1,1} + k_1 - R_{1,2} - k_2)^2 + 4k_1 k_2}$$



Susceptibility induced relaxation:

$$M_z = \chi H_0 \quad (H = B_0 / \mu_0)$$

Langevin equation:

$$M = Nm \left[\coth \left(\frac{\mu_0 m H_0}{k_B T} \right) - \left(\frac{k_B T}{\mu_0 m H_0} \right) \right]$$

For paramagnetic agents, Curie law approximation valid < 50 T ...

$$M = \frac{Nm^2 \mu_0 H_0}{3k_B T} = \chi H_0$$

\rightarrow

$$\chi = \frac{Nm^2 \mu_0}{3k_B T}$$

Susceptibility constant

Background & Theory

Iron oxides have much larger magnetic moment than gadolinium and are therefore much more potent relaxation enhancers

Magnetic Field (Tesla)	Iron Oxide Nanoparticle (mu_B)	gadolinium (mu_B)
0	0	0
0.2	2.5	0.1
0.4	3.5	0.2
0.6	4.0	0.3
0.8	4.3	0.4
1.0	4.5	0.5

Background & Theory

Microscopic field inhomogeneities due to intravascular CA

Diamagnetic blood + CA
 $M_i \gg M_c$

Susceptibility induced relaxation

$$S(TE, \omega) = S_0 \int_{-\infty}^{\infty} P(\omega) \exp(-j\omega TE) d\omega$$

P(omega) Gaussian or Lorentzian

Lorentzian lineshape

$$P(\omega) \propto \frac{\sigma}{\sigma^2 + \omega^2}$$

\downarrow

$$S(TE, \omega) = S_0 \int_{-\infty}^{\infty} \left[\frac{\sigma}{\sigma^2 + \omega^2} \right] \exp(-j\omega TE) d\omega$$

$$= S_0 \exp(-\sigma TE)$$

'Effective 1/T2*' is proportional to linewidth of spectrum => monoexponential signal decay

Gaussian lineshape

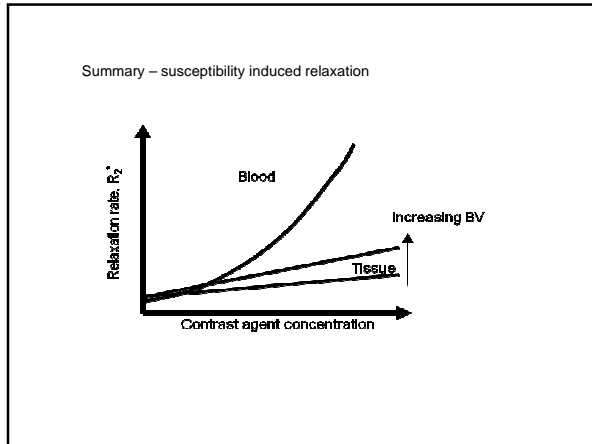
$$P(\omega) \propto \exp\left(\frac{-\omega^2}{2\sigma^2}\right)$$

\downarrow

$$S(TE, \omega) = S_0 \int_{-\infty}^{\infty} \exp\left(\frac{-\omega^2}{2\sigma^2}\right) \exp(-j\omega TE) d\omega$$

$$= S_0 \exp[-(\sigma TE)^2]$$

'Effective 1/T2*' is proportional to $\sigma^2 TE$: i.e. measured T2* is function of TE.



Advanced Applications of MR Contrast Agents

General requirement: estimate contrast agent concentration in vivo (at least to within a scaling constant)

T_1 -based dynamic imaging

Most accurate approach: quantification of $1/T_1$

$$\Delta R_1(t) = R_1(t) - R_1^0 = C(t) \rho_1$$

Then, assuming fast water exchange: $C(t) = k \Delta R_1(t)$

Estimation of $C(t)$ from T1-GRE sequence:

Requirement: $TR \ll T_1$ and $TE \ll T_2^*$ then:

$$SI \propto \frac{M_0 TR}{T_1}$$

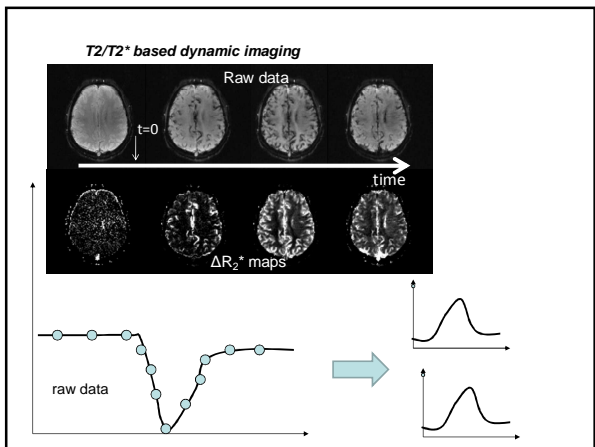
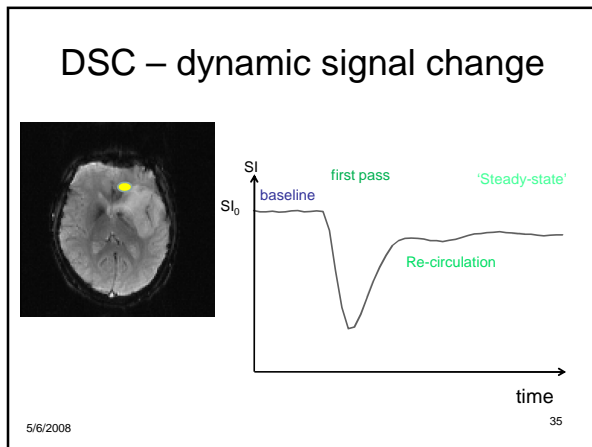
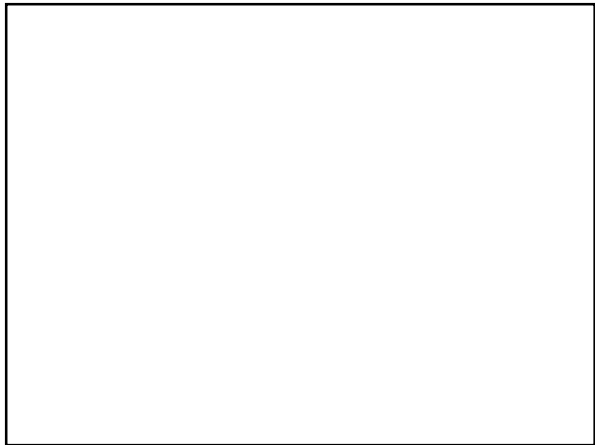
T_2/T_2^* based dynamic imaging

Assumption: monoexponential T_2/T_2^* signal decay (Lorentzian lineshape)

Then:

$$SI(t) = SI(0) f(M_0, T_1) \exp(-TE \cdot \Delta R_2(t))$$

If T1-effects are negligible then:

$$\Delta R_2(t) = k \ln \left(\frac{SI(t)}{SI(0)} \right) / TE \propto C(t)$$


Perfusion imaging

Central volume principle:

$$V_t = F_a \cdot MTT = \frac{F_a}{q_{in}} \int_0^{\infty} C_t(\tau) d\tau$$

Blood volume maps in tumor imaging:

$$\Delta R_2'(t) = k \cdot C(t) = -\frac{k}{TE} \log\left(\frac{S(t)}{S_0}\right) \quad rCBV \propto \int \Delta R_2(t) dt$$

$$V_b = k \frac{\int_0^{\infty} C_t(\tau) d\tau}{\int_0^{\infty} C_a(\tau) d\tau} \propto \frac{\int \Delta R_{2t}(\tau) d\tau}{\int \Delta R_{2a}(\tau) d\tau}$$

Measurement of flow (perfusion) and MTT:

Introducing the *tissue residue function* $R(t)$

$$C_t(t) = F_a \int_0^t R(t-\tau) C_a(\tau) d\tau = F_a R(t) \otimes C_a(t)$$

Perfusjons-MR

arterie

kapillærnett

vene

Vevets residualfunksjon

Standard dekonvolusjonsproblem

$$\kappa C(t) = F \int_0^t C_a(\tau) R(t-\tau) d\tau$$

MR perfusjonsavbilding

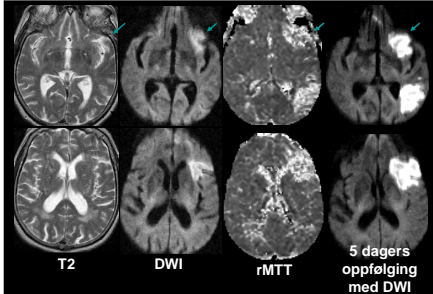
MTT = V/F

BV

BF

MTT

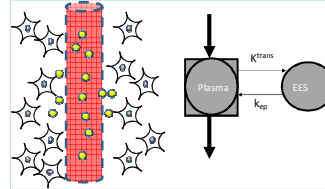
Slag: Forlenget MTT = area at risk



Østergaard et al, Århus, Danmark

Dynamic contrast enhanced imaging

Two-compartment model:



K^{trans} and K^{ep} related to 'leakiness of capillary wall'

Dynamic contrast enhanced imaging

Two-compartment model:

$$\frac{dC_t}{dt} = K^{trans} C_p(t) - k^{ep} C_t(t)$$



$$C_t(t) = K^{trans} \int_0^t C_p(\tau) \exp[-k^{ep}(t-\tau)] d\tau$$

Transfer constants can be determined if $C(t)$ can be measured in tissue and artery (AIF)

Dynamic contrast enhanced imaging

If not, need to assume $C_p(t)$ to be monoexponential

$$\frac{dC_t}{dt} = K^{trans} C_{p,0} \exp\left(-\frac{t}{T_1}\right) - k^{ep} C_t(t)$$



$$C_t(t) = \frac{K^{trans} C_{p,0}}{T_{1/2} - k^{ep}} \left[\exp(-k^{ep}t) - \exp\left(\frac{-t}{T_{1/2}}\right) \right]$$

Dynamic contrast enhanced imaging

