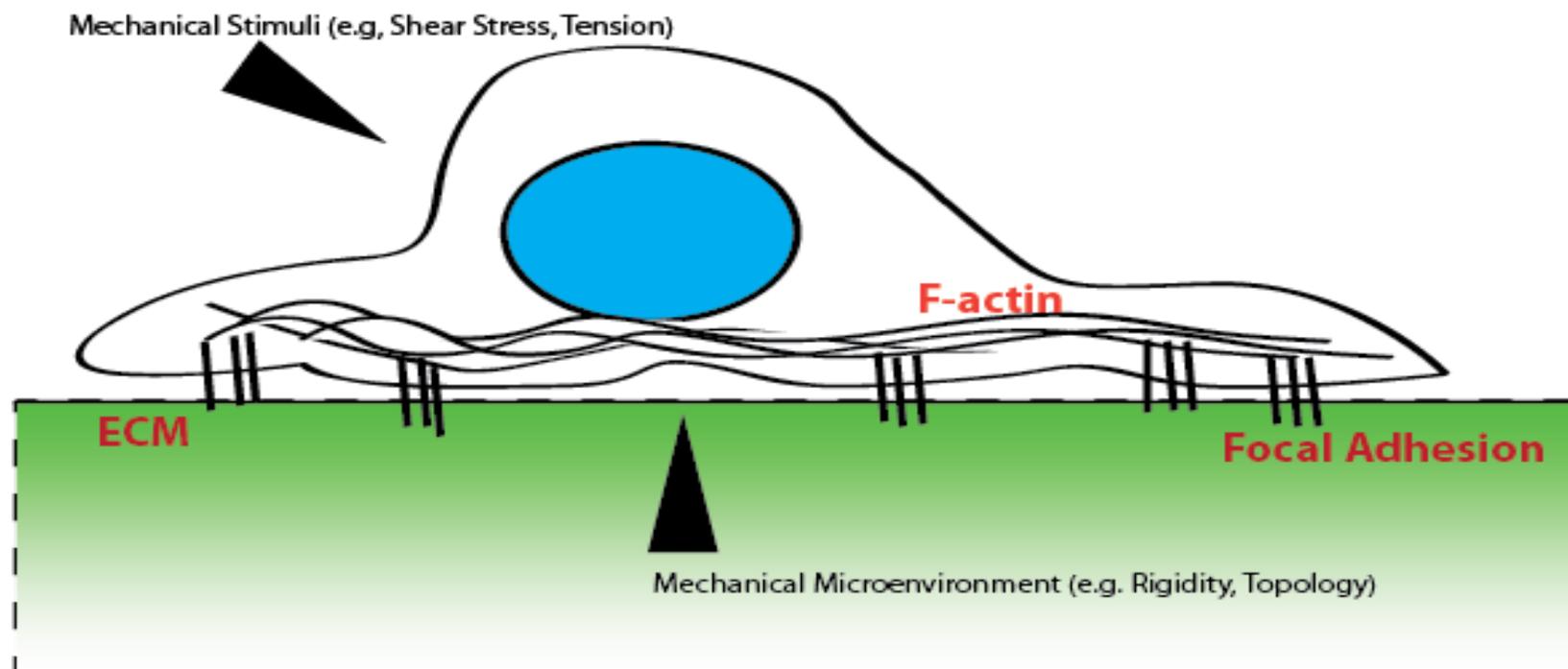
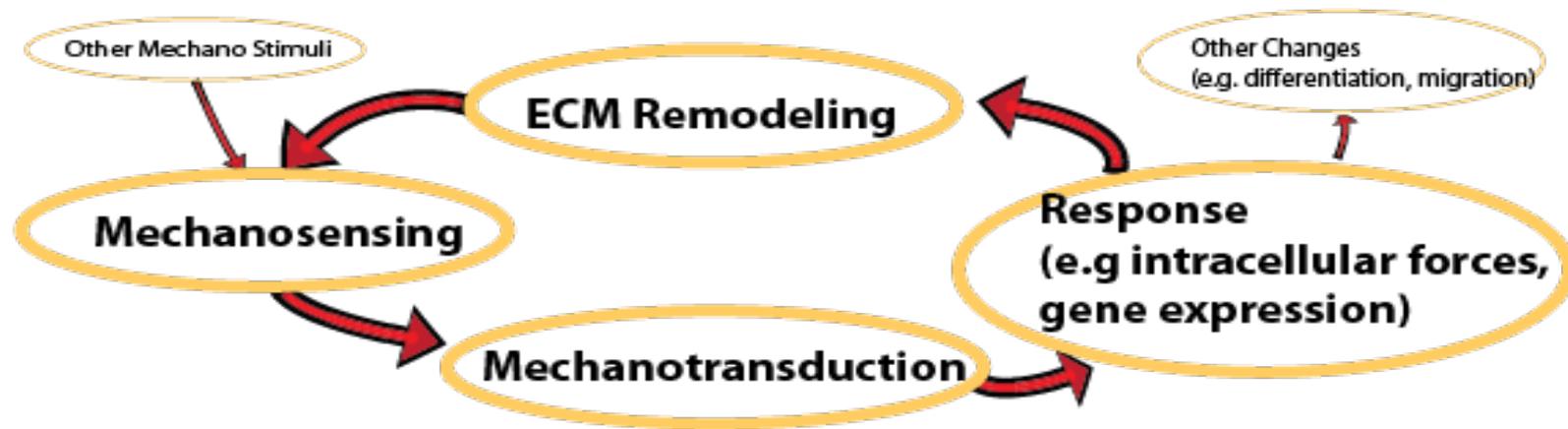


# Biological Applications of MechanoBiology

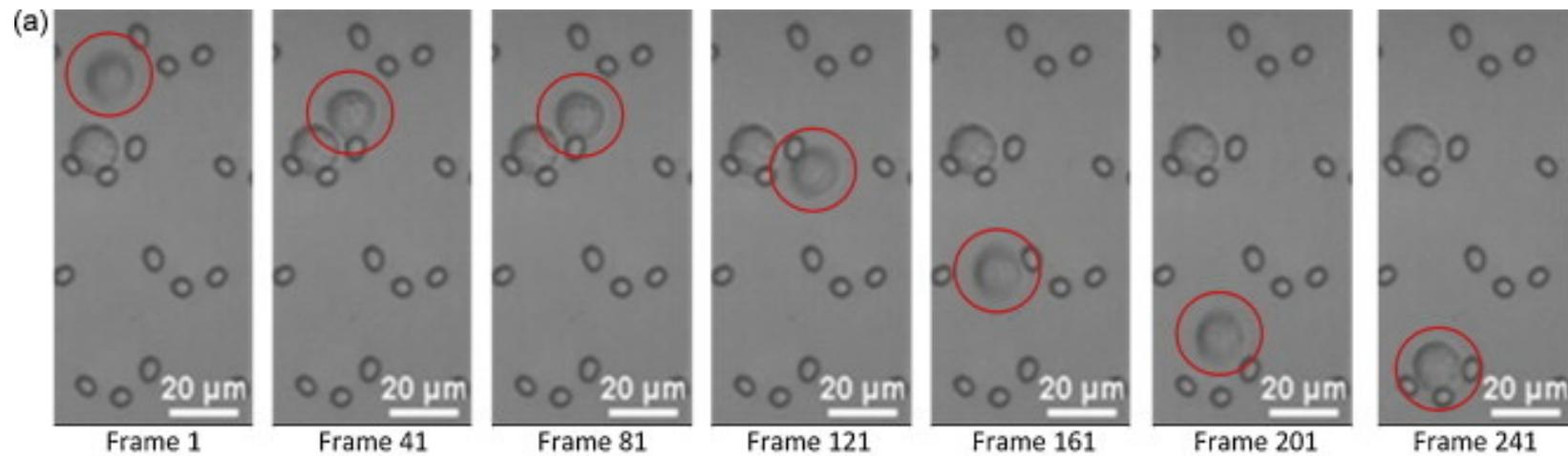
FYS4715 Høst 2021

Xian Hu(Edna)



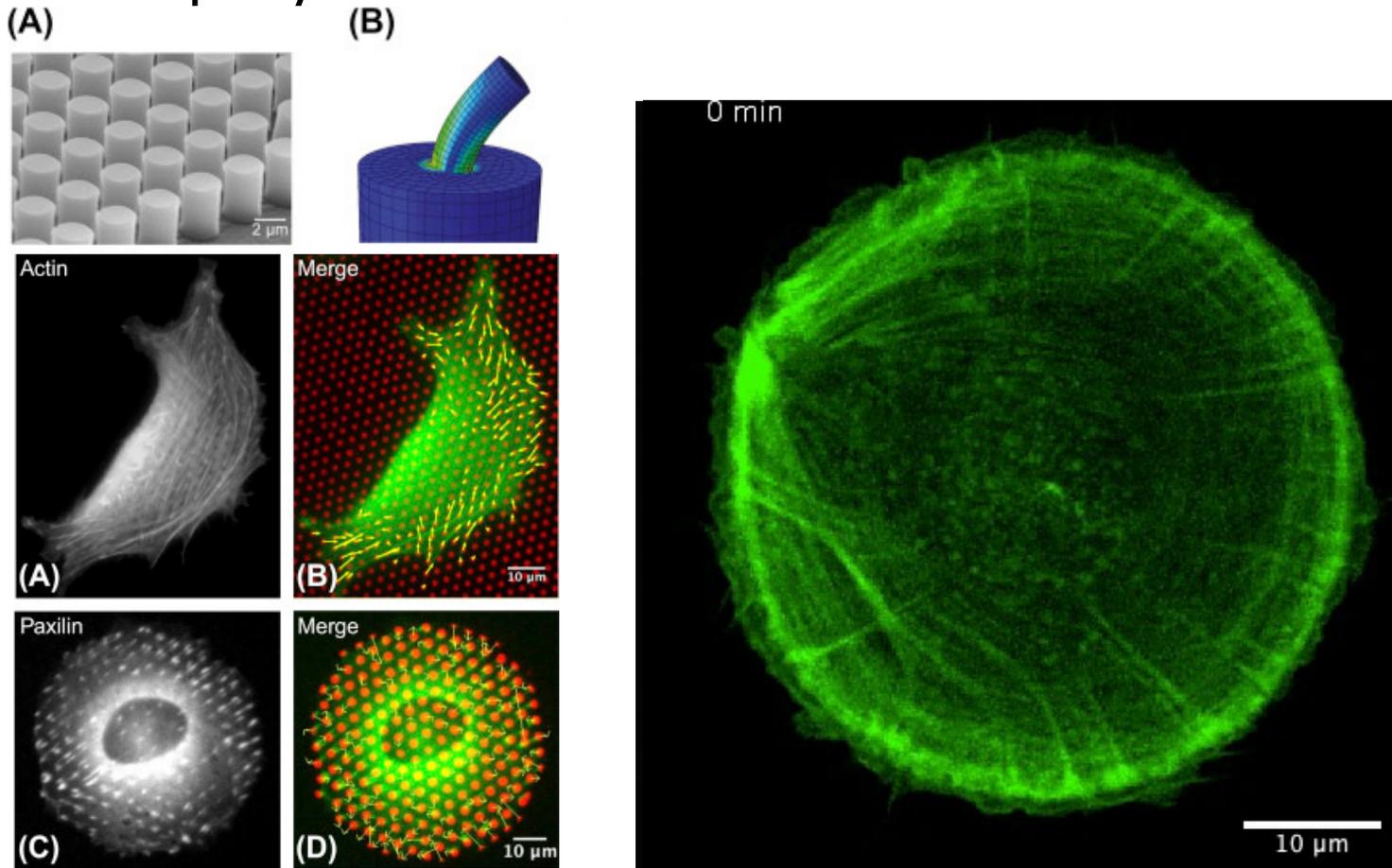
# Cellular Rigidity

Microfluidics device that capture CTC by rigidity



Tan et al, 2010 Biosensors and Bioelectronics

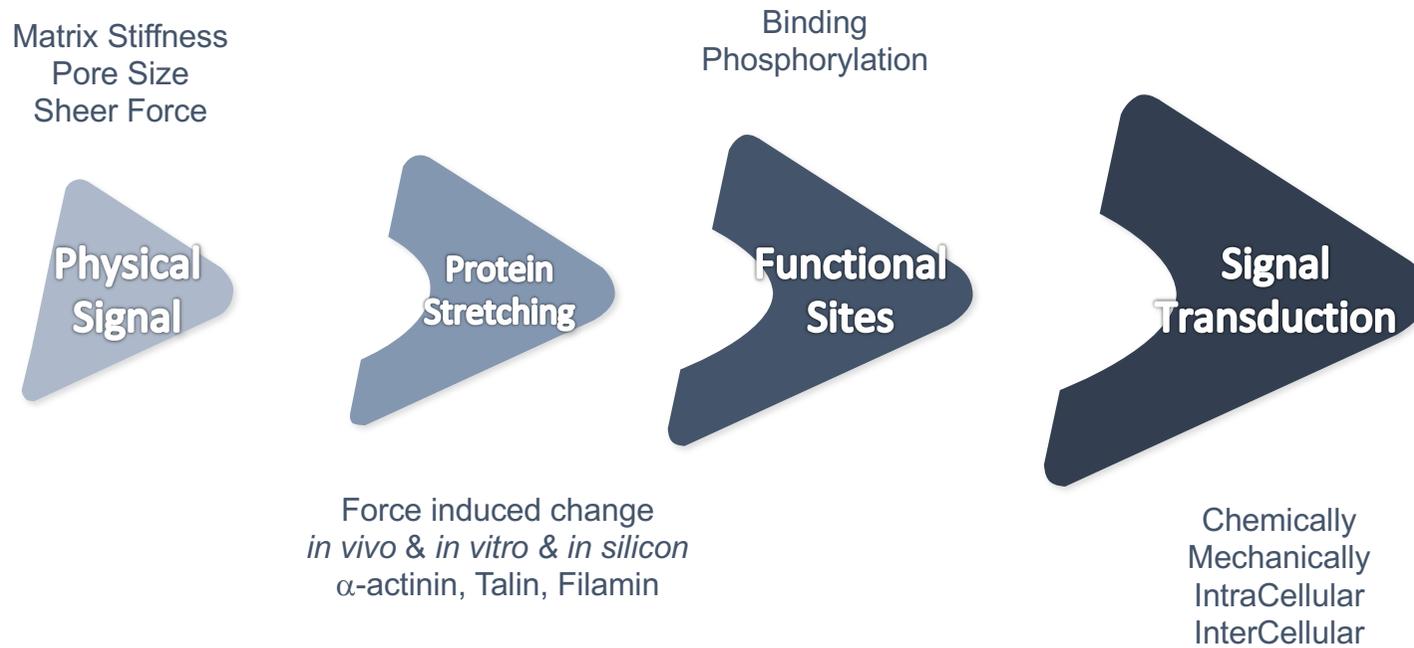
# Cell exert physical forces on substrate



Gupta et al, 2015 Methods in Cell Biology

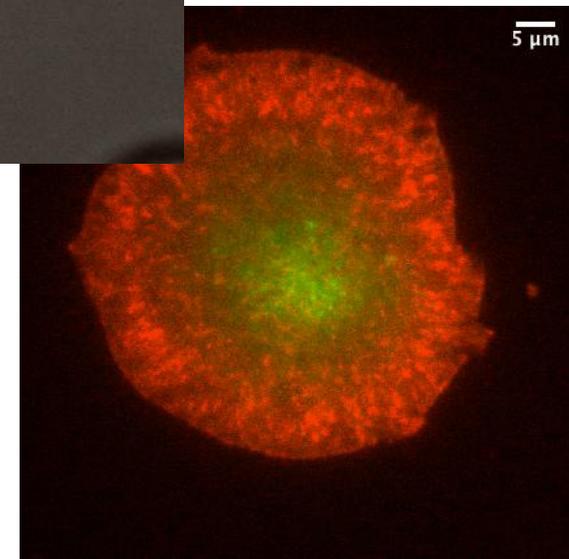
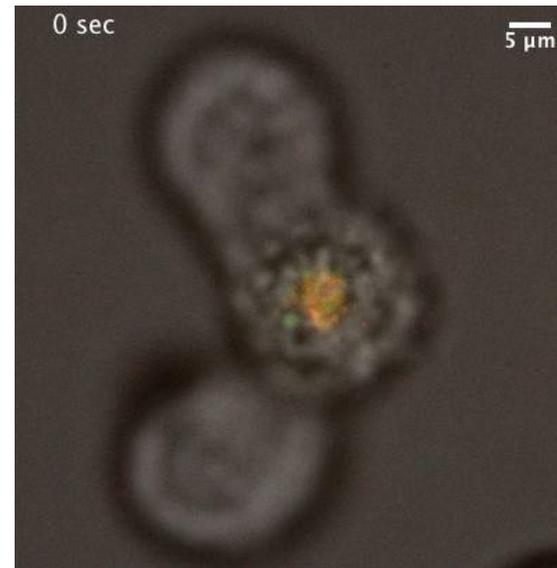
Tee et al, 2015 NCB

# Rigidity Sensing and Molecular Mechanism

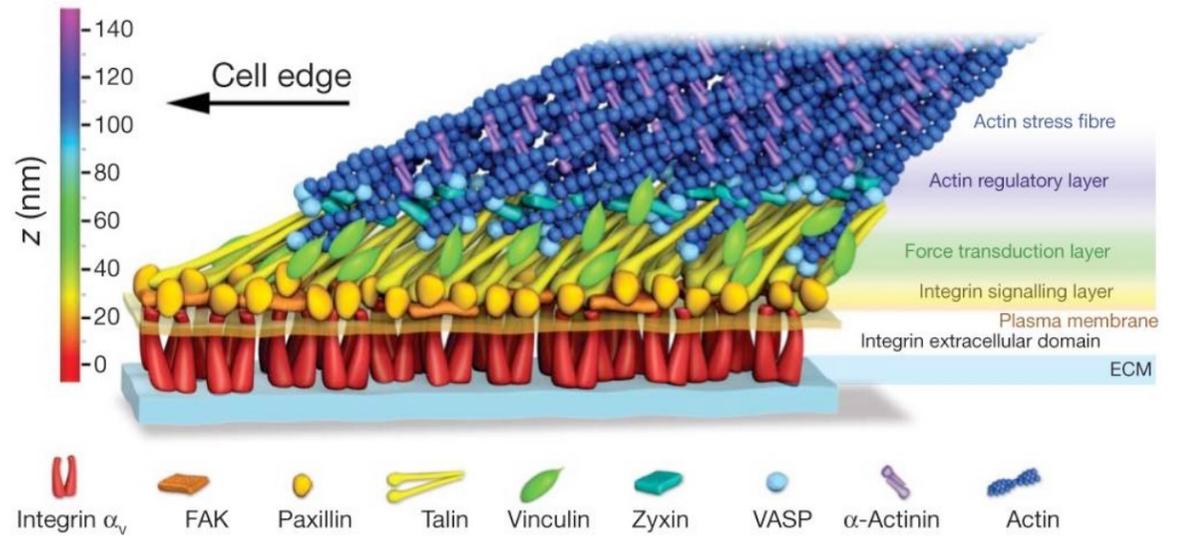
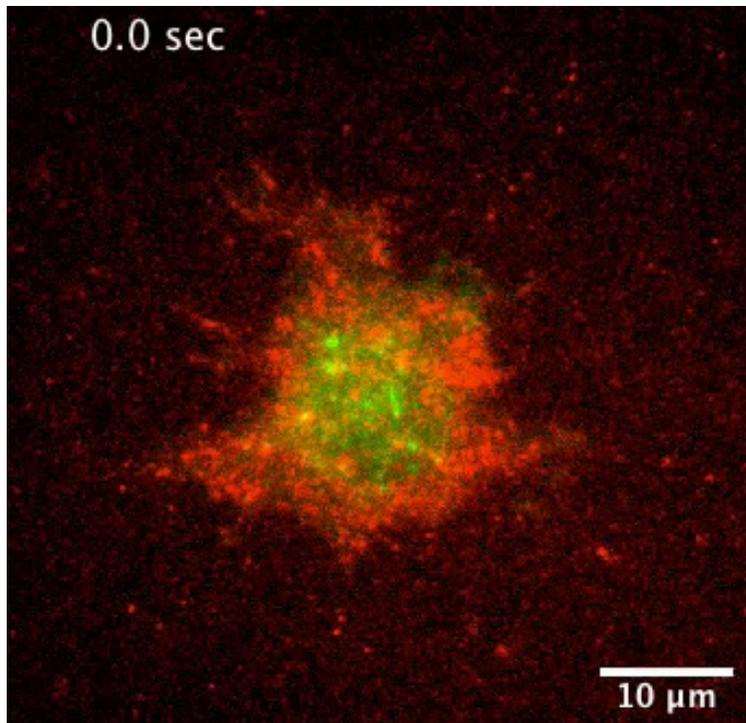


# Understand Complex Function in Cells

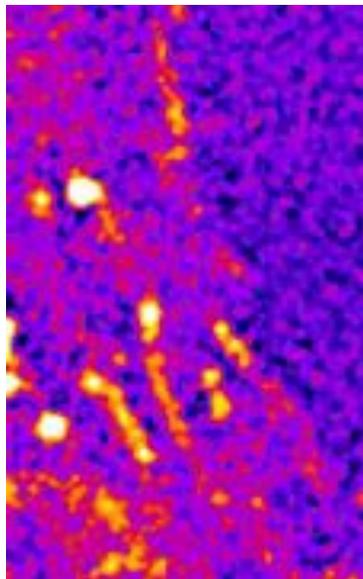
- Cells has many complex functions, integrated by different cellular states—high noise
- Need to standardize experimental procedure, isolate the events and repeat
- Rigidity Sensing- Cell Sensing Matrix-Cell Binding to Matrix- Cell Spreading Assay
- Have several different experiments to test the same thing



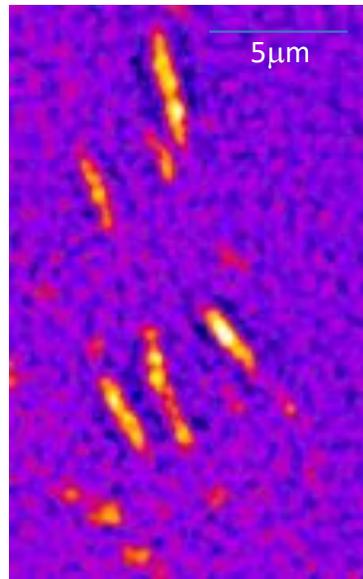
# Focal Adhesion



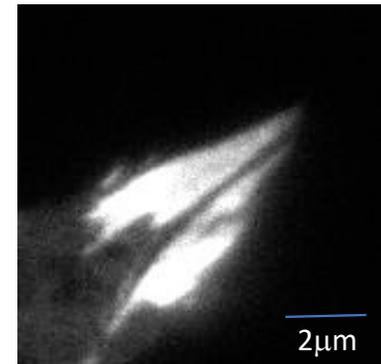
# Dynamics of Focal Adhesion Proteins



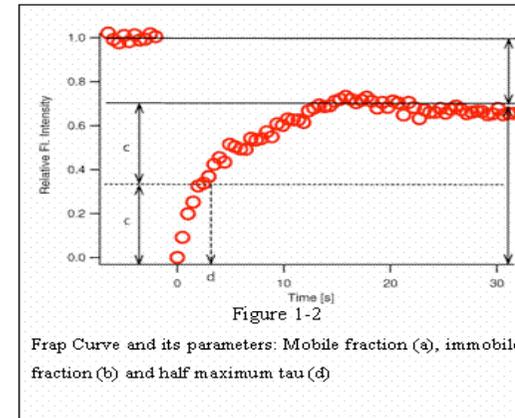
Talin-GFP



Vinculin-mCherry



Vinculin-GFP

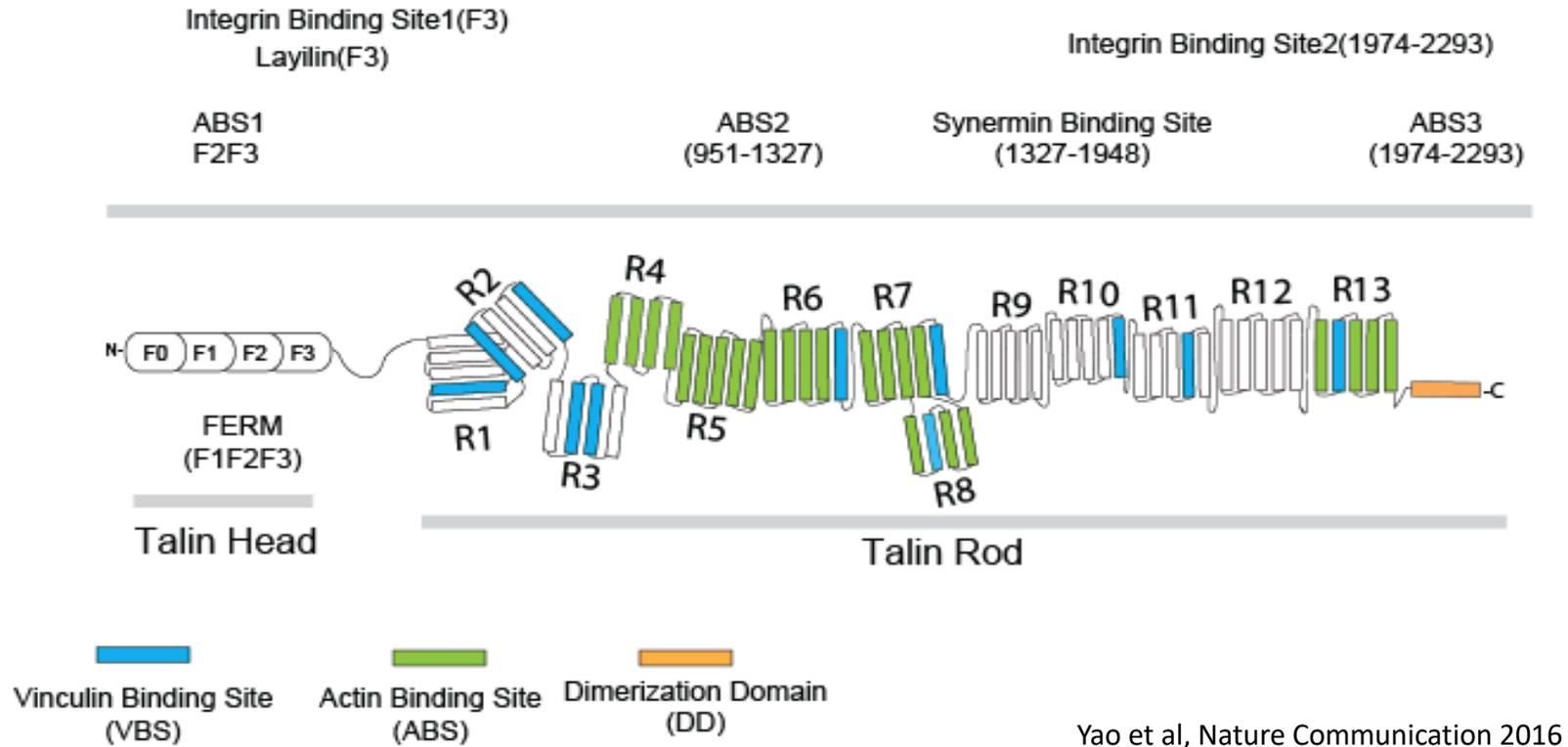


Glass/Fibronectin      MEF(Fibroblast)  
100ms/Frame  
Stream Mode

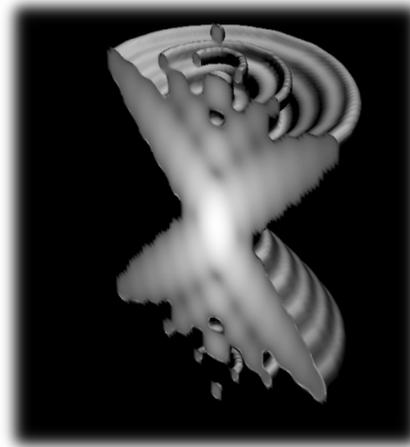
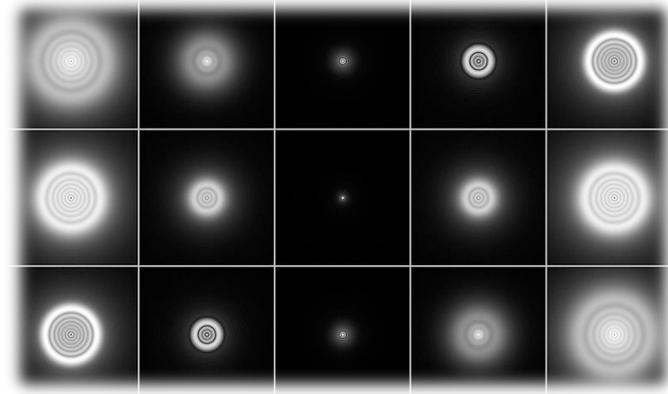
# Dynamics of FA Proteins

Protein Name	FRAP t1/2(s)	Cell Type
Vinculin	1.9/14/50/11/21	HelaJW/MEF/ REF52
Paxillin	1.5/25/17.7/53.2/20	HelaJW/MEF/ Bovine capillary endothelial cells/MEF, TIRF FRAP/ REF52
FAK	2.1/7/6.7/14	HelaJW/MEF/ Bovine capillary endothelial cells/ REF52
Zyxin	0.5/12/7/10	HelaJW/MEF/ REF52
Talin	2.2/23/77/18	HelaJW/MEF/ Bovine capillary endothelial cells/ REF52
ILK	9.2/12	HelaJW/ REF52
□-actinin	36/14.1/27	MEF/ Bovine capillary endothelial cells/ REF52
beta3-Integrin	38.5	Bovine capillary endothelial cells
p130Cas	5.8/14	MEF / REF52
Tensin	55	REF52
Src/CSK	9	REF52
Pinch	14	REF52
VASP	0.7/15	HelaJW/ REF52

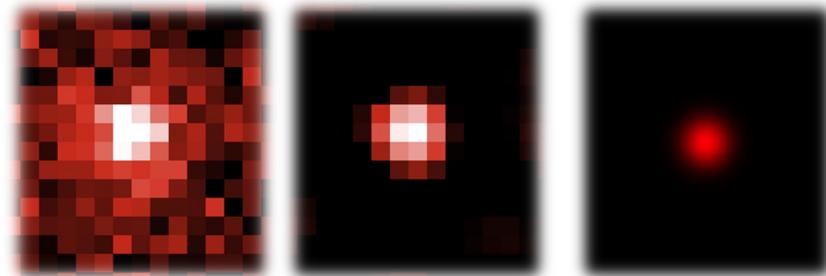
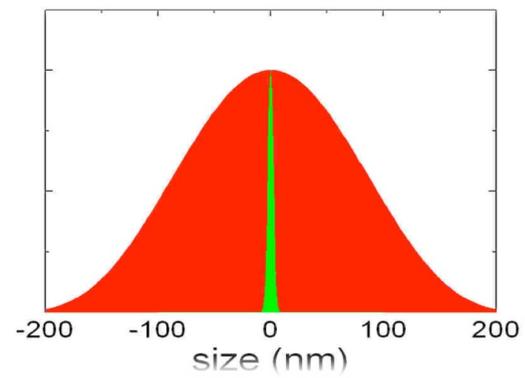
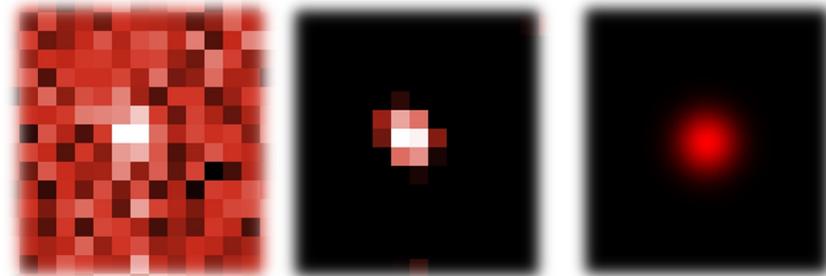
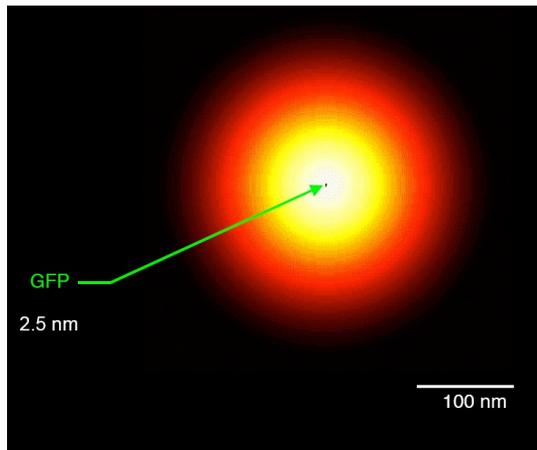
# Talin



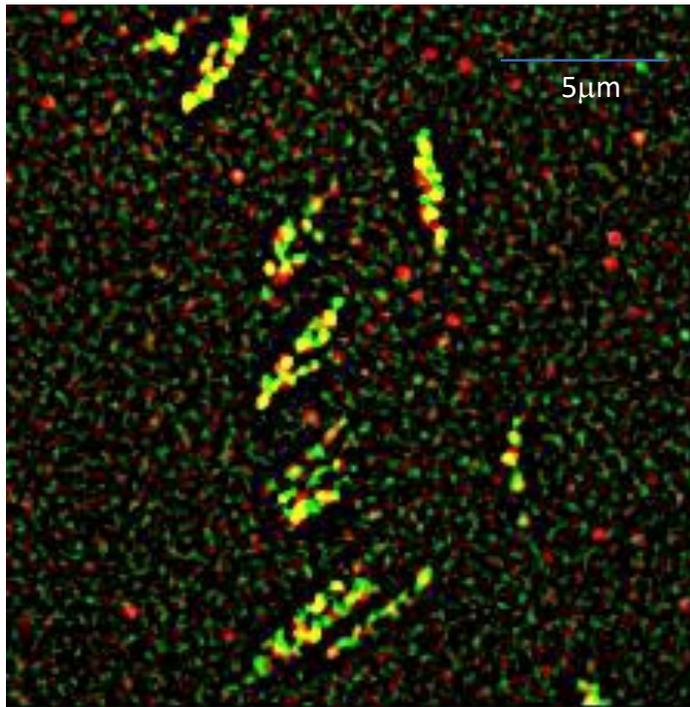
# Resolution Limit/PSF



# Single Molecule Localization



# Talin Stretching *in vivo*

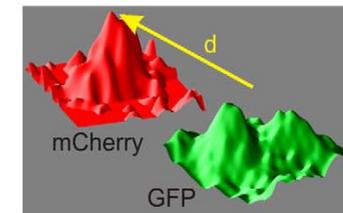
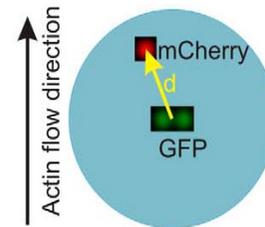
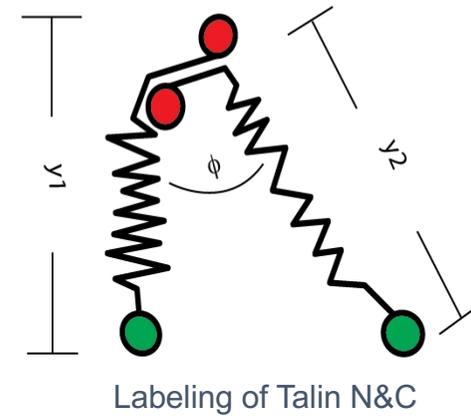


GFP-Talin-mCherry

Paired Single Molecule  
Live Cell Imaging  
site

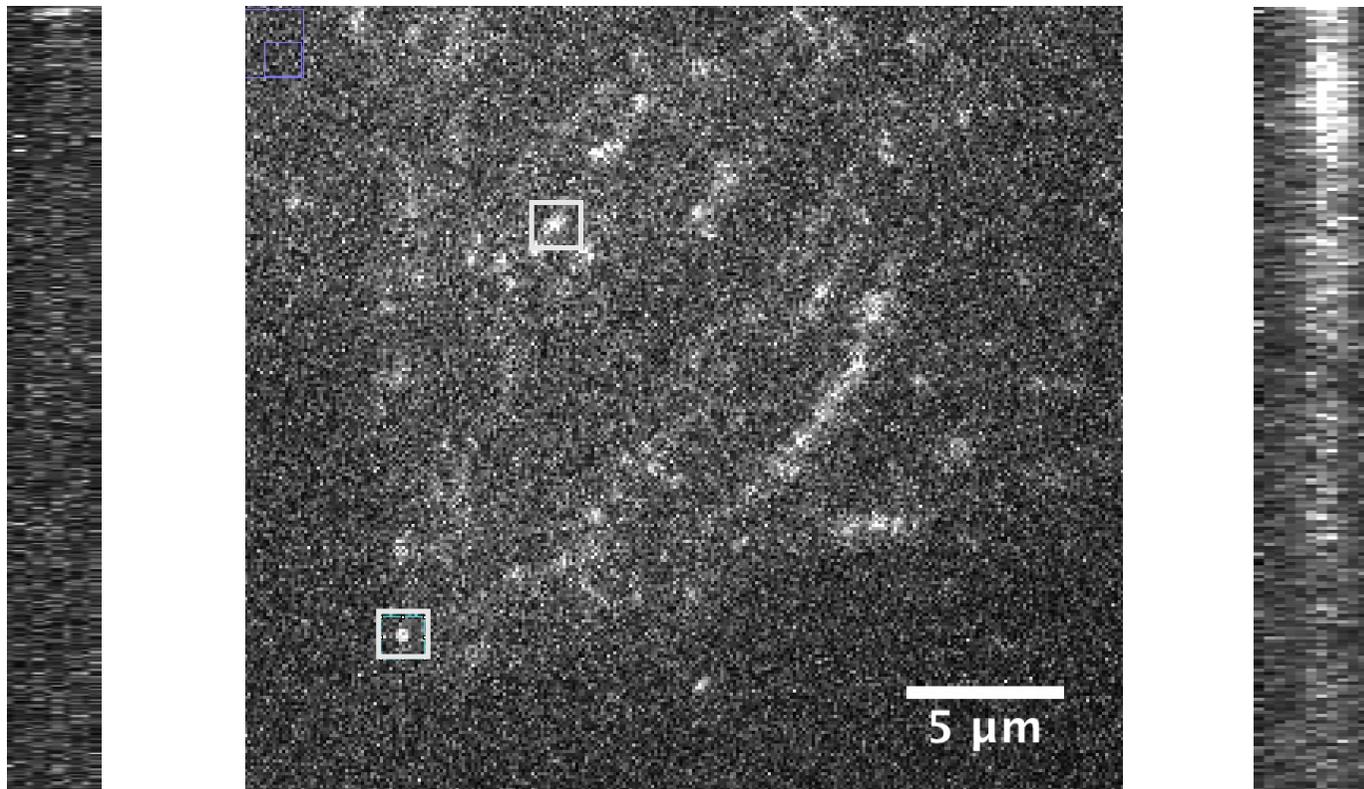
100ms/frame, stream mode, total duration 10s

GFP: integrin binding site  
mCherry: actin binding site

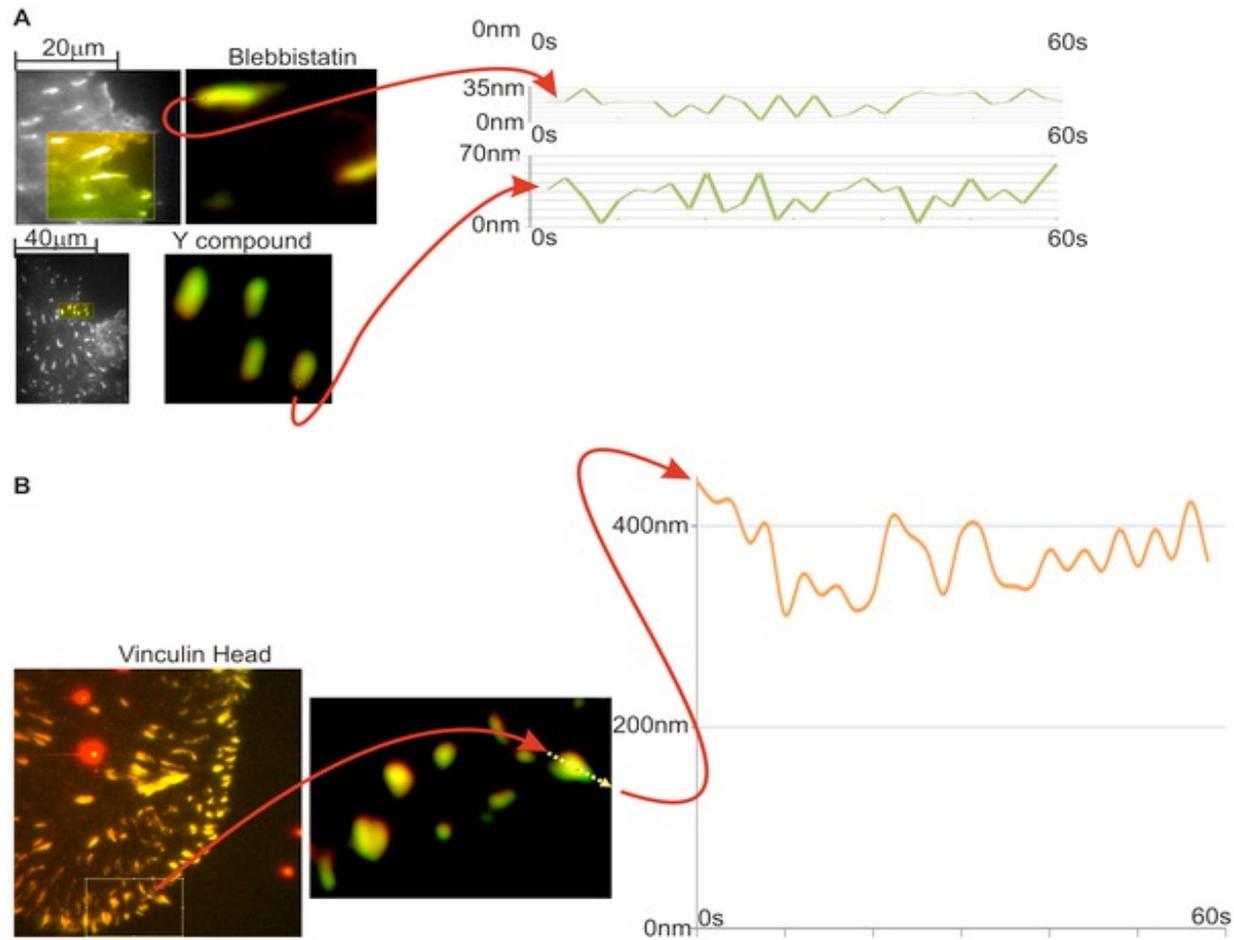


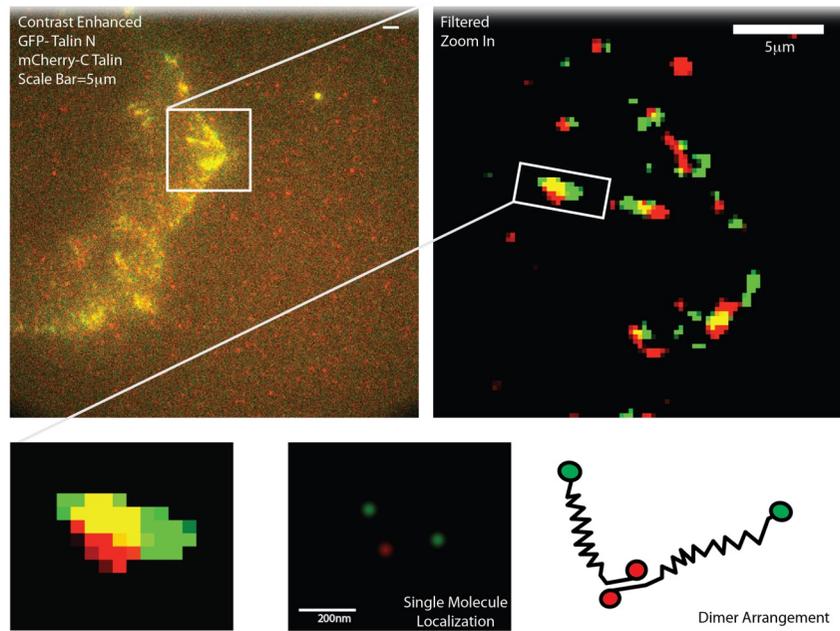
Dimer Tolerant Detector: Search for Signal Pairs

# Single Molecule Imaging

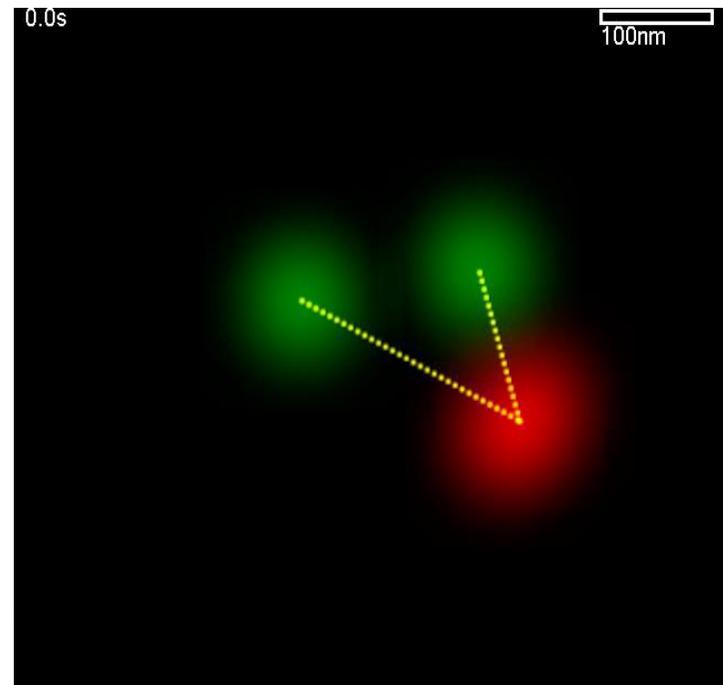


# Talin Stretching *in vivo*



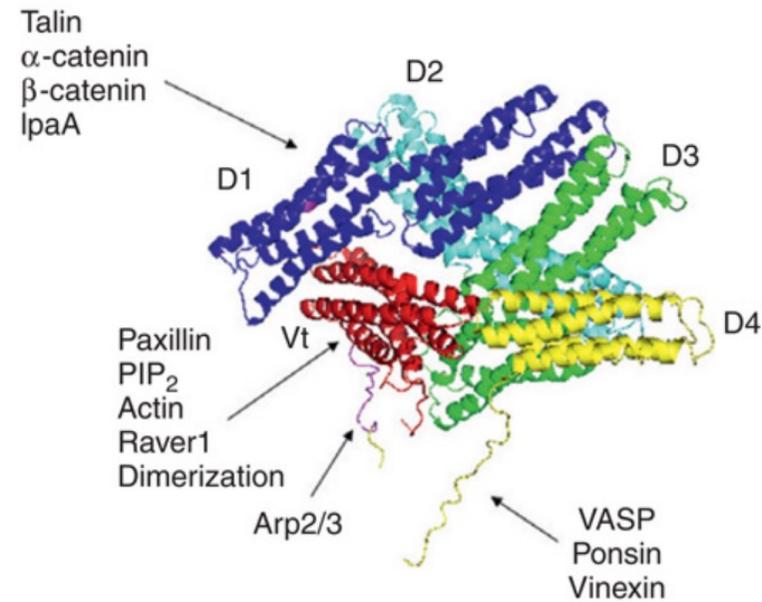


GFP-Talin-mCherry  
Paired Single Molecule Dynamics



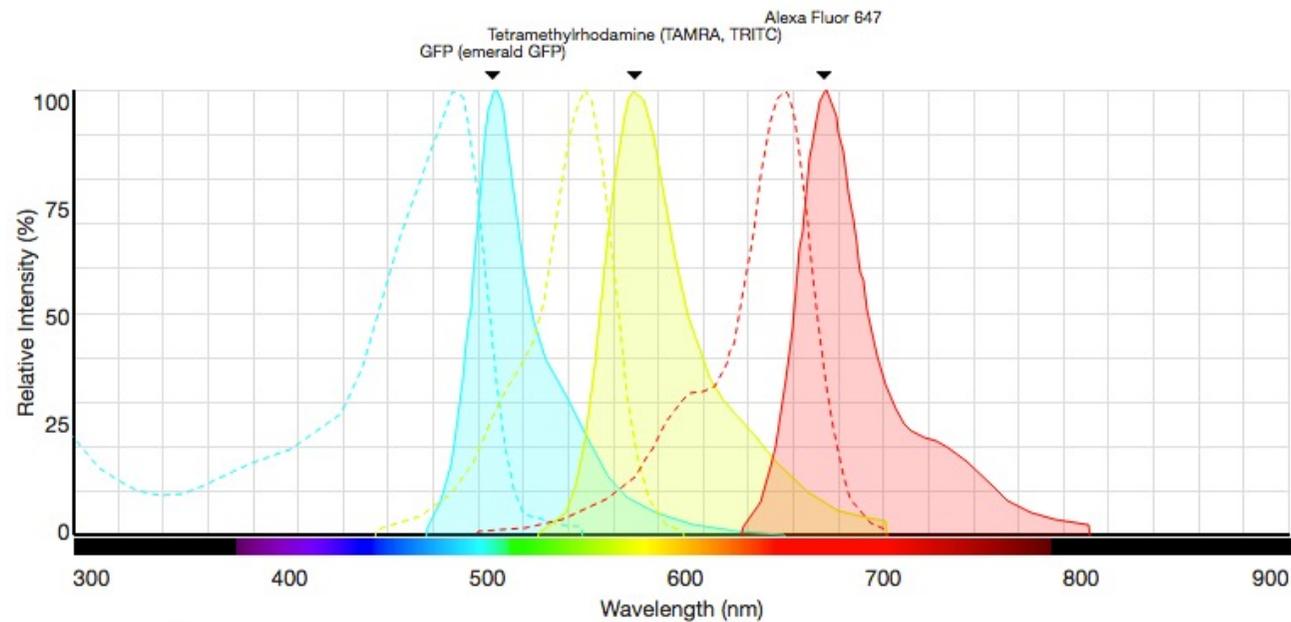
# Vinculin

- Function:
  - adaptor
  - non enzymatic activity
  - regulating FA assembly
  - force transmission
- Work with talin:
  - one talin binding site
  - activated by talin



# Vinculin and Talin

- **Question:** vinculin recruitment event *in situ*
- **Challenge:** Three color live cell imaging with single molecule localization accuracy.
  - Wavelength: ~~Blue, Green, Red~~, Far-red
  - Short term imaging
  - Superresolution localization precision
  - Organic dyes: delivering system: eDHFR-TMP



**Legend**

- GFP (emerald GFP)
- Tetramethylrhodamine (TAMRA, TRITC)
- Alexa Fluor 647

**Key**  
Excitation: - - - Emission: - - - -

Pointer Location: 635nm, 7%  
Fluorophores: Tetramethylrhodamine (TAMRA, TRITC), Alexa Fluor 647

**Fluorophores** | **Light Sources** | **Excitation Filters** | **Emission Filters** | **Reset**

Category (Optional)	Fluorophore	Excitation	Emission	Action
1. <input type="text" value="Click to Select"/>	GFP (emerald GFP)	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	Remove
2. <input type="text" value="- All Categories -"/>	Tetramethylrhodamine (TAMRA, TRITC)	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	Remove
3. <input type="text" value="Click to Select"/>	Alexa Fluor 647	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	Remove

+ Add

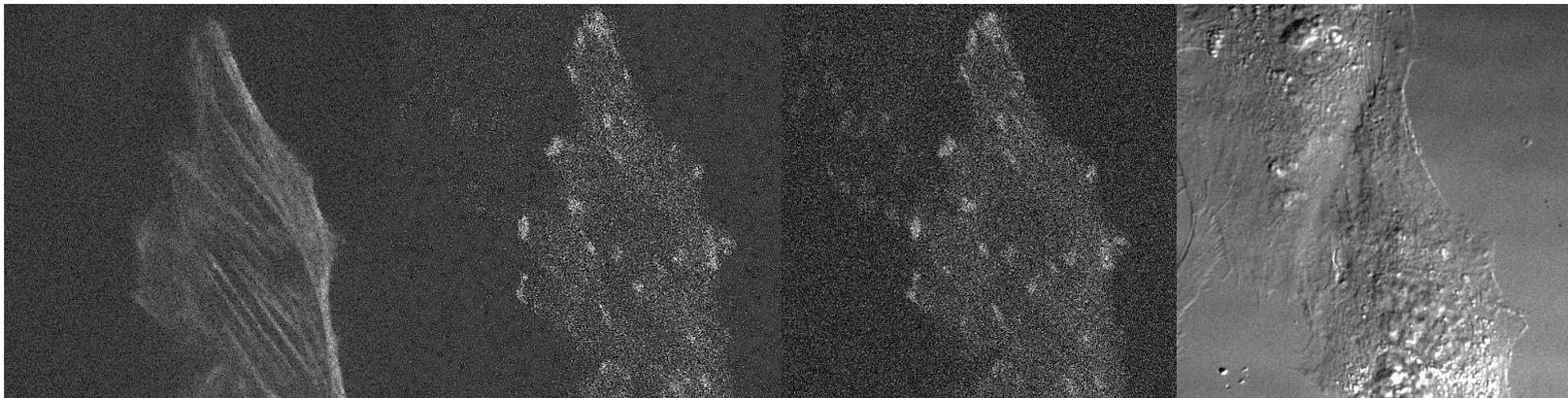
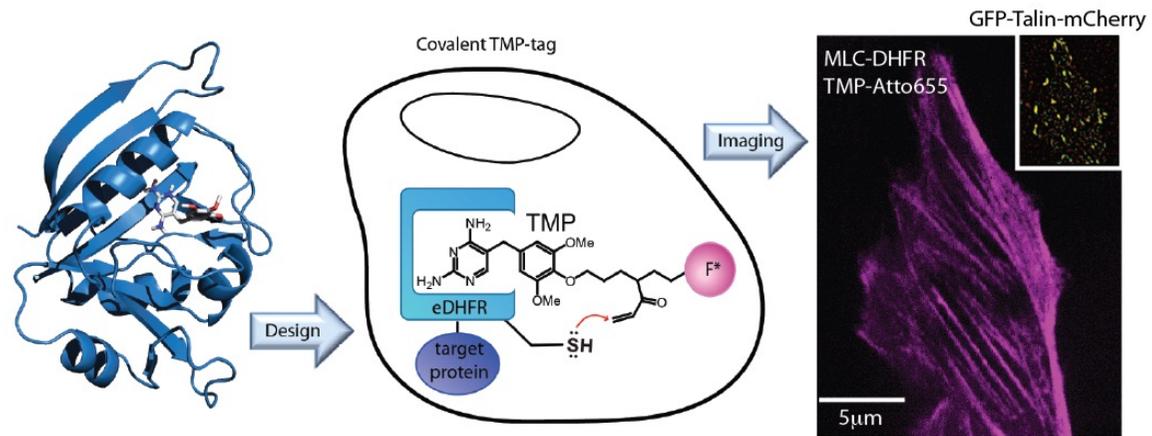
**Display Options**

- Grid
- Labels
- Excitation Plots
- Emission Plots
- Light Sources
- Filters
- Show Emission Plot Normalization

## Choosing a Fluorophore

Excitation and emission	Together with filter set/laser line
Brightness	quantum yield, extinction coefficient
Photo stability	quenching and photo-bleaching rate
Size	GFP 27kDa
Blinking or photometry	Localization or quantitation
Maturation speed	mCherry ~ 15min
Labeling Ratio	monomer/oligomer
Delivering method	SNAP, CLIP, eDHFR-TMP
Biocompatibility	Membrane Permeability, Cytotoxicity, PH stability(pKa)

# Labeling Vinculin



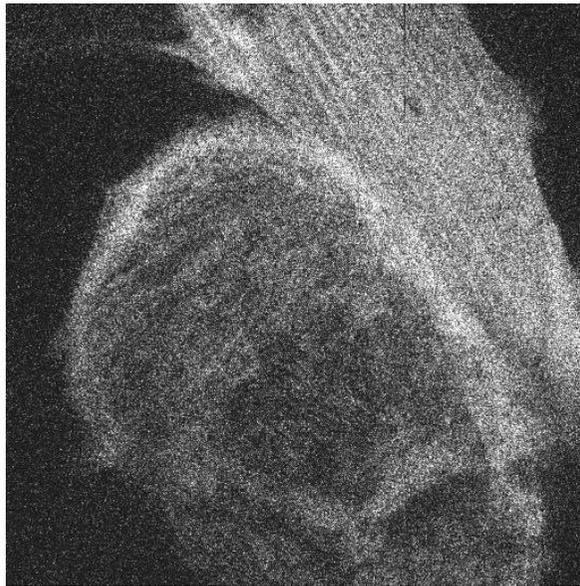
MLC-DHFR, TMP Atto655

GFP-Talin

Talin-mCherry

DIC

# BioCompability



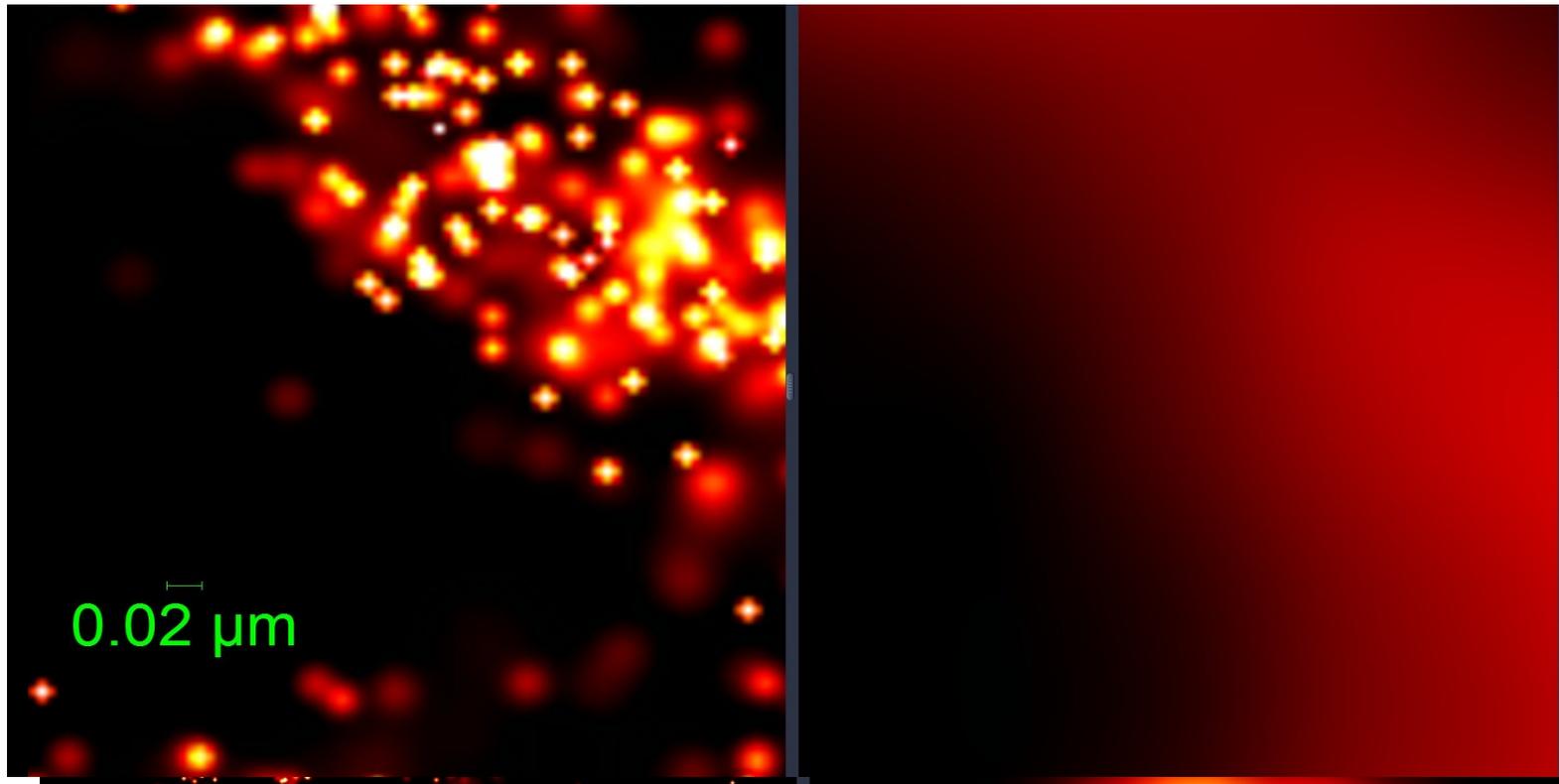
20 uM TMP-Fluorescein,  $\tau=55$  sec N=5



20 uM TMP-Atto655,  $\tau=64$  sec N=5



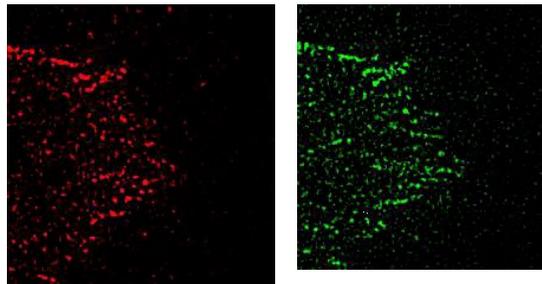
# Single Molecule Localization

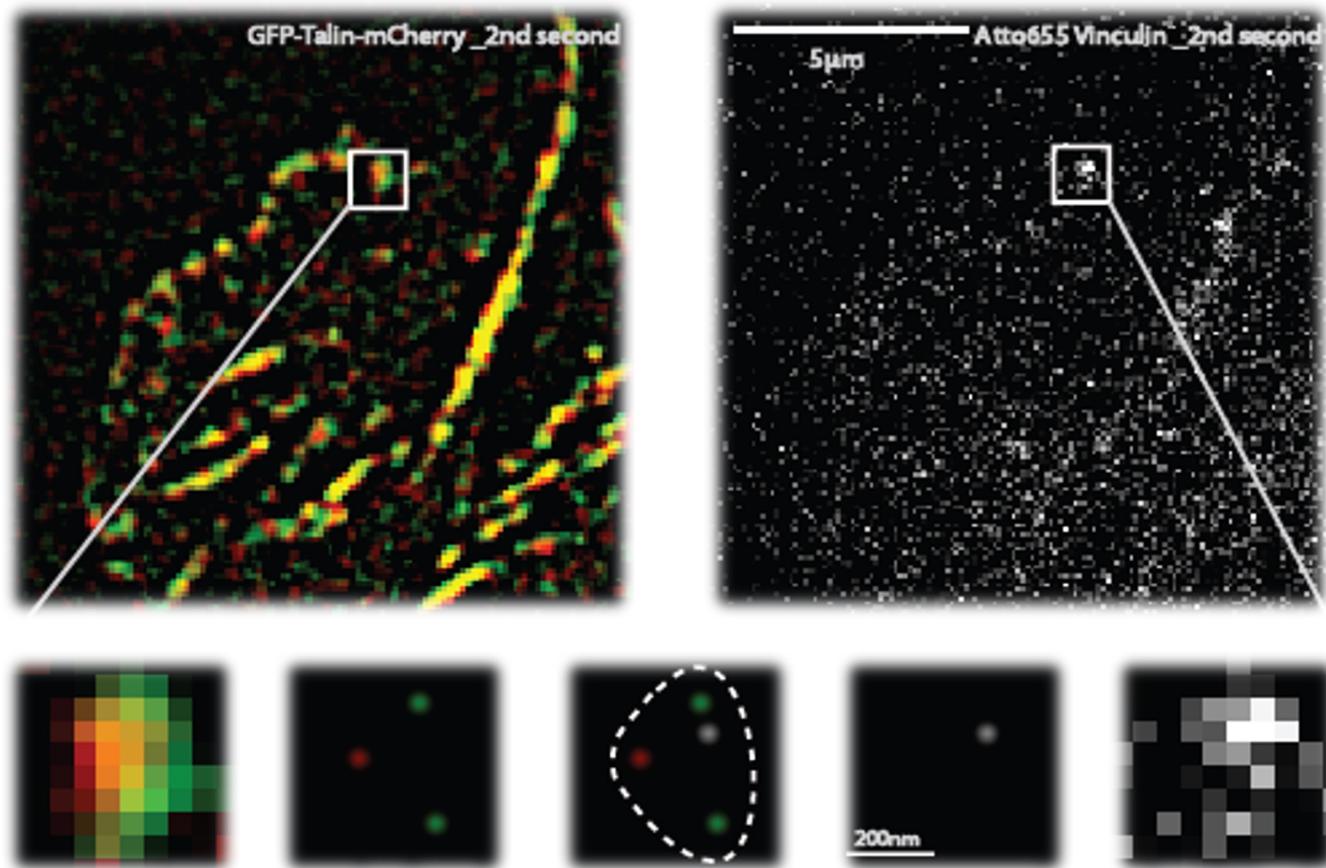


5 $\mu$ m



GFP-Talin-mCherry  
Paired Single Molecule

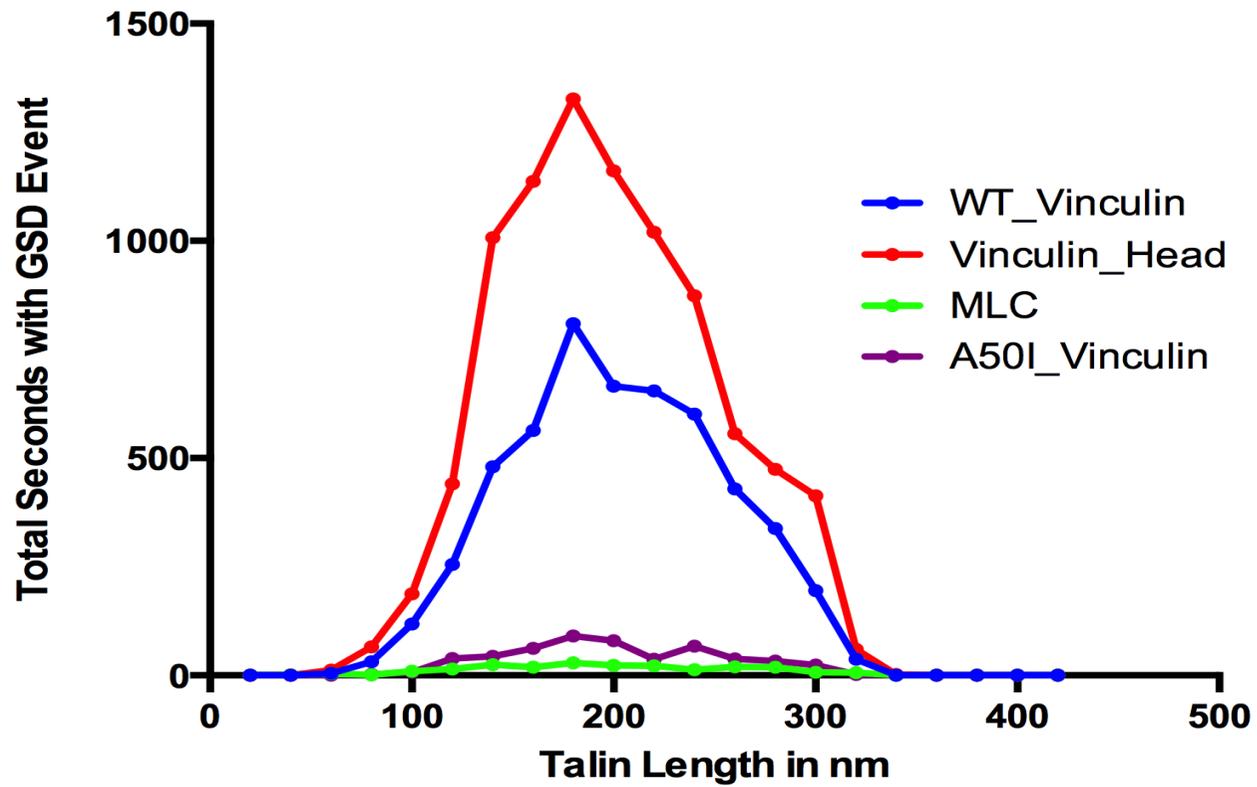


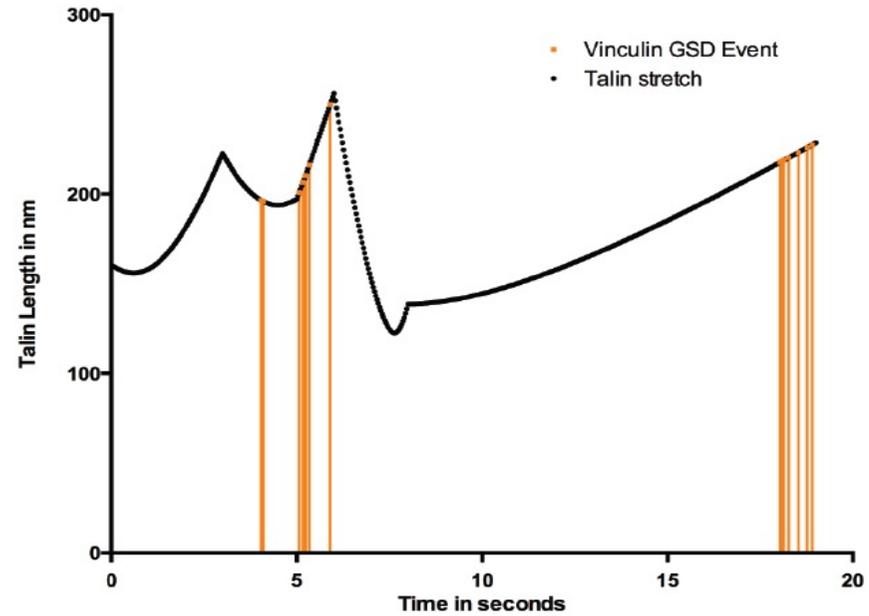
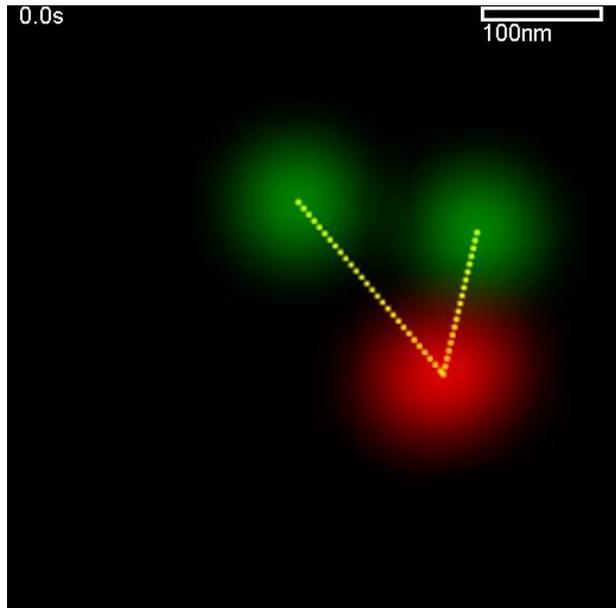


- Simultaneous Imaging of
  - GFP: N terminal, Integrin Binding
  - mCherry: C terminal, Actin Binding, Dimerization
  - Atto655: Vinculin

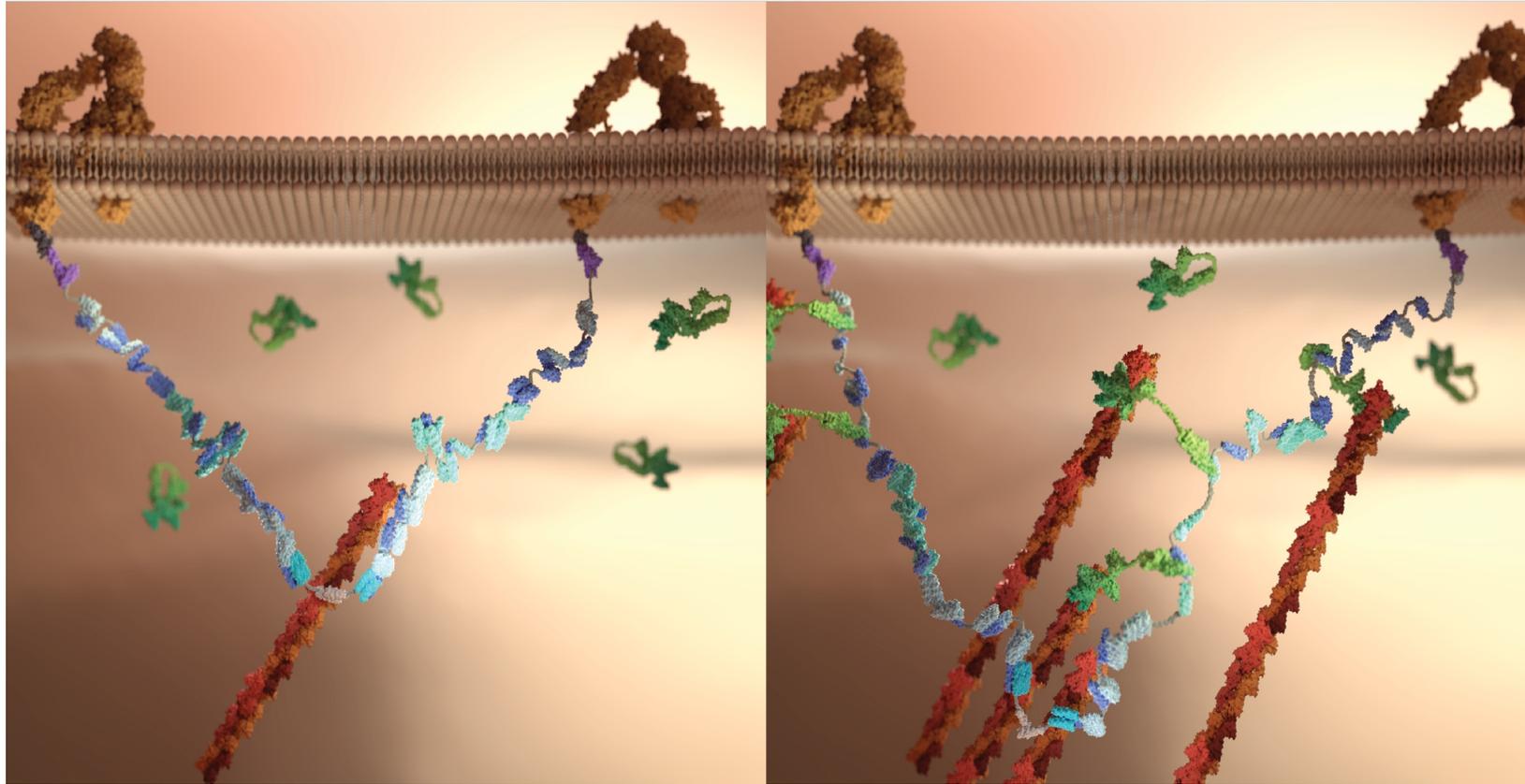
Hu.et al, Nano Letters, 2016

Vinculin binds to talin most when talin is 180nm





- GFP: N terminal, Integrin Binding
- mCherry: C terminal, Actin Binding
- Atto655: Vinculin



Integrin Complex

F-actin

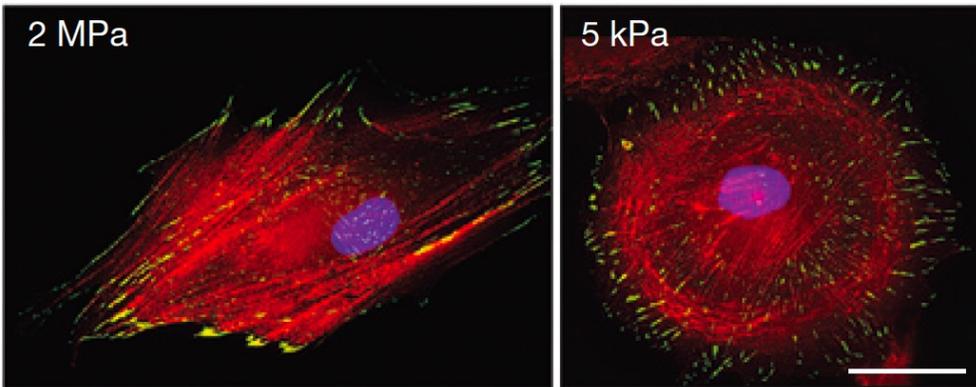
Talin (Unstretched)  
Talin (Stretched)

Vinculin(Auto-inhibition)  
Vinculin(Activated)

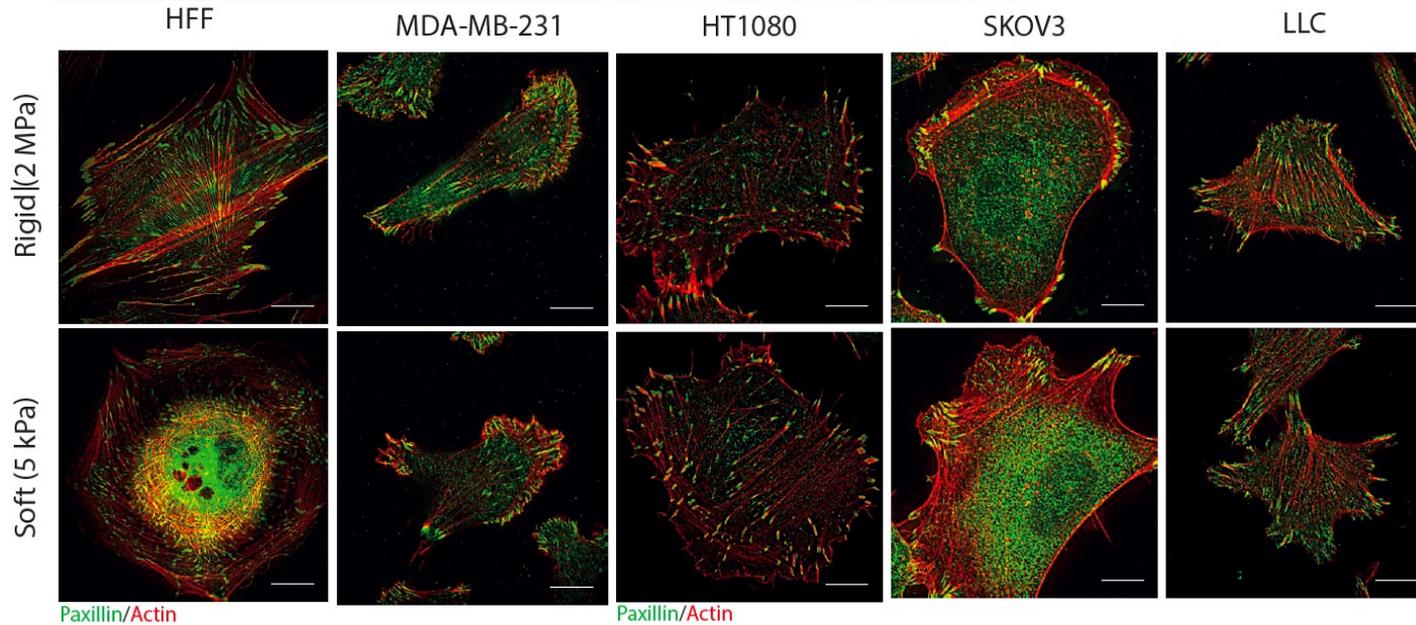
Hu et al, Pro Sci. 2017

# Rigidity sensing is impaired in transformed cell

6 h



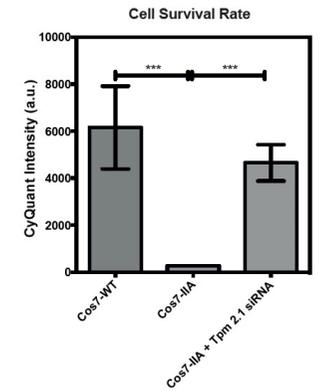
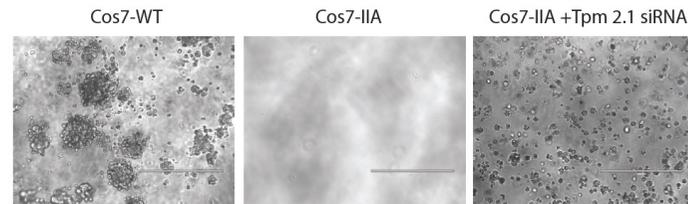
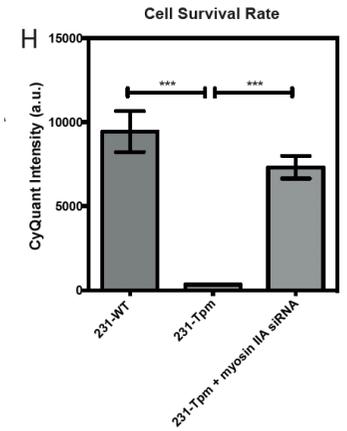
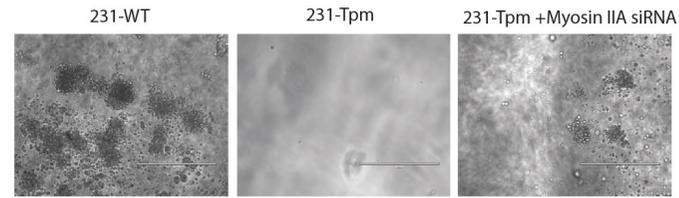
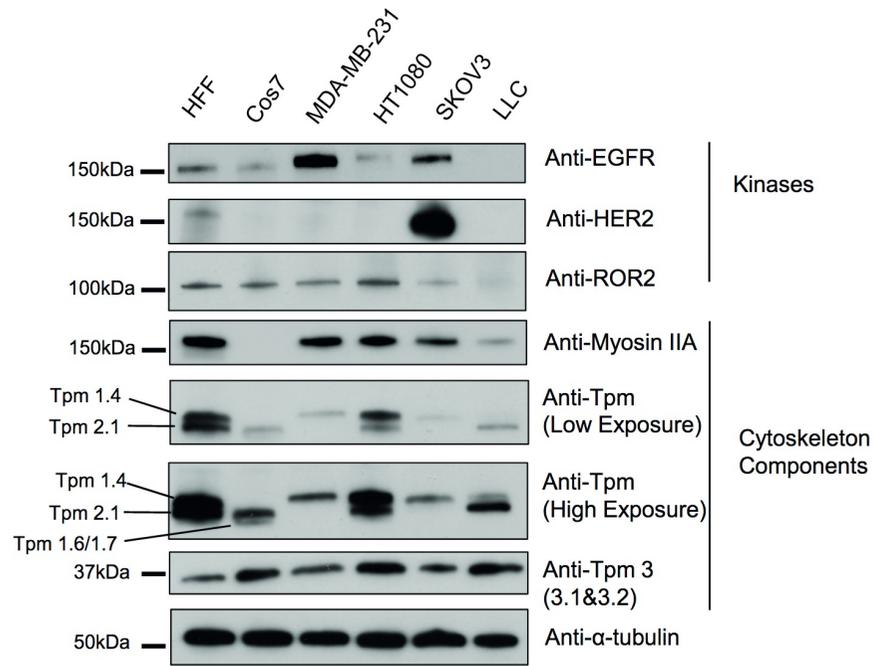
Normal Cell behave differently on soft substract



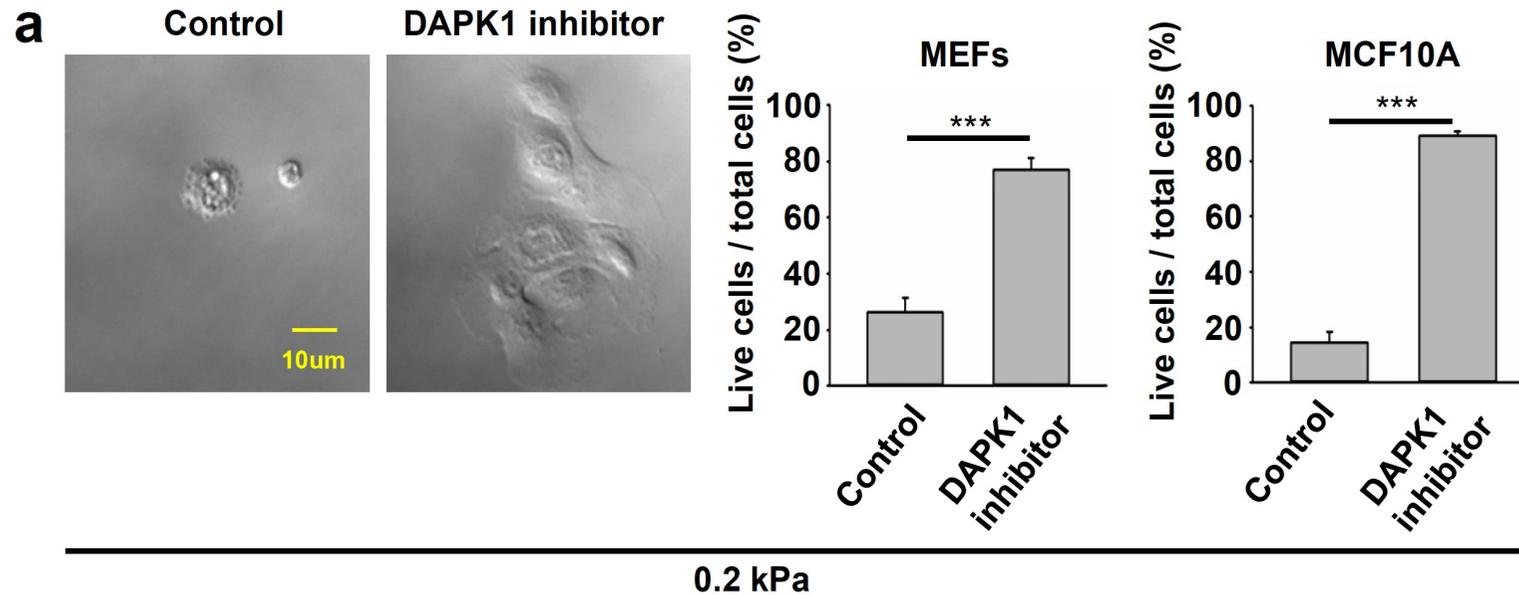
Prager-Khoutorsky, M. *et al*; Nat Cell Bio 2011

Yang, B. , Wolfenson, H., Nakazawa, N. *et al*, Nat Mater 2019

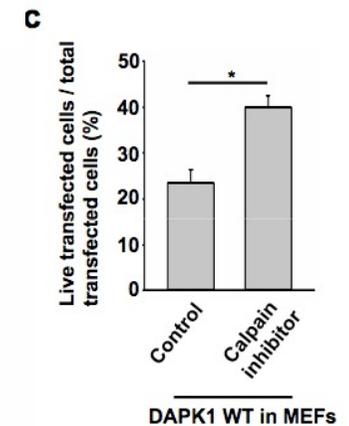
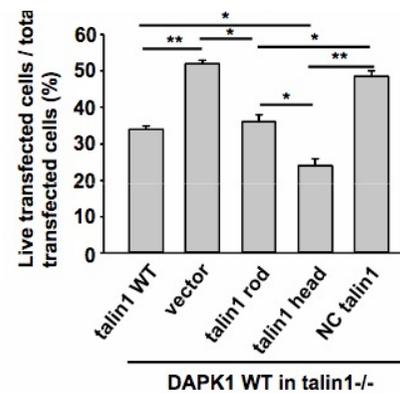
# The Missing Mechanosensing Component



# Missing Link between Sensing and Apoptosis



- DAPKI activity linked with Talin cleavage



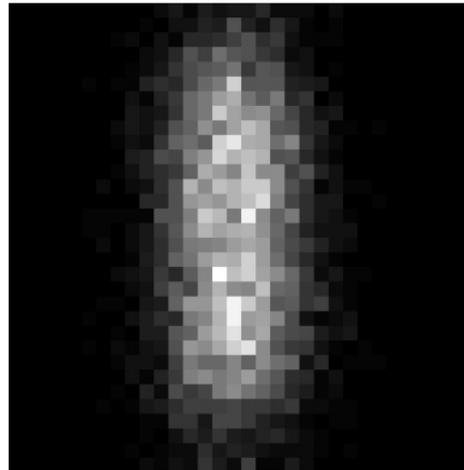
# Bio Imaging For Quantitative Measurement

- Qualitative and Quantitative
- What Makes Quantitative Measurement Possible:
  - Scientific grade detectors are linear
  - Fluorescent proteins are linear

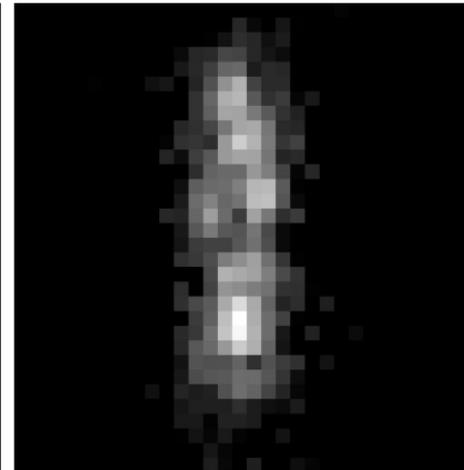
- **Live Cell Imaging: Photon budget**
- Artifacts in Quantitative Imaging
  - Protein expression level?
  - Bleaching
  - Laser power
  - Light efficiency of microscope
  - Undersampling



line object



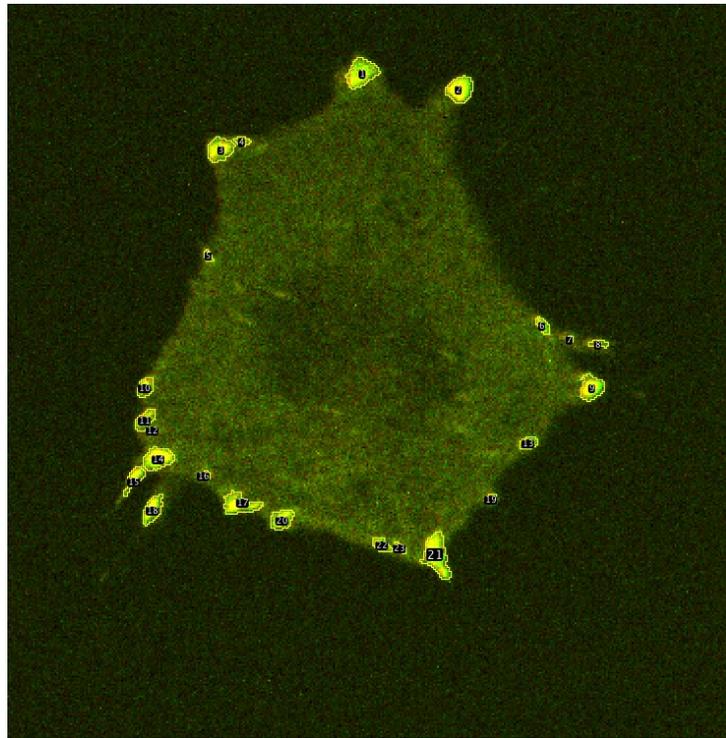
convolved with PSF and Poisson noise



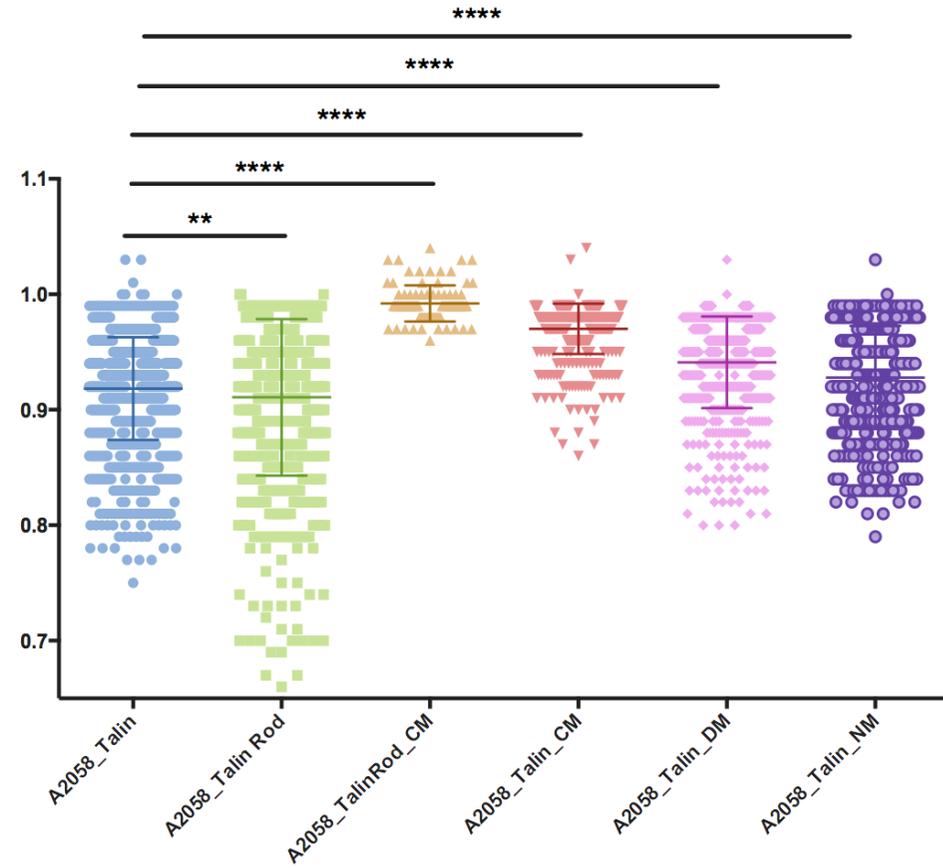
Undersampling simulation

Simulation by Felix Margadant

# mCherry/GFP Ratio

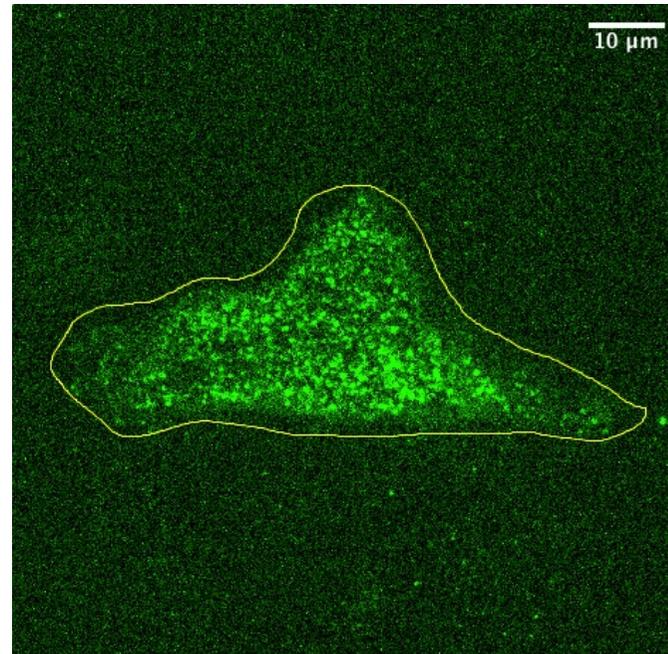
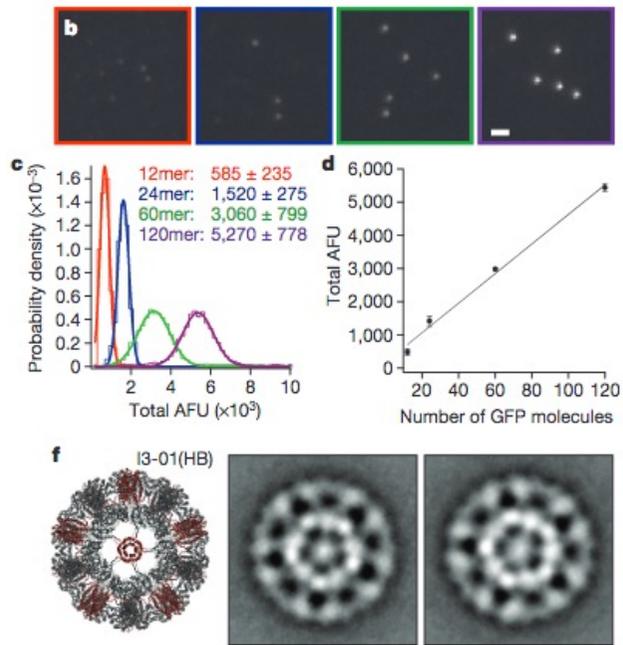


Ratio of mCherry vs GFP in Fas, A2058, SiTalin  
Sample Size each group 500+

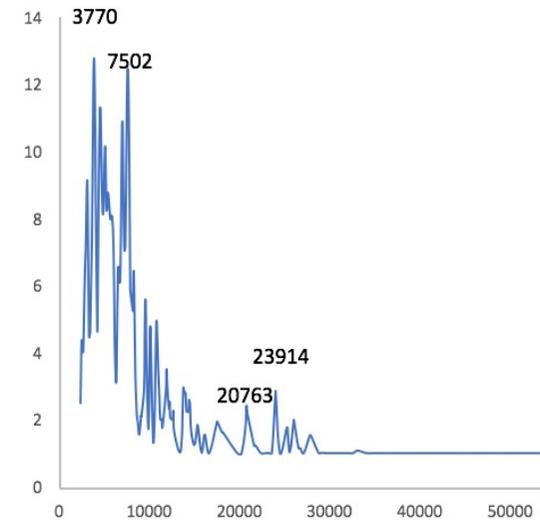


# mCherry/GFP Ratio- Calibration

- GFP2-tag, icosahedron, I3-01(14nm)
- Cloned to 12mer mCherry, 12mer GFP, 60mer mCherry, 60mer GFP



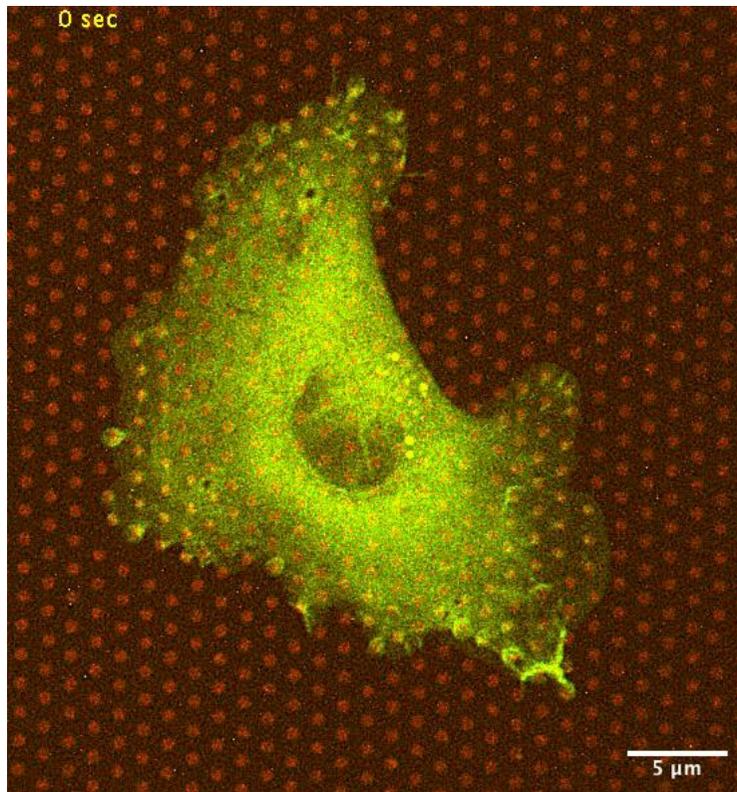
60Mer-GFP



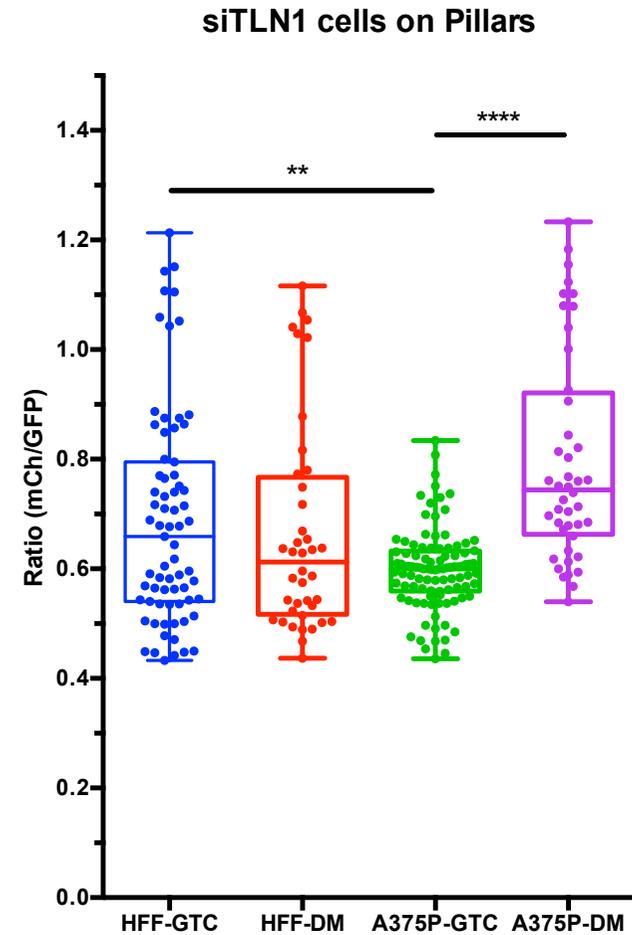
mCherry-60Mer Peaks

1 single GFP intensity ~ 61.2

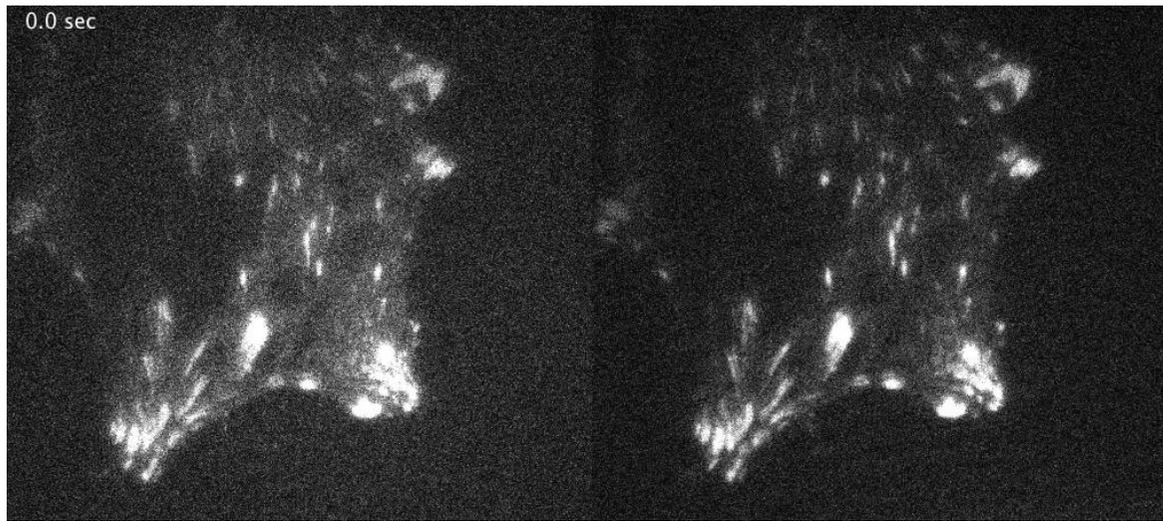
# More Cleavage in Transformed Cells on Soft surface



REF, GFP-Talin-mCherry, 2 $\mu$ m pillar, 6 hours seeding



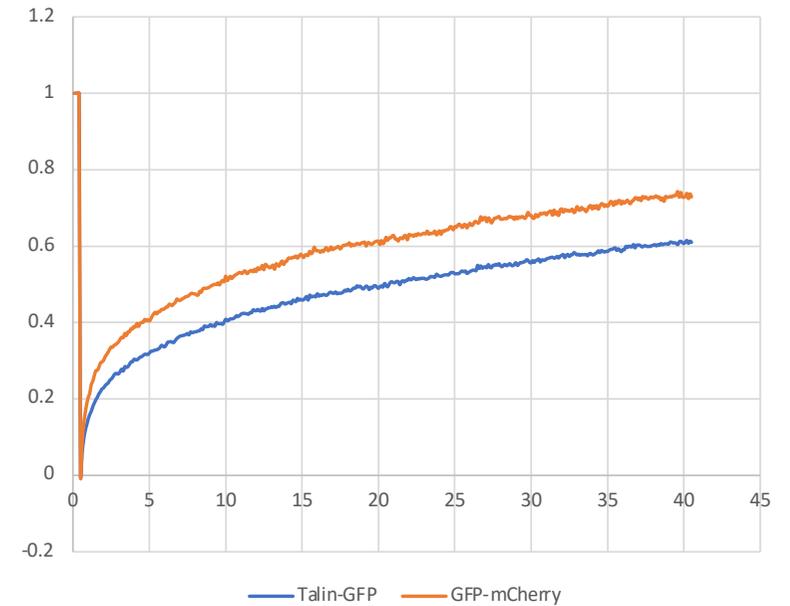
# FRAP Difference between GFP/mCherry



mCherry-C

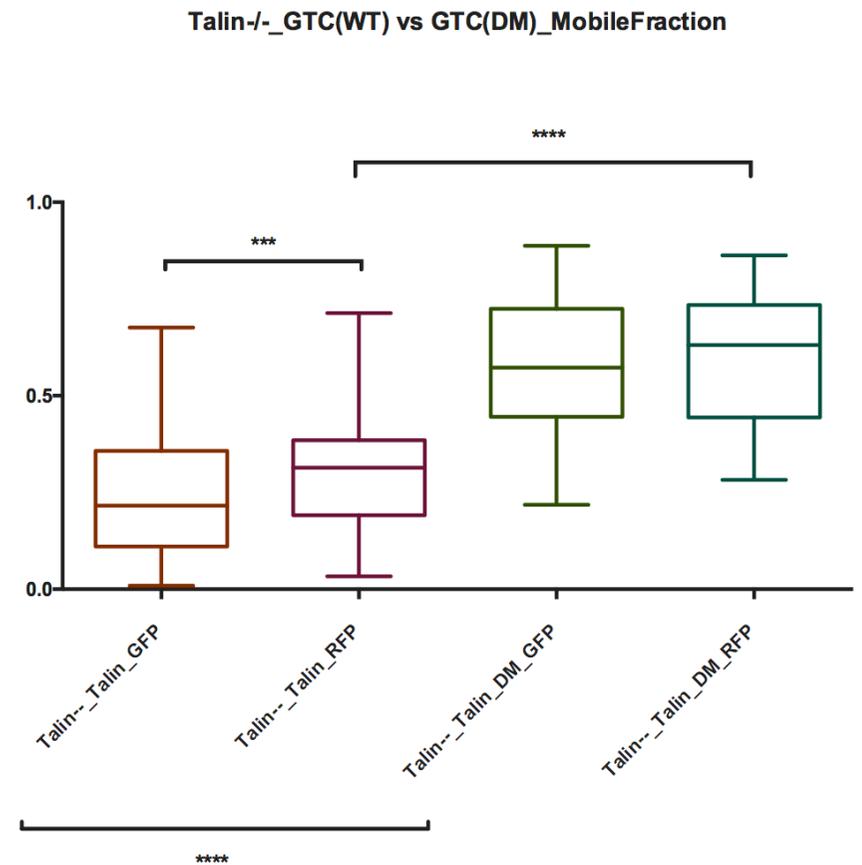
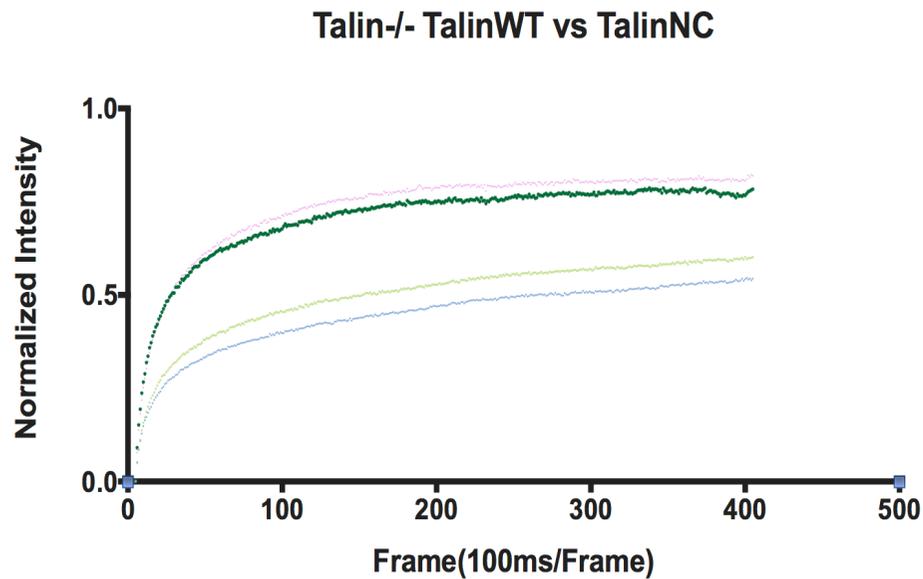
GFP-Talin-mCherry

GFP-N

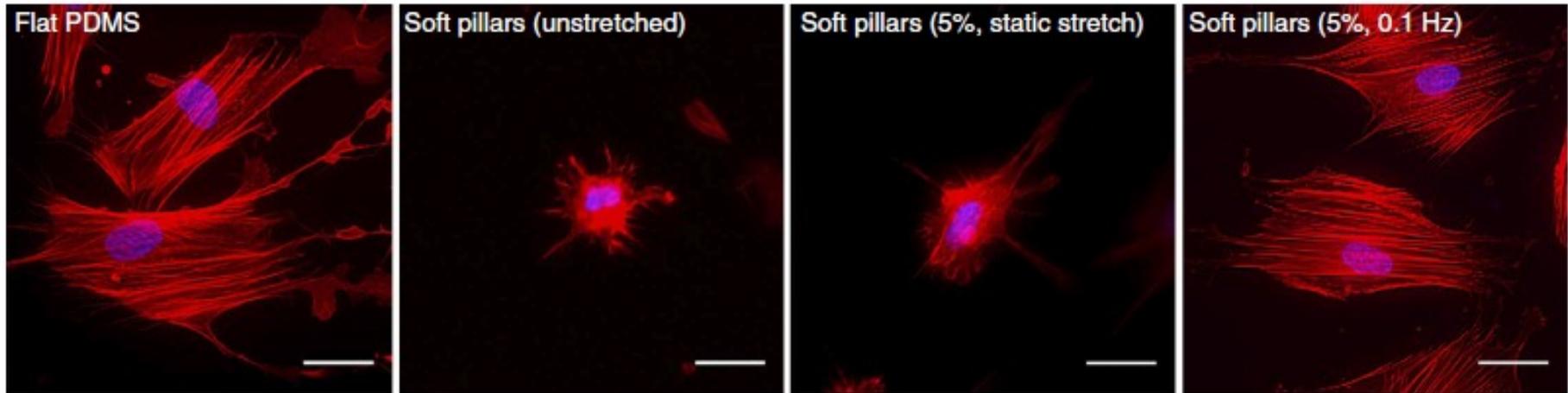
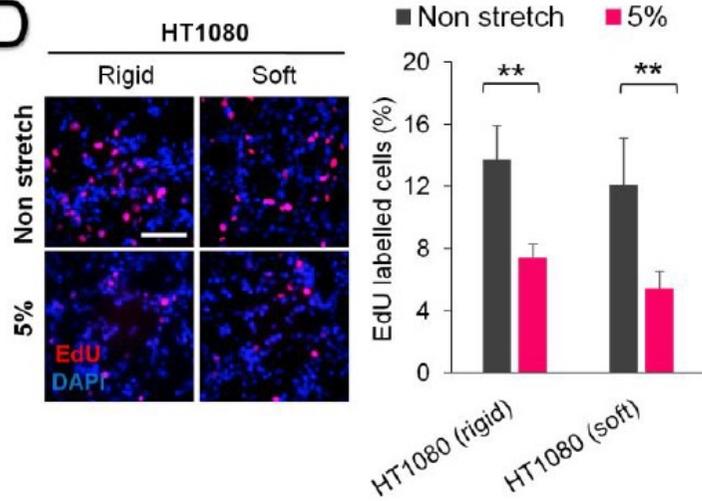
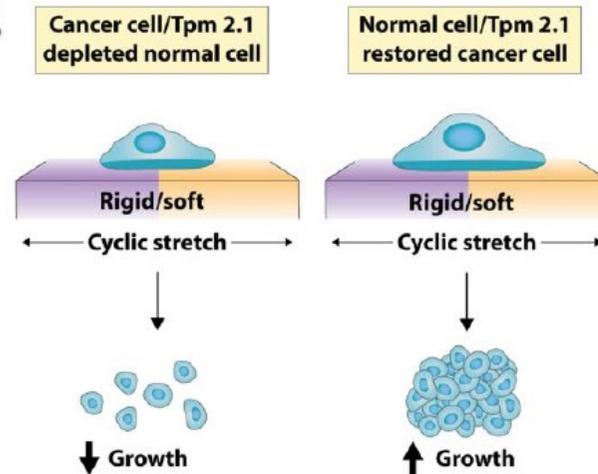


RPTP- GFP-talin-mCherry Dual FRAP, Sample Size: 25 cells

# FRAP Difference between GFP/mCherry



Tau of four groups shows no statistically significant difference, median around 2 sec

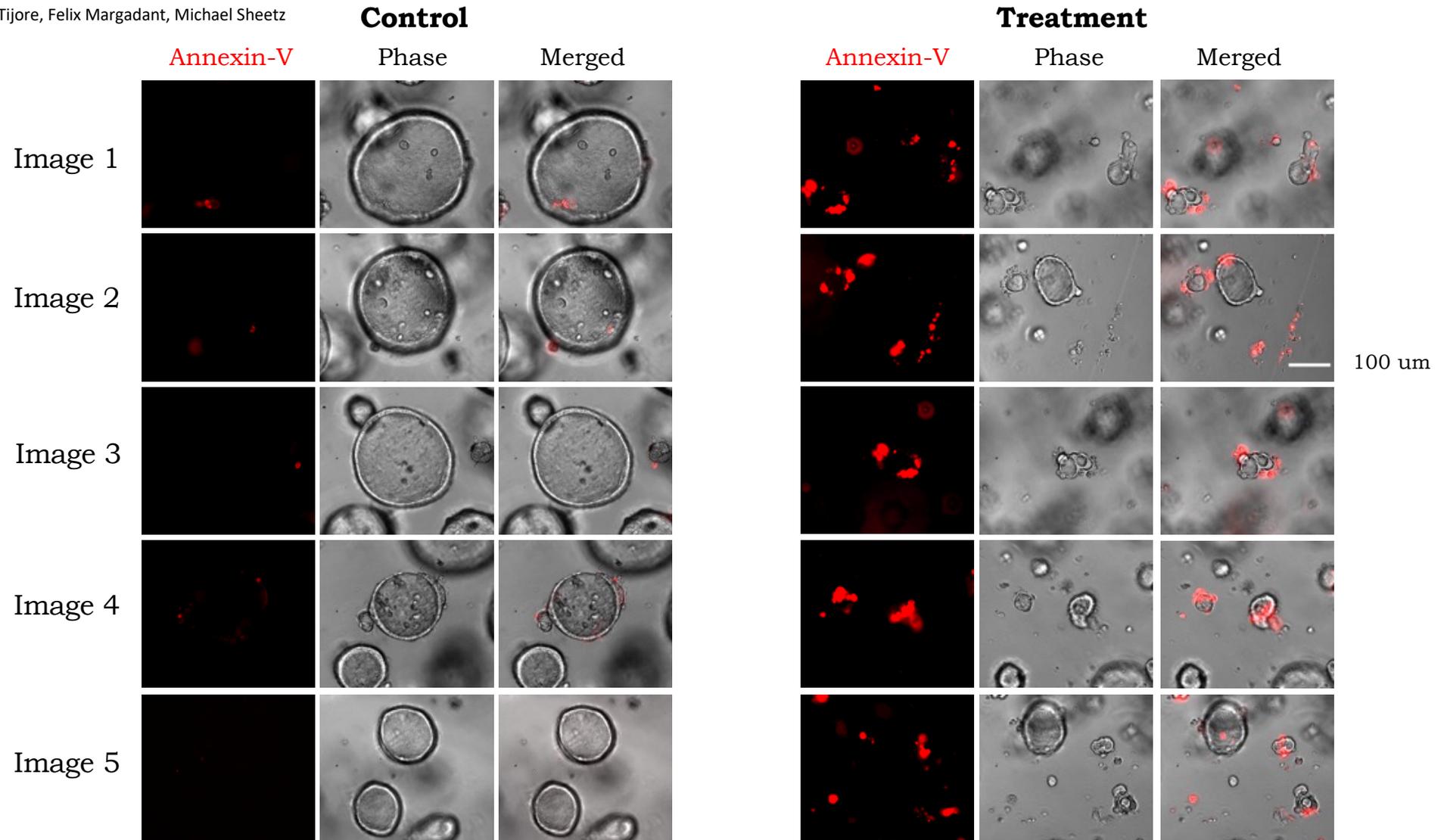
**a****D****E**

Cui et al, Nat Commun, 2015

Tijore et al, BioArchive, 2018

# Ultrasound Promotes Apoptosis in Pancreatic Tumor Organoids

Ajay Tijore, Felix Margadant, Michael Sheetz



Thank you



UiO : **University of Oslo**



Prof. Michael Sheetz  
Prof. Alexander Bersdhasky  
Prof. Henry Yu  
Prof. Oddmund Bakke

*Talin Project:* Xian Hu, Salma Jalali, MingXi Yao, Felix Margadant, Michael Sheetz

*Endosome Project:* Xian Hu, Duarte Mateus, Vinodha Manovasegaran,  
Felix Margadant & Oddmund Bakke

# 9th NorMIC Workshop on Microscopy Image Processing

29th Nov to 2nd Dec, 2022, University of Oslo



## Topics Covered:

Image Formation  
Image Processing Pipelines  
Segmentation  
Localization and Tracking  
Deconvolution  
Image J From Basic to Scripting  
Surface Rendering

## Faculty:

Rainer Heintzmann, Jena, Germany  
Felix Margadant, Radium Hospital, Oslo  
Ping Ching Cheng, Buffalo, USA  
Kay Schink, Radium Hospital, Oslo  
Frode Skjeldal, UiO  
  
Harald Stenmark  
Oddmund Bakke, UiO  
Xian Hu (Edna), UiO