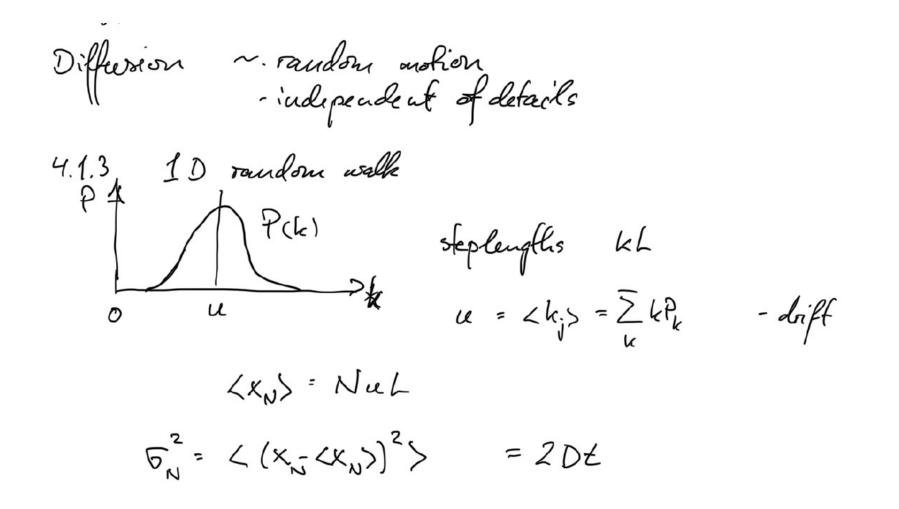
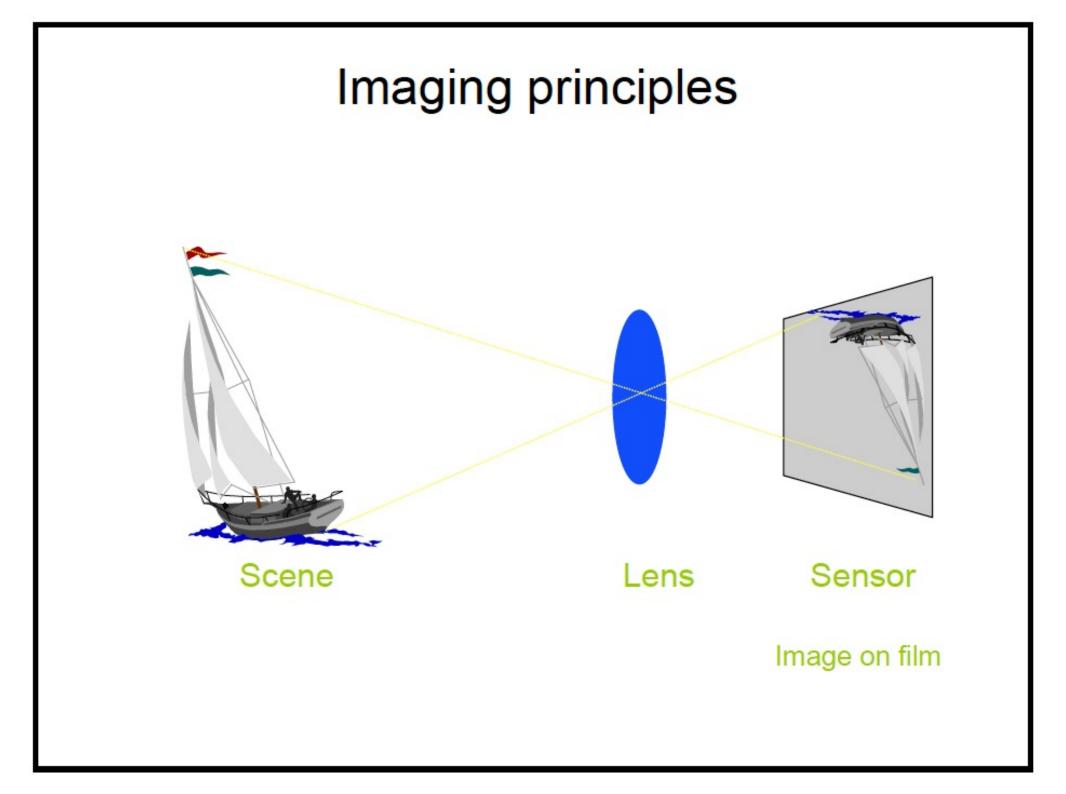
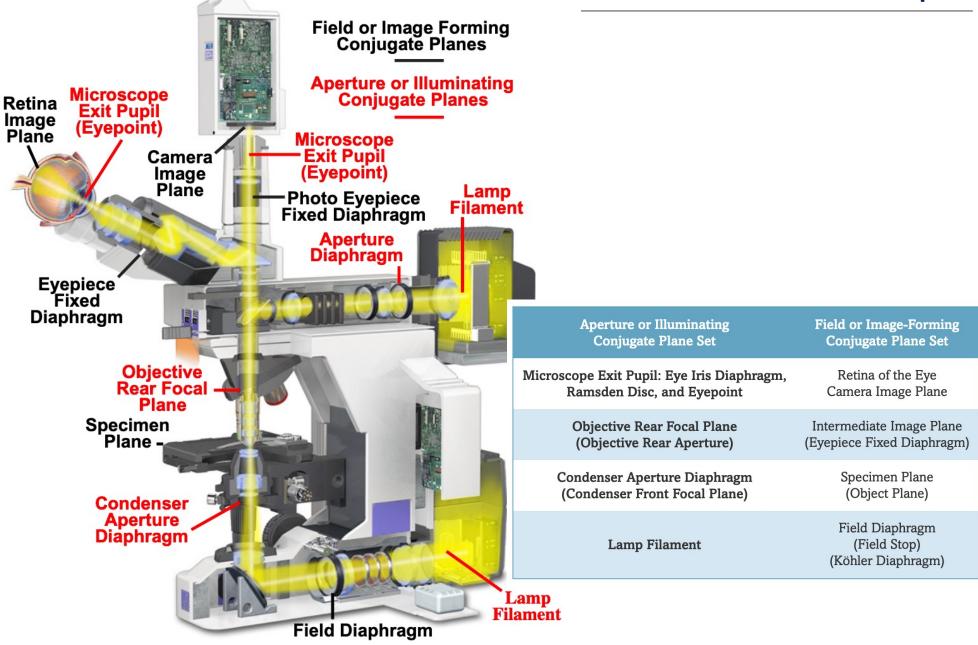
- Did you do 2D RW?
- Demo 1drw?



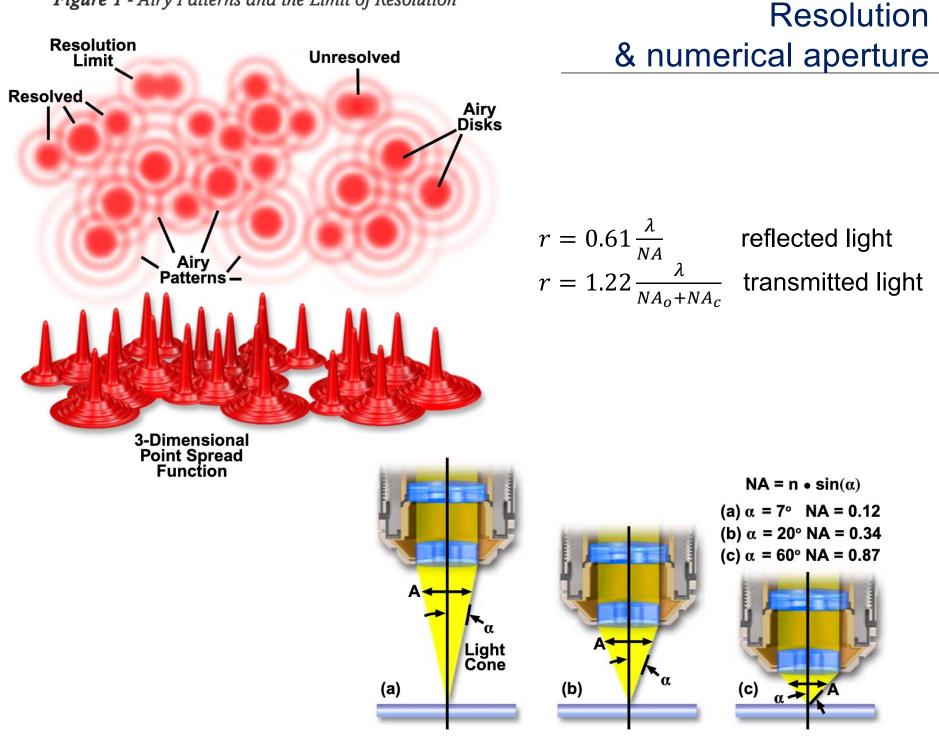
- Microscope imaging and cameras
- What is a digital image?
- Image types and resolution
- Why do we need image analysis?
- How to do image analysis (basic steps)?
- Morphological operators
- Watershed algorithm
- Examples



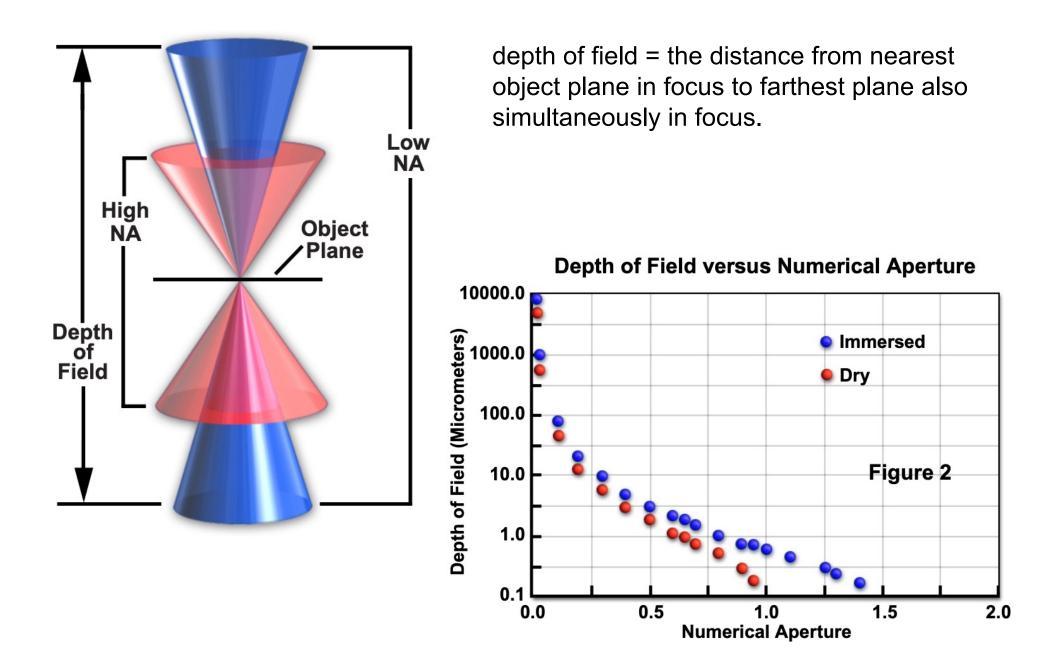


Microscope

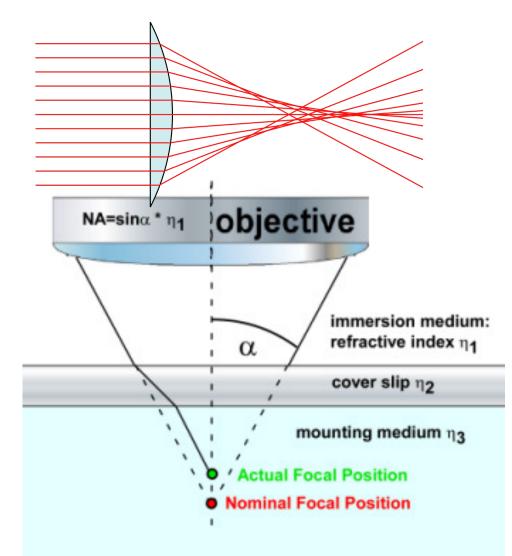
Figure 1 - Airy Patterns and the Limit of Resolution

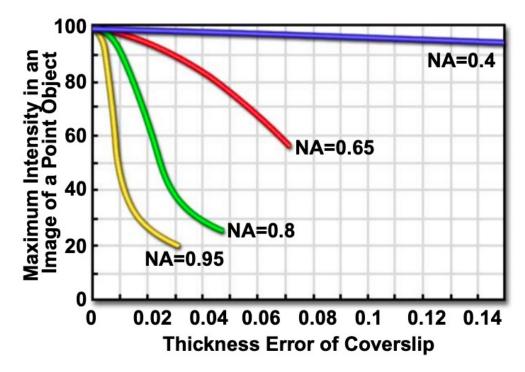


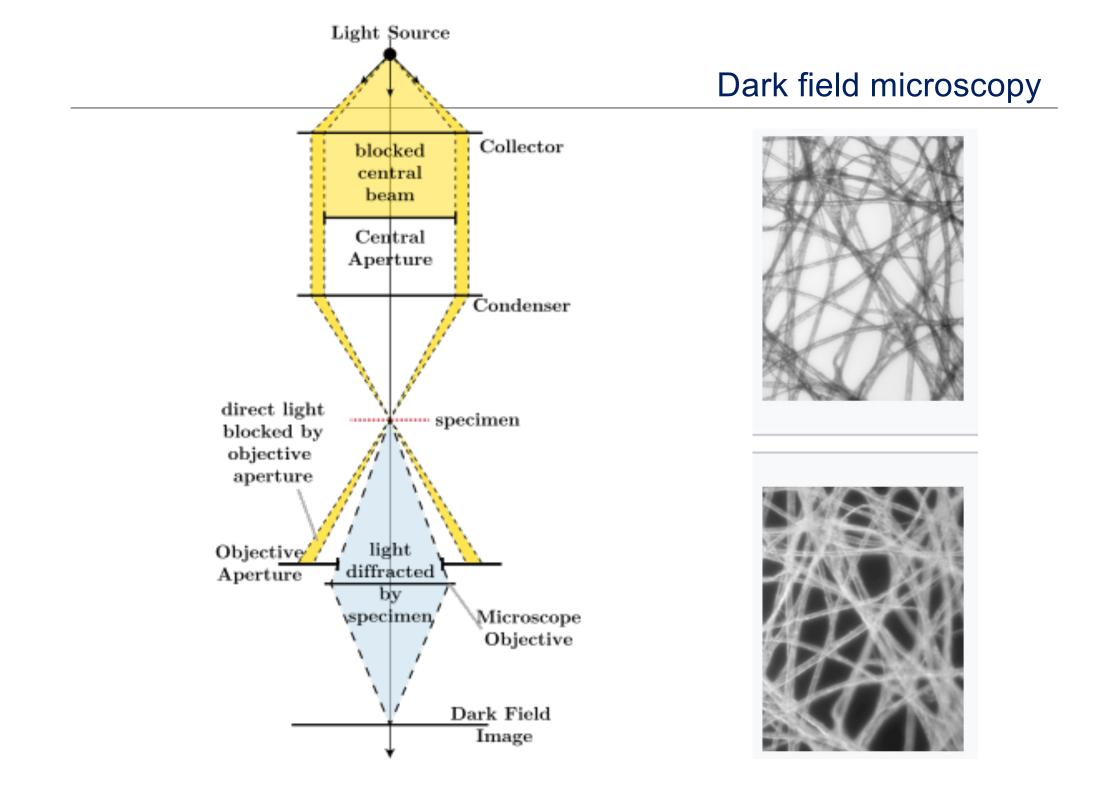
Depth of field/focus



Spherical aberration & coverslip correction







making phase changes visible in phase-contrast microscopy is

- to separate the illuminating (background) light from the specimen-scattered light (which makes up the foreground details) and
- to manipulate these differently.

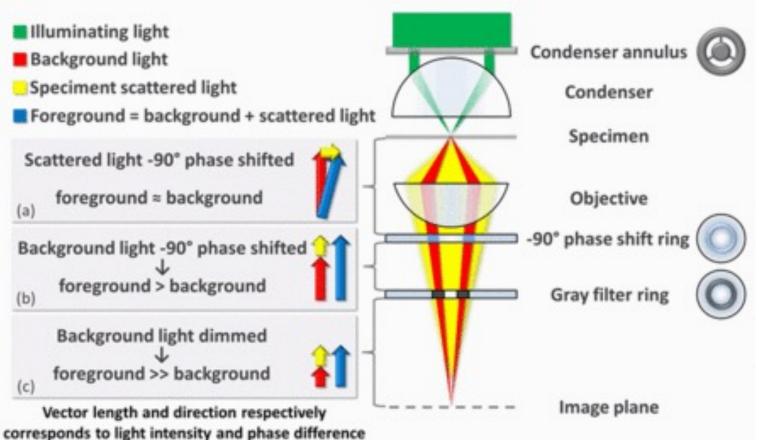
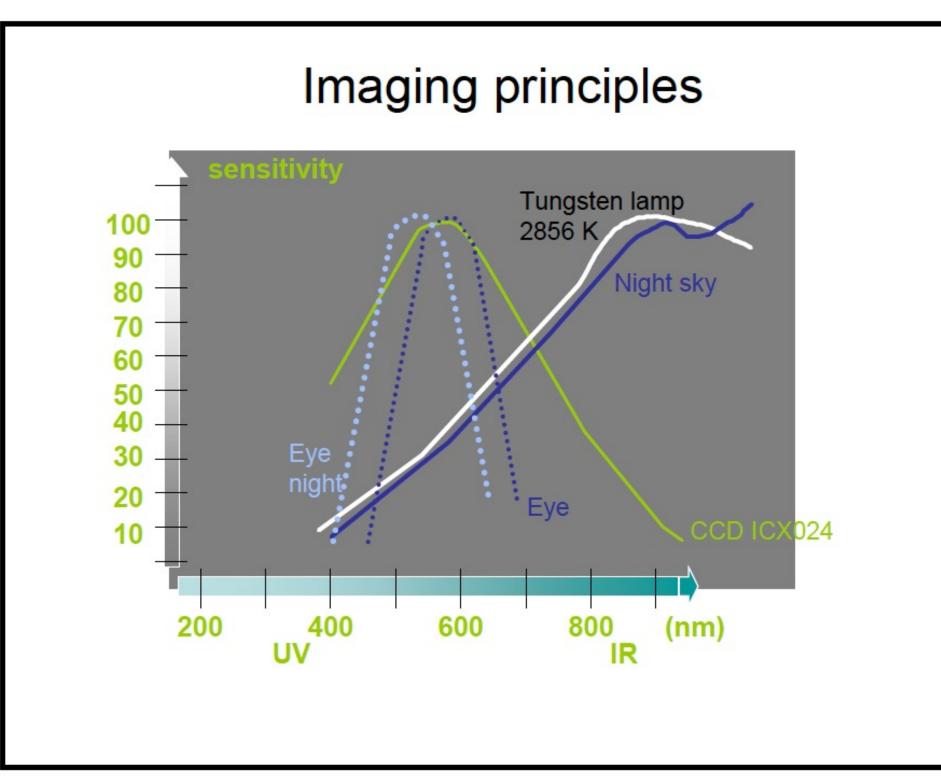
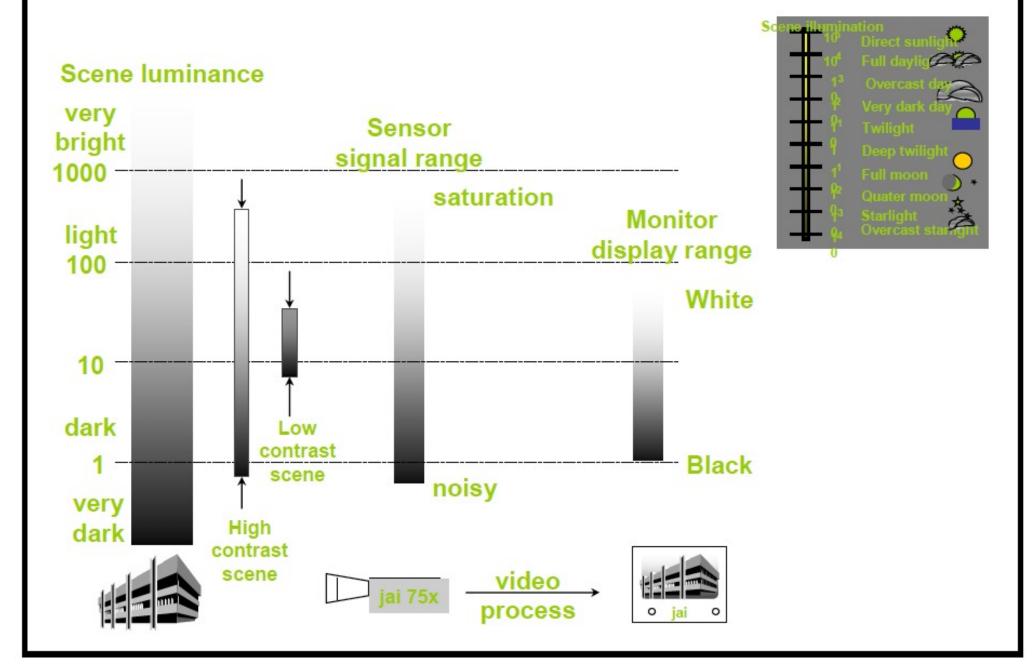


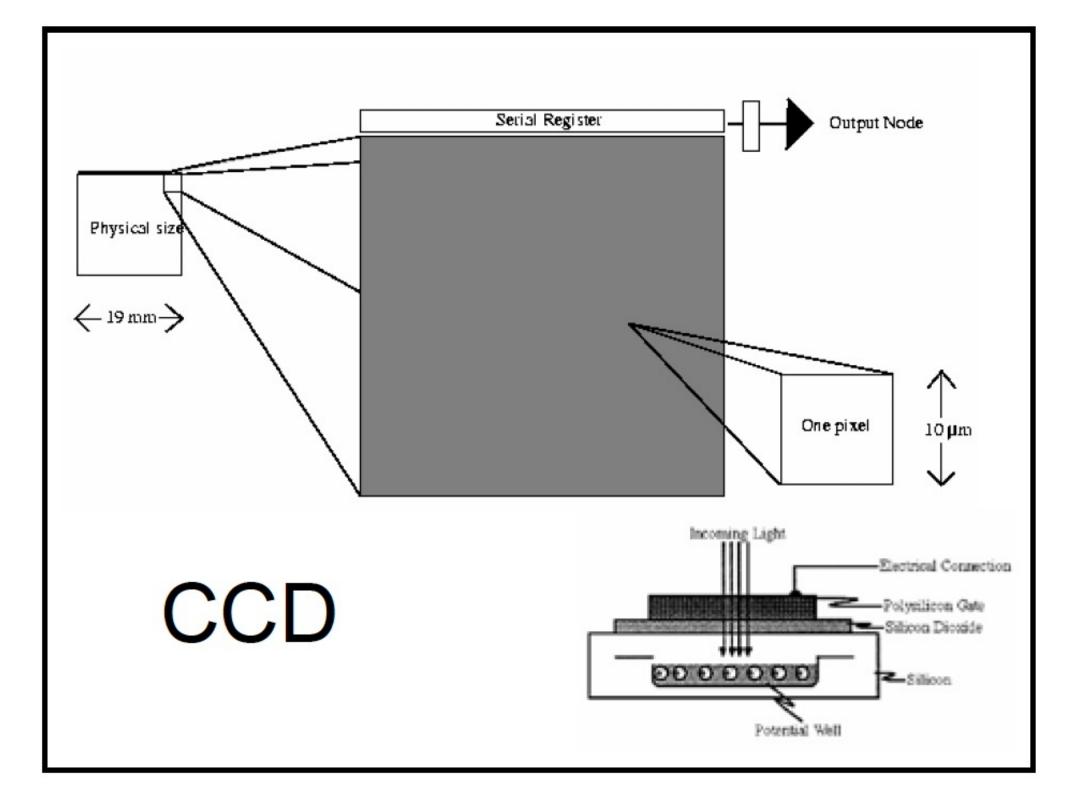
image contrast is increased in two ways:

- by generating constructive interference between scattered and background light
- by reducing the amount of background light that reaches the image plane



Luminance and contrast





Noise

- Shot noise / thermal / dark current
- Read-out noise
- Saturation / Glare / Blooming
- High energetic "cosmic" rays
- "Digital noise" / Moirè patterns

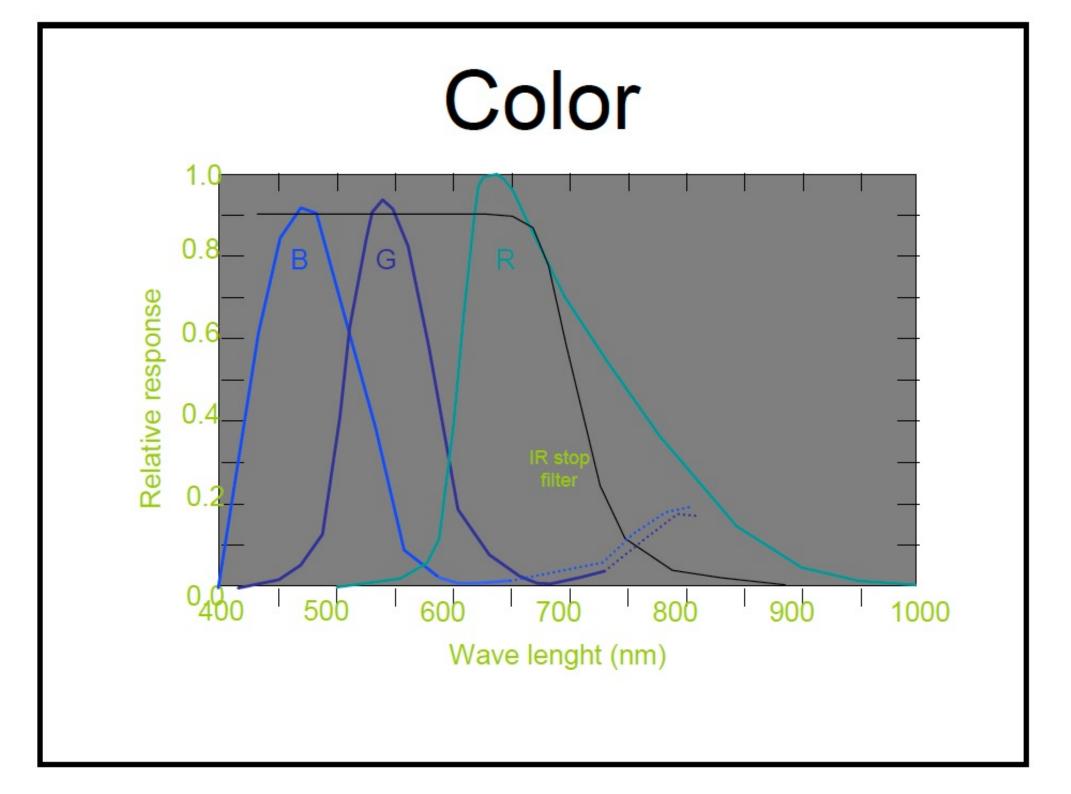
High Resolution Digital Cameras Advantages Drawbacks

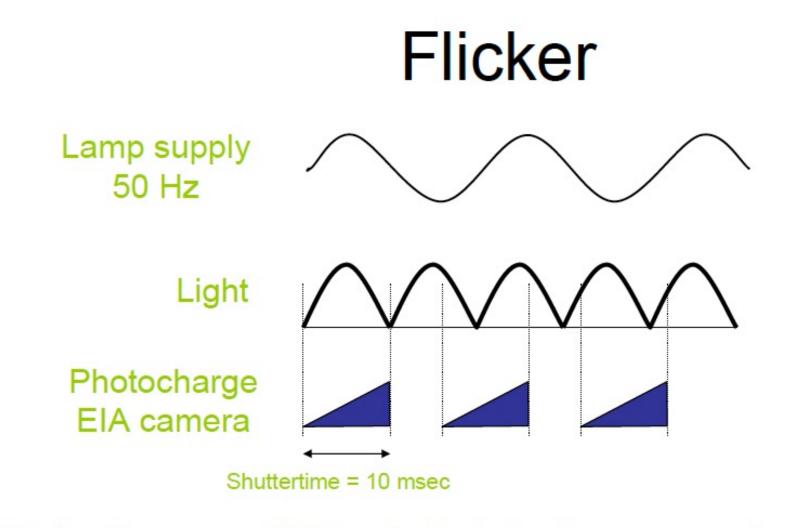
- Light sensitive
- High spatial and <u>dynamical</u> resolution
- Low noise

- Slow data transfer
- Produces much data
- Requires custom made software
- Not user friendly
- Expensive

MORE SENSITIVE THAN THE EYE

16 bit: cooled sensor





Shutter time = one light period, photocharge = constant

Result = no flicker and reduced sensitivity

Practical tips for adjusting video camera

- Turn off automatic adjustments
- Turn down Gain (it only adds noise)
- Adjust light intensity and shutter speed
 - until histogram covers intensity range
 - shutter speed must be short enough for desired frame rate
 - shutter speed long enough to avoid flicker

What is an image?



f = f(x, y)

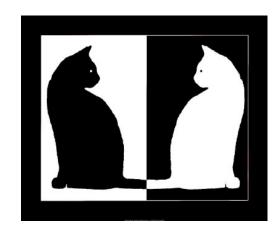
12	0	234	122	54	65	
78	34	215	23	23	34	
109	65	30	117	54	54	
140	23	111	214	65	76	
11	12	245	213	235	189	
155	0	78	0	0	67	
178	198	201	0	12	42	

Pixels MxN



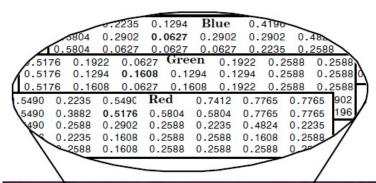
- Intensity images grey level
- Binary images black and write
- RGB images color images







Color image



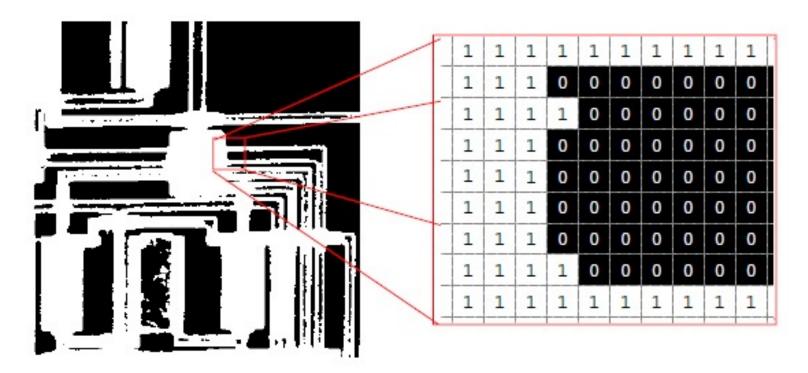


[RGB] Red Green Blue M x N x 3

Matlab image processing toolbox:

im = imread('landscape.jpg');
figure(1),imshow(im)
whos im
imfinfo('landscape.jpg')
A = im(1000:1010,1000:1010,:);

Binary image



im_bw = imread('black_and_white_cats-1541.jpg'); im_bw = rgb2gray(im_bw); im_bw = im2bw(im_bw); imwrite(im_bw,'bw_cats.png'); figure,imshow(im_bw) whos im_bw unique(im_bw)

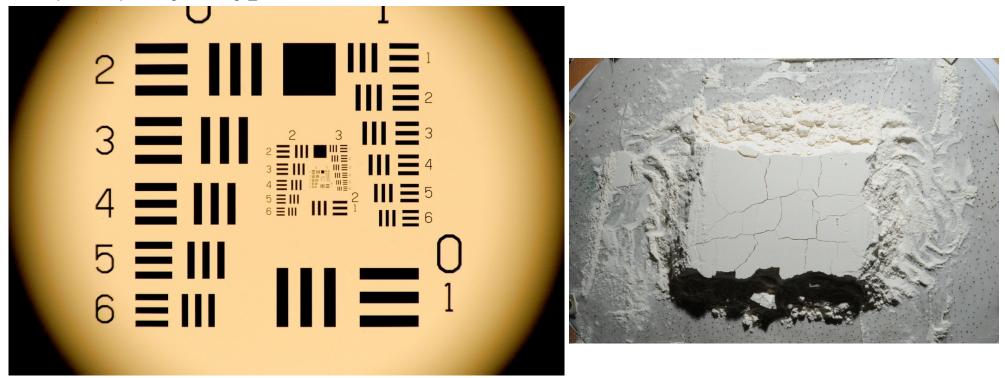
Image quality:

- Number of pixels in the matrix image size
- Intensity range

1 bit depth (2¹ = 2) - black and white
8 bit depth (2⁸ = 256) - gray scale 0..256
12, 16 bit gray scale
24 bit depth (256 shades of RGB) - true color

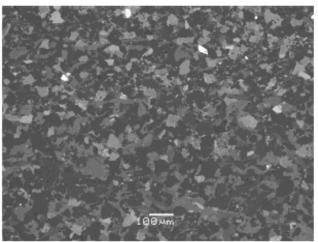
Spatial resolution of images

http://en.wikipedia.org/wiki/Image_resolution



Spatial resolution of optical system - Number of independent pixels per unit length

Morphological analysis – a mathematical tool to investigate geometrical structure of binary or grayscale image



Segmentation

procedure

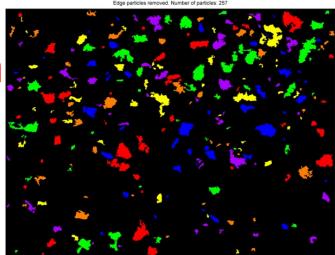


Image segmentation quick steps:

- $\mathsf{RGB} \to \mathsf{gray}$
- Filter
- Thresholding \rightarrow binary
- Labeling connected components
- Geometrical analysis of connected components

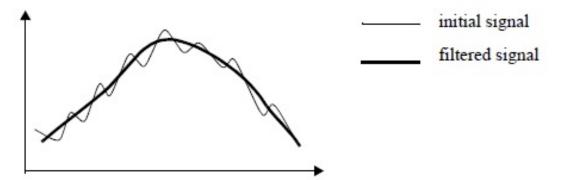
RGB to gray scale

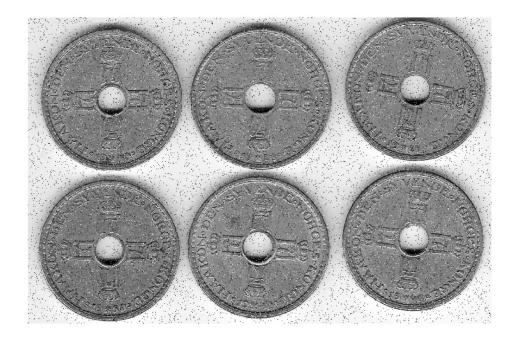
```
im_bw = rgb2gray(im_bw);
Im_bw = im(:,:,1);
Im_bw = (im(:,:,1) + im(:,:,2) + im(:,:,3))/3;
```



Noise removal

- Filtering smoothing
- Background correction







Convert to black and white

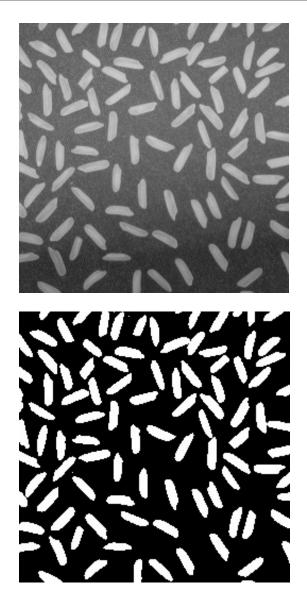
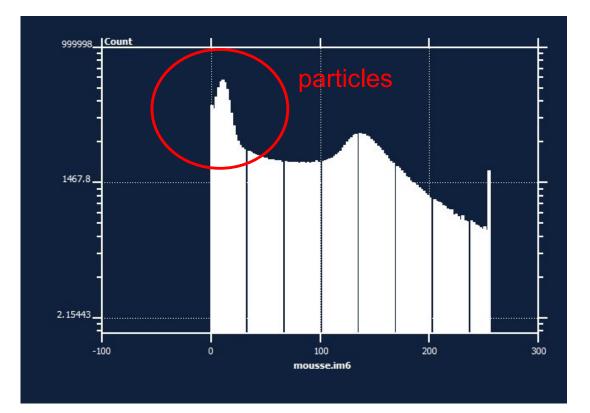


Image histogram



Thresholding intensity interval (a,b)

Labeling connected components

0	0	0	140	140	140	140	140	140	140
o	o	o	o	140	140	140	140	140	140
o	o	0	o	0	140	140	140	140	140
o	0	0	0	0	140	140	0	0	о
o	o	o	o	0	o	0	0	0	0
o	o	o	o	0	0	0	0	0	o
0	o	o	o	o	o	o	o	O	0

4 or 8 neighbor connectivity

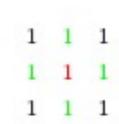
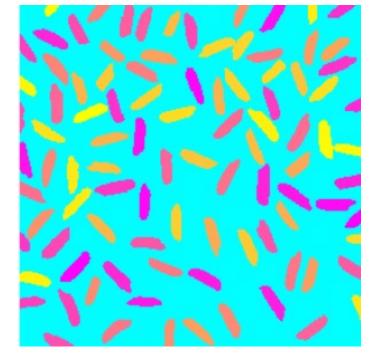
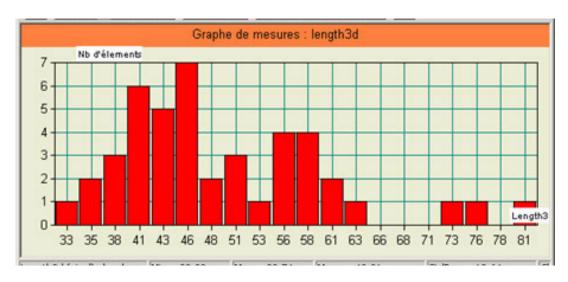


Figure B.9: 4-connectivity of pixels in a 3x3 pixel-environment. The center pixel (1) is connected to its nearest neighbours (1's), but not its next nearest neighbours (1's).

minutes

Particle size distribution





How to make segmentation

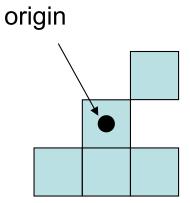


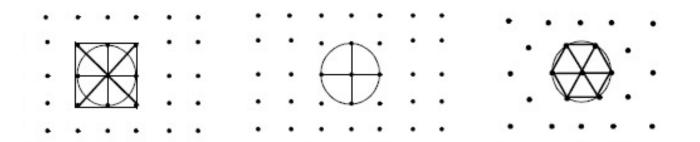
?

Morphological transformation are based on a structural element



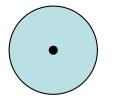
- shape
- center location





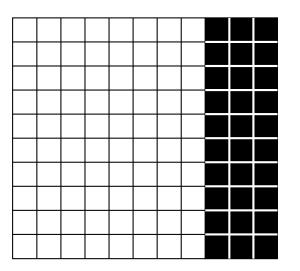
Erosion and dilation – basic operations

Erosion

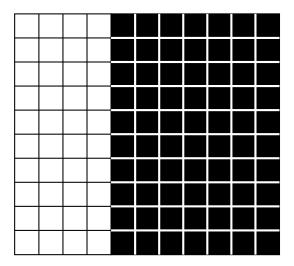


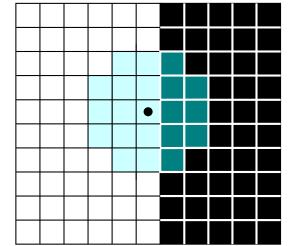
Dilation

"Set the value at the origin to the <u>maximum</u> value of pixels in the structural element"



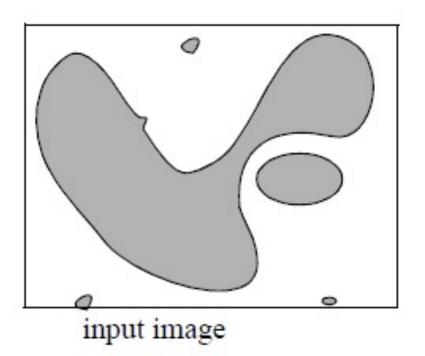
"Set the value at the origin to the <u>minimum</u> value of pixels in the structural element"

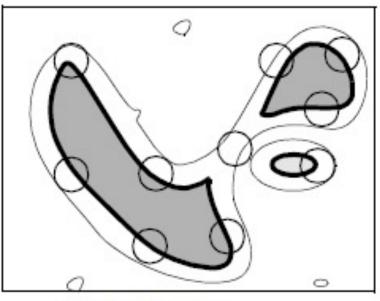






- removes isolated points
- discards peaks on the boundaries
- disconnects some particles

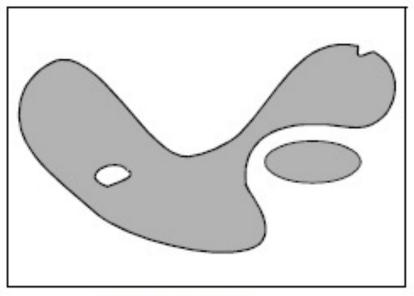




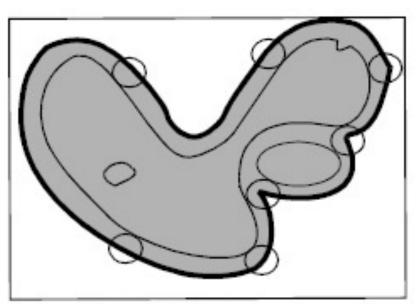
eroded image



- fills small holes inside particles
- enlarges the size of the particles
- connects neighboring objects



input image



dilated image

Opening and closing

Opening = Erosion + Dilation Closing = Dilation + Erosion

Original image \longrightarrow Erosion \longrightarrow Dilation



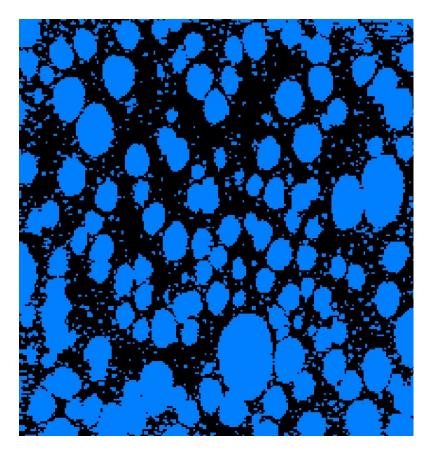


Original image → Dilation → Erosion

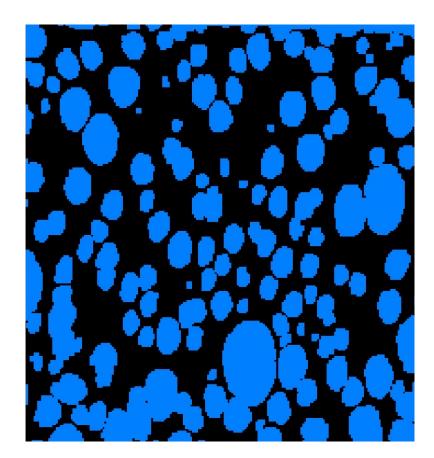


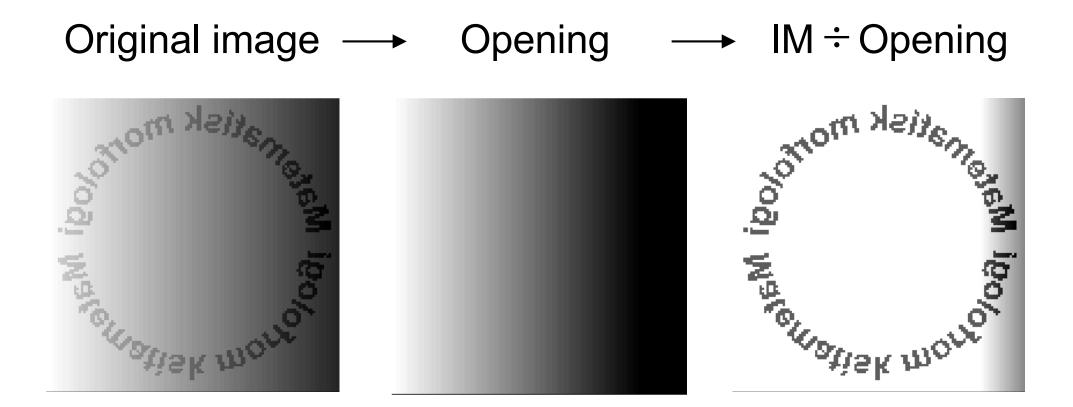


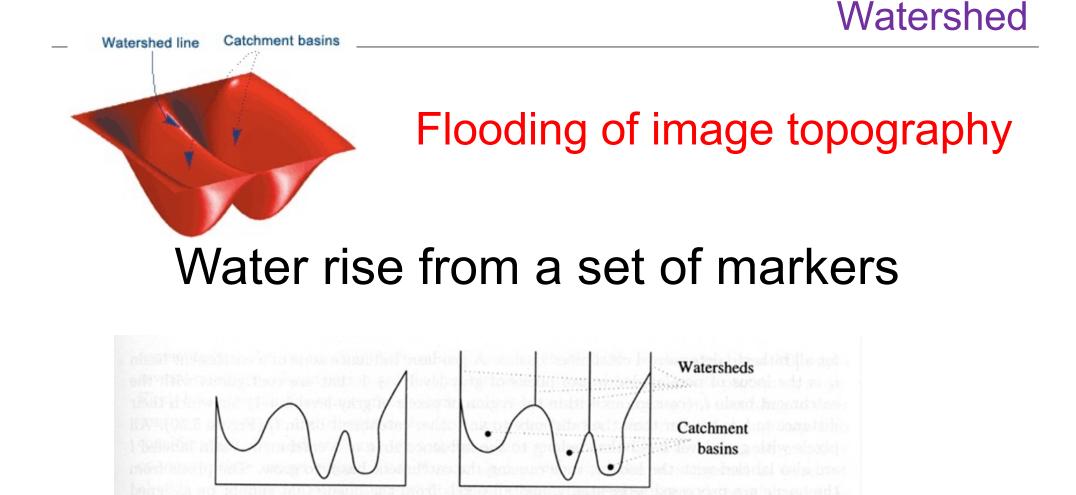
Original image



After opening





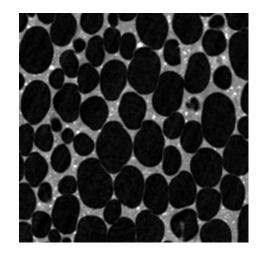


(b)

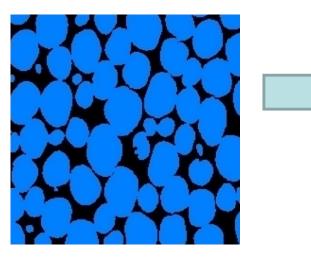
Figure 5.49: One-dimensional example of watershed segmentation: (a) gray-level profile of image data; (b) watershed segmentation—local minima of gray-level (altitude) yield catchment basins, local maxima define the watershed lines.

(a)

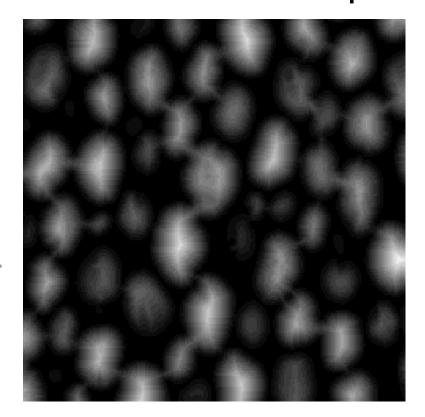
Gray level image



Binary image



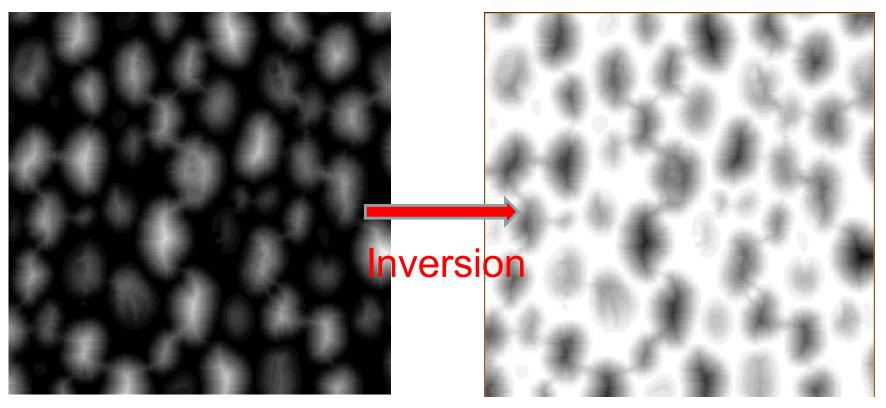
Reconstruction of individual pores in foam Distance map



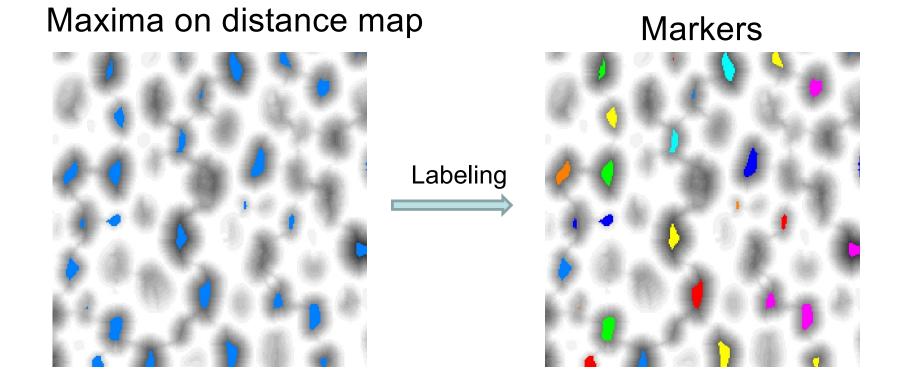
Valleys for watershed

Distance map

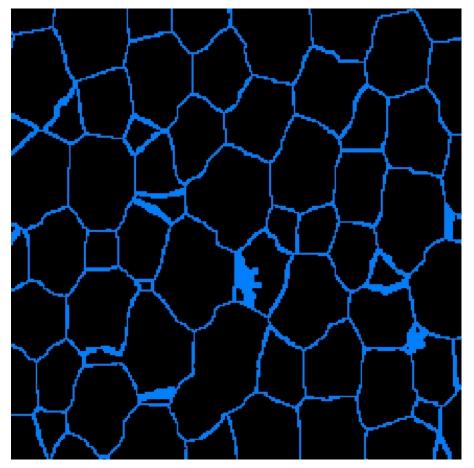
Inversed distance map

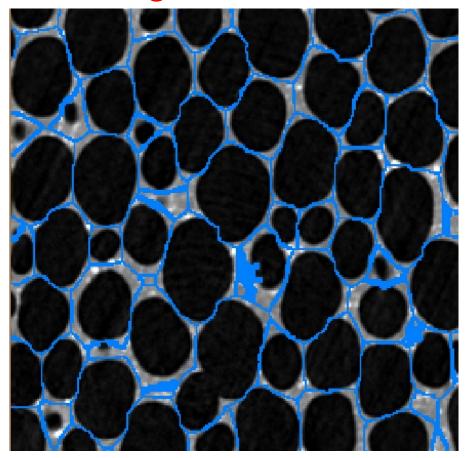


Create markers

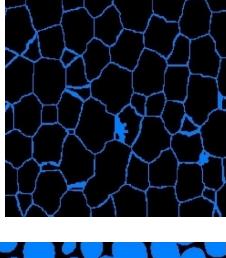


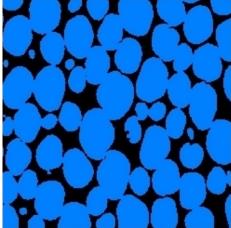
Watershed lines – boundaries between regions

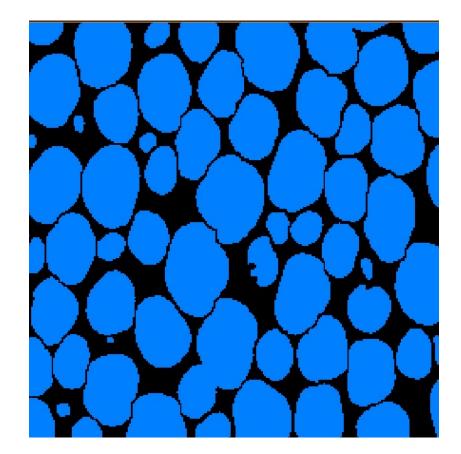




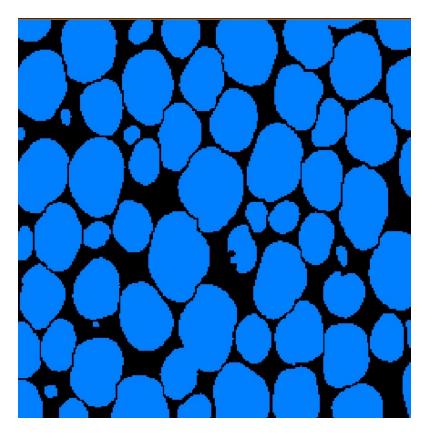
Subtraction of watershed lines gives separated pores

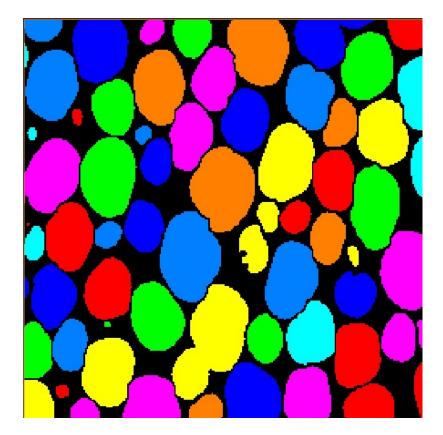






Labeling of connected components





Best tools

- Fiji (imageJ)
- Matalb
- Python

Principle of fluorescence microscopy

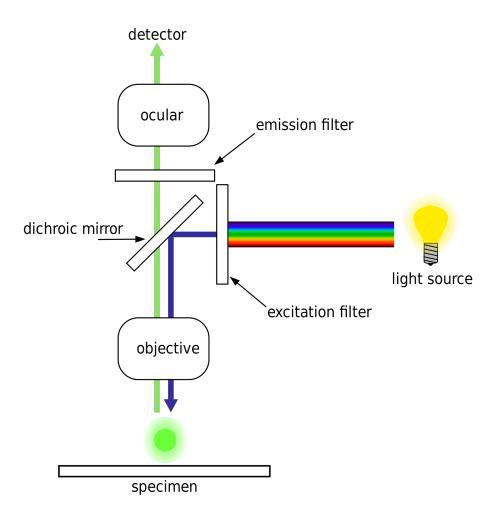
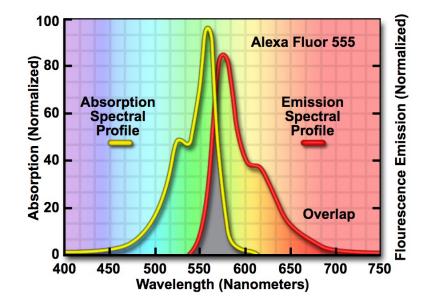
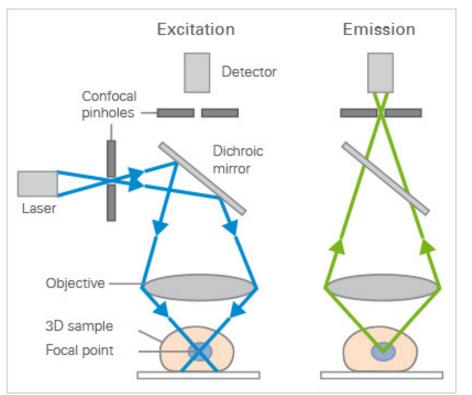


Figure 3 - Fluorophore Absorption and Emission Profiles



Confocal fluorescence microscopy



Confocal laser scanning microscopy

Yokogawa Spinning Disk Unit Optical Configuration Shaped and Collimated — Laser Illumination Monochrome CCD Camera Microlens Array Lens Disc Barrier Filter Emission Light Sweep Pattern Tube Lens Dichromatic Pinhole (Nipkow) Disc Beamsplitter 50 µm Excitation Pinhole and Array Emission Light Objective Figure 8 Specimen

Spinning disk confocal microscopy

Lower resolution, faster

High resolution, slow

Multichannel

