

# Microsystems Design, 11/11-2008

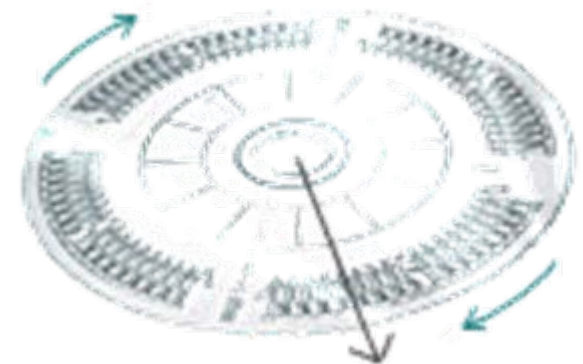
- Surface tension
- Capillary forces
- Ideal gas law
- Viscosity
- Navier Stokes equation
- Reynolds number
- Poiseuille flow
- Electroosmotic flow
- Electrophoresis
- Mixing

These topics are important for design of well-functioning fluidic microsystems.



# Gyros AB microfluidics platform

- world leader in the miniaturization and integration of laboratory applications through its proprietary microfluidics platform, Gyrolab CD (compact disc) microlaboratory.

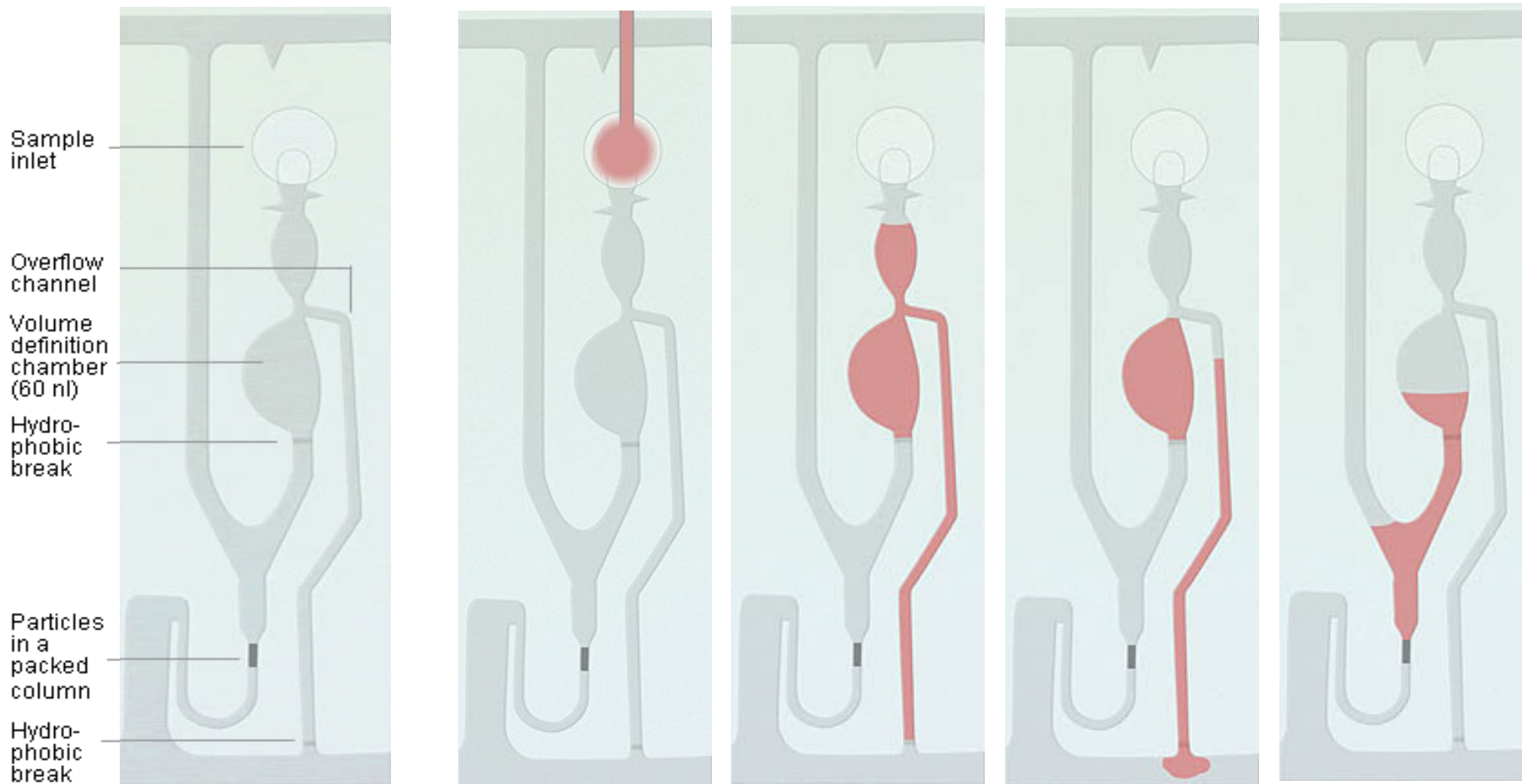
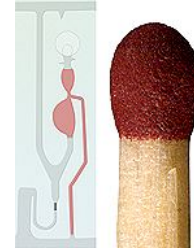


# Gyros AB, Sweden

## GyroLab CD microlaboratory

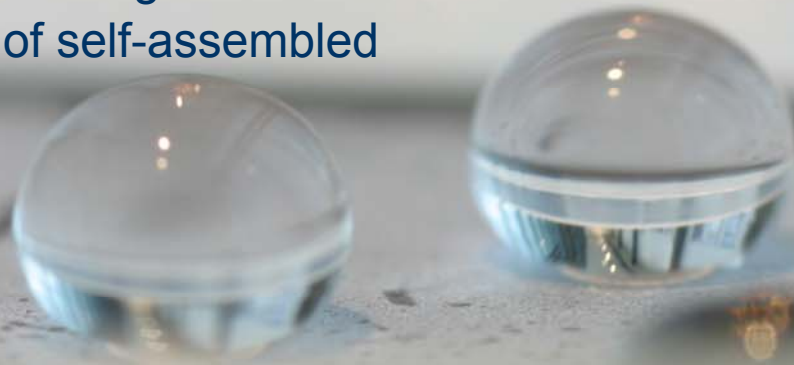
- Protein quantification
  - Microchannels in polymer disk
  - Multiple analyses
  - Centrifugal forces
- 
- Instrument
  - Polymer disk





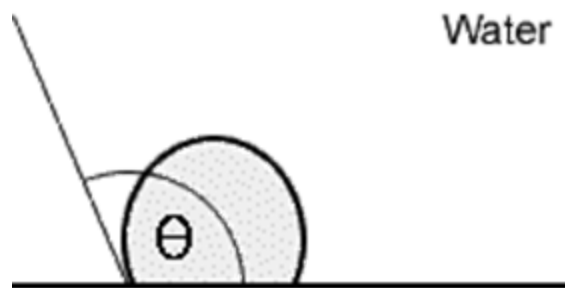
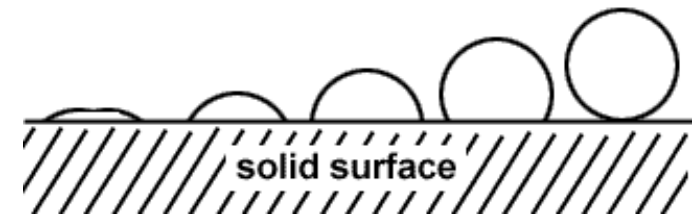
# Surface modification

- Hydrophilic / hydrophobic surfaces
- Wetting/non-wetting droplets
- Fuktende/ ikke-fuktende væsker
  
- Lithographic patterning
- E.g. deposition of self-assembled -monolayers

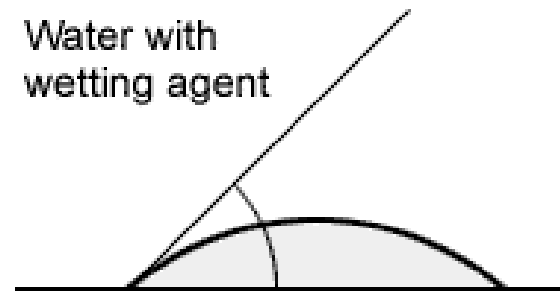


# Wetting / Non-wetting

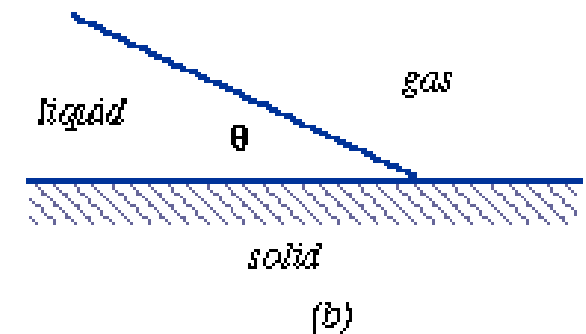
- Contact angle depends on the solid/liquid/gas that meet in one point
- Wetting fluid: Contact angle  $< 90$
- Non-wetting: Contact angle  $> 90$



Water

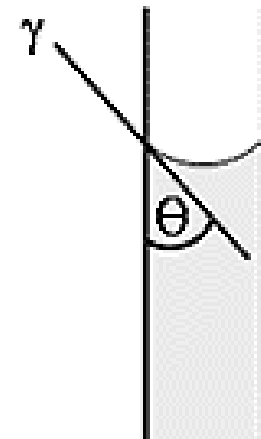
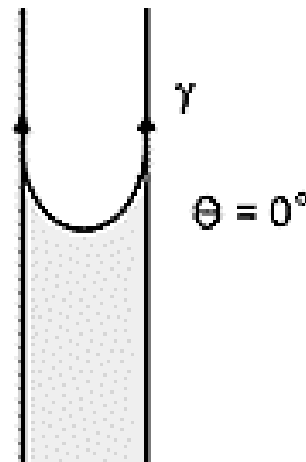
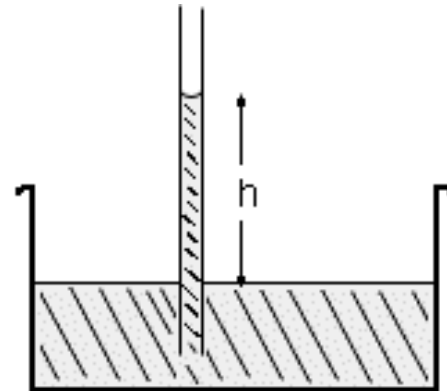


Water with wetting agent



# Definition of wetting angle

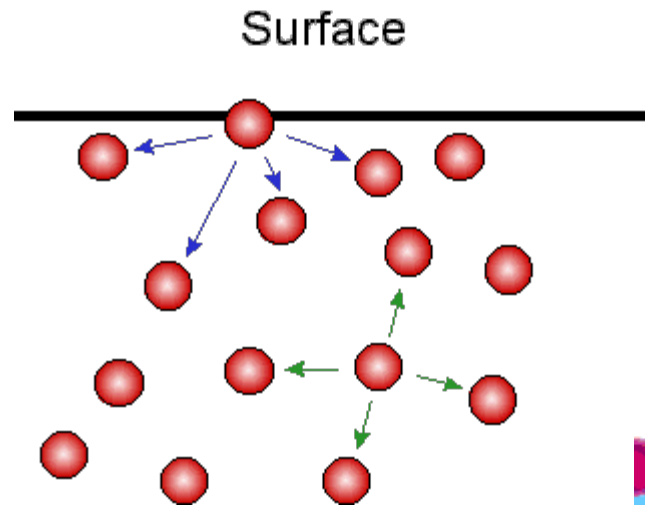
- Can be modified by (chemical) surface treatment



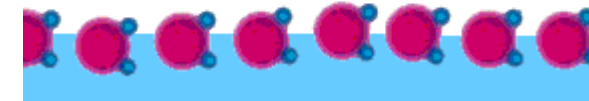


# Surface tension

- Surface between two fluids
  - Gas-Liquid
  - Liquid-Liquid
- Energy per surface area
- Force per unit length [N/m]

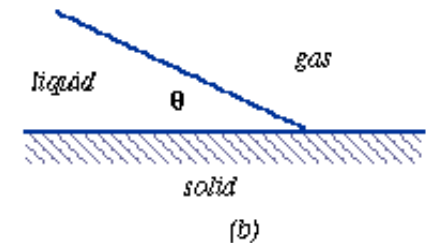
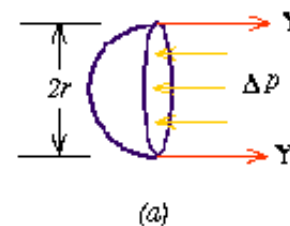


Surface Tension



- Surface tension along periphery
- Pressure on section area

$$2\pi r\Gamma = \Delta P \pi r^2$$



Pressure difference outside/inside drop:  $\Delta P = 2\Gamma / r$



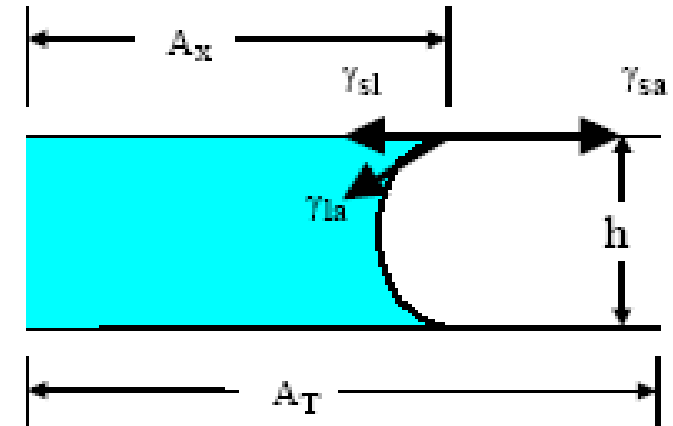
# Equilibrium pressure difference across meniscus in capillary tube

- Young-Laplace equation, circular capillary tube:

$$\Delta p = -\frac{2\gamma \cos \theta}{r}$$

- Young-Laplace equation, rectangular capillary tube:

$$\Delta p = -2\gamma \left[ \frac{\cos \theta_{wall}}{w} + \frac{\cos \theta_{top}}{h} \right]$$



Tube radius  $r$

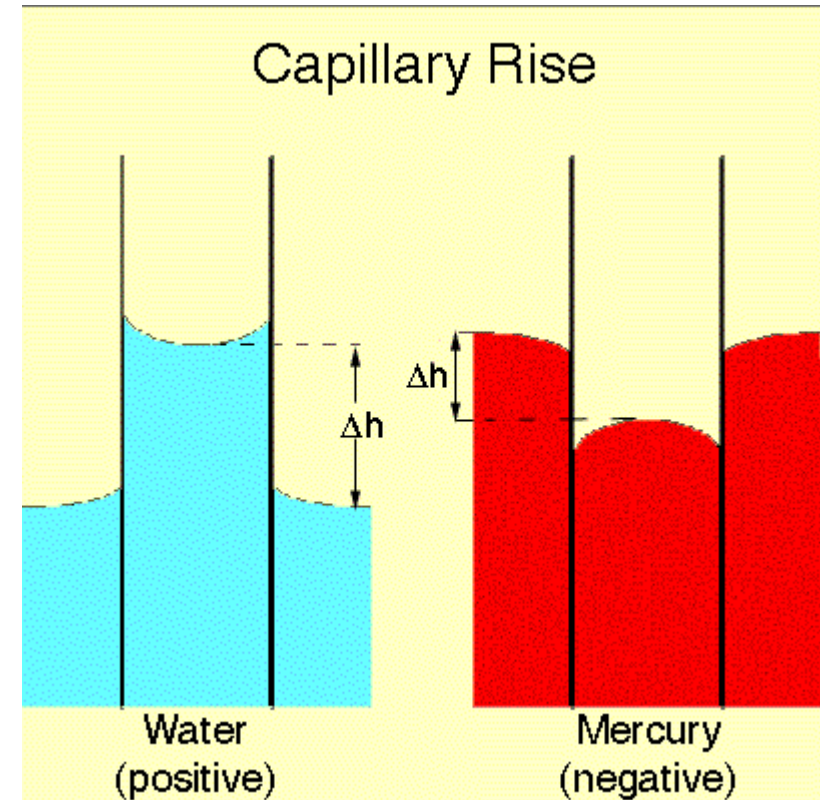
Channel width  $w$

Channel height  $h$

# Capillary rise

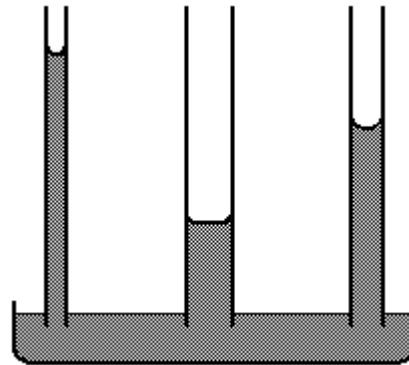
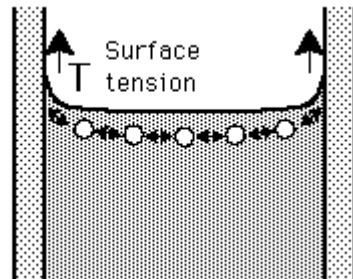
Senturia 13.2.3

- A liquid that wets the walls will rise to a height  $h$  in a capillary tube
- Equilibrium is when weight of liquid column equals surface forces that pull meniscus up
- Forces:
  - Surface forces pull meniscus up  $2\pi r\Gamma \cos\Theta$
  - Gravity pull liquid down  $\rho g h \pi r^2$



$$\rho g h \pi r^2 = 2\pi r \Gamma \cos \Theta$$

Capillary action is the result of adhesion and surface tension. Adhesion of water to the walls of a vessel will cause an upward force on the liquid at the edges and result in a meniscus which turns upward. The surface tension acts to hold the surface intact, so instead of just the edges moving upward, the whole liquid surface is dragged upward.



$$h = \frac{2\Gamma \cos \theta}{\rho g r}$$

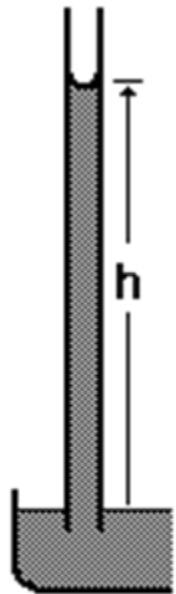
Capillary action occurs when the adhesion to the walls is stronger than the cohesive forces between the liquid molecules. The height to which capillary action will take water in a uniform circular tube is limited by surface tension. Acting around the circumference, the upward force is

The height  $h$  to which capillary action will lift water depends upon the weight of water which the surface tension will lift:

The height to which the liquid can be lifted is given by

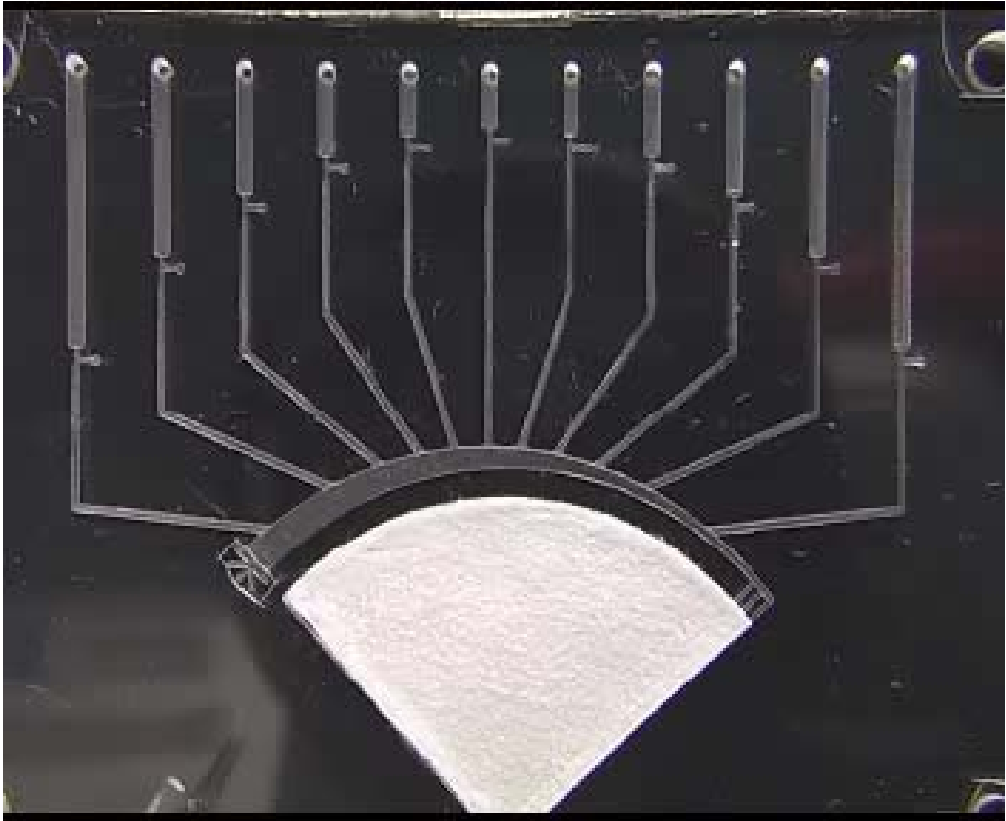
$$T 2\pi r = \rho g (h\pi r^2)$$

$$h = \frac{2T}{\rho r g}$$



Since it is weight limited it will rise higher in a smaller tube

# Capillary filling



# Manufacturing of microfluidic chips

## ■ Plastic

- Master manufactured in silicon with lithographic methods, then in nickel by electroplating
- Use foundry (støpeform) for plastic forming
- Hot embossing
- Injection moulding

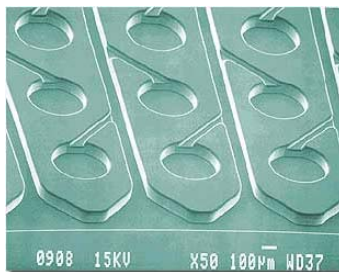
## ■ Glass

- Lithographic patterning of resist on e.g. gold chromium
- Wet etch of glass

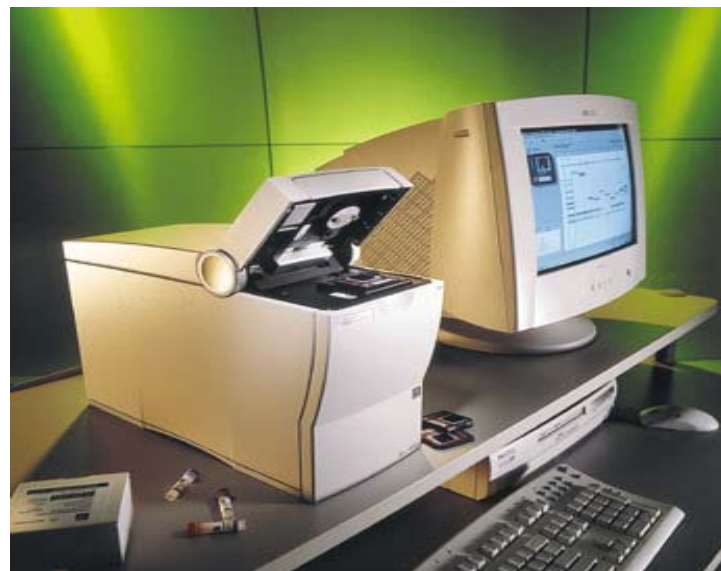
## ■ Sealed by glass or plastic film -> bonding or lamination

## ■ Typical depth of microfluidic channel: 10-100 $\mu$ m

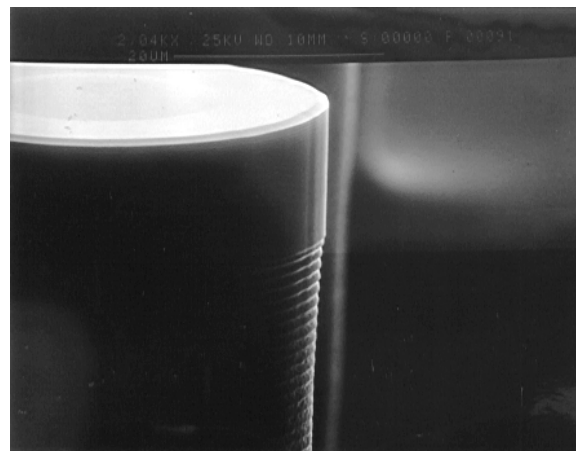
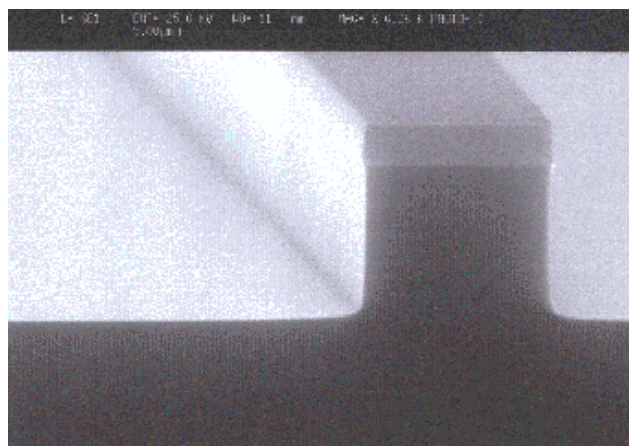
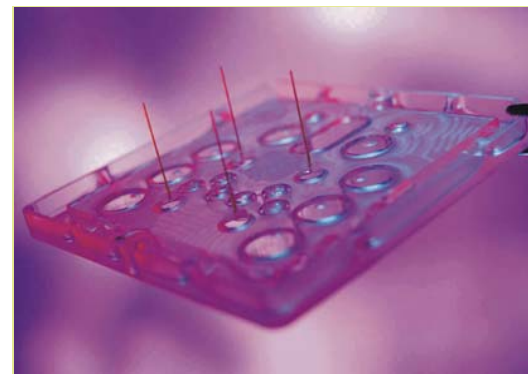
# Mikro-kanaler



- Kanaler med vertikale vegger i silisium
- Sprøytetøping av plast  
Støpeformene generert f.eks. via silisium + elektroplatering



Caliper

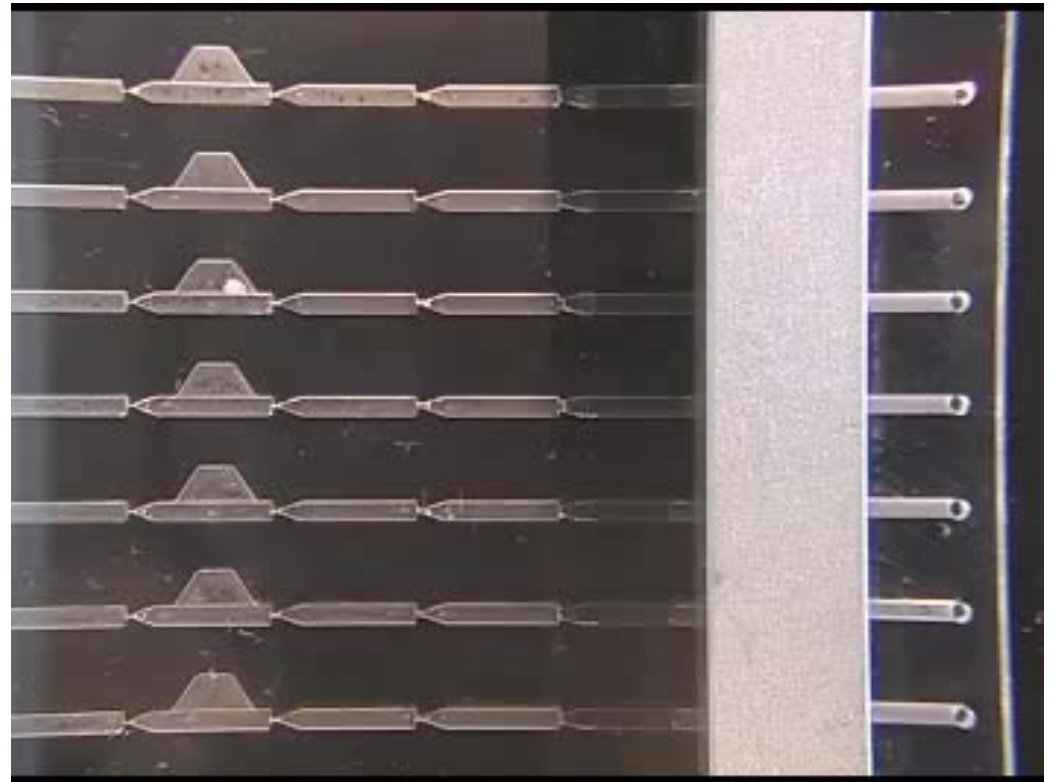


New:  
Deep reactive ion  
etch DRIE,  
BOSCH process



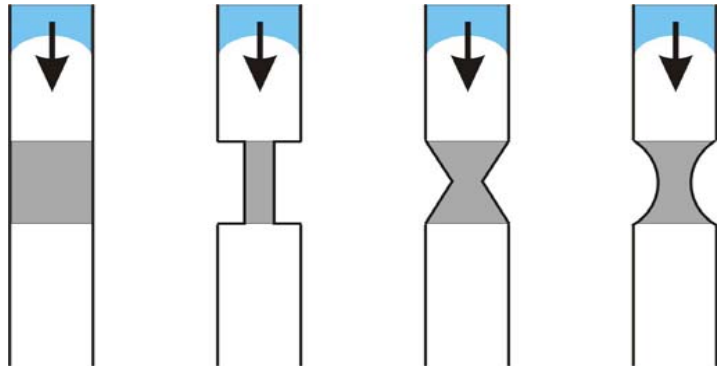
# Parallel plug displacement chip

- 7 liquid plugs in parallel
- Plug volume  $\sim 1\mu\text{l}$
- Working liquid: DI water
- Teflon spotted hydrophobic valves
- Single pump source, pump velocity  $10\mu\text{l}/\text{min}$

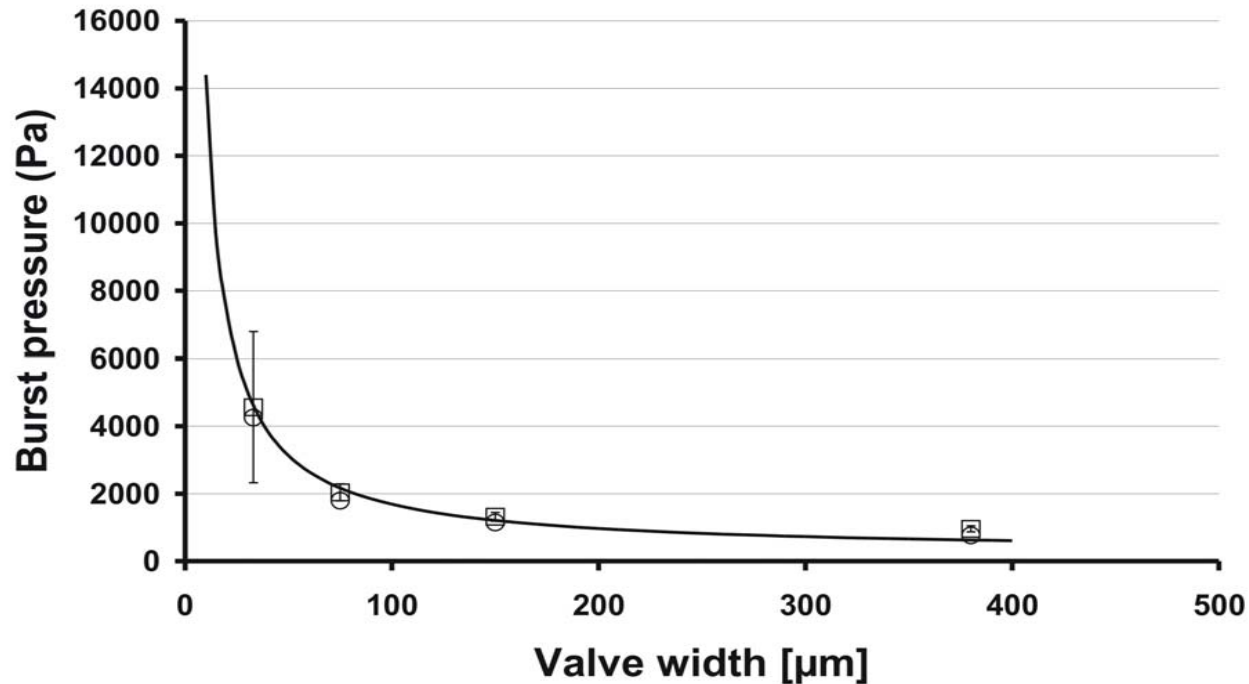


# Capillary valving (surface manipulation to change wetting properties)

- Grey –teflon
- Hydrophobic valves
- Small restriction – high pressure required to “break” the valve
- Can efficient flow control using a single pressure source for many parallel channels be achieved?



# Burst pressures water, reagents



Burst pressures of the capillary valves as function of valve width. Symbols: □ DI water; ○ reagents; solid line represents the analytical values for water. Contact angle of DI water on Teflon was measured to be approx.  $110^\circ$ . The pressure data for each valve represents an average of 35 measurements (5 chips with 7 parallel channels each).

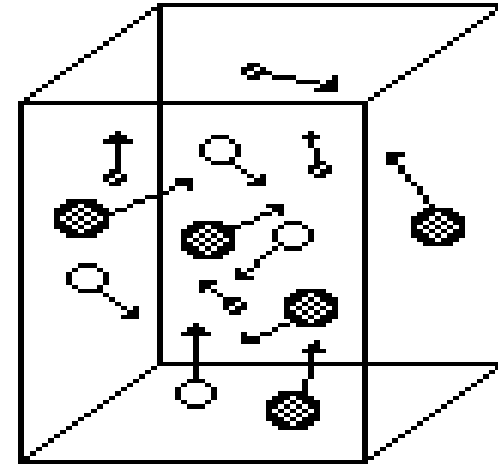
# Ideal Gas Law

- Equation of state for (ideal) gases
- $pV=NkT$
- $k=1.38 \cdot 10^{-23}$  J/K, Boltzmann constant

- Senturia:

$$P = \rho_m \left( \frac{R}{M_w} \right) T$$

- $R=8.31$  J/(mol K), universal gas constant



State variables

V volume

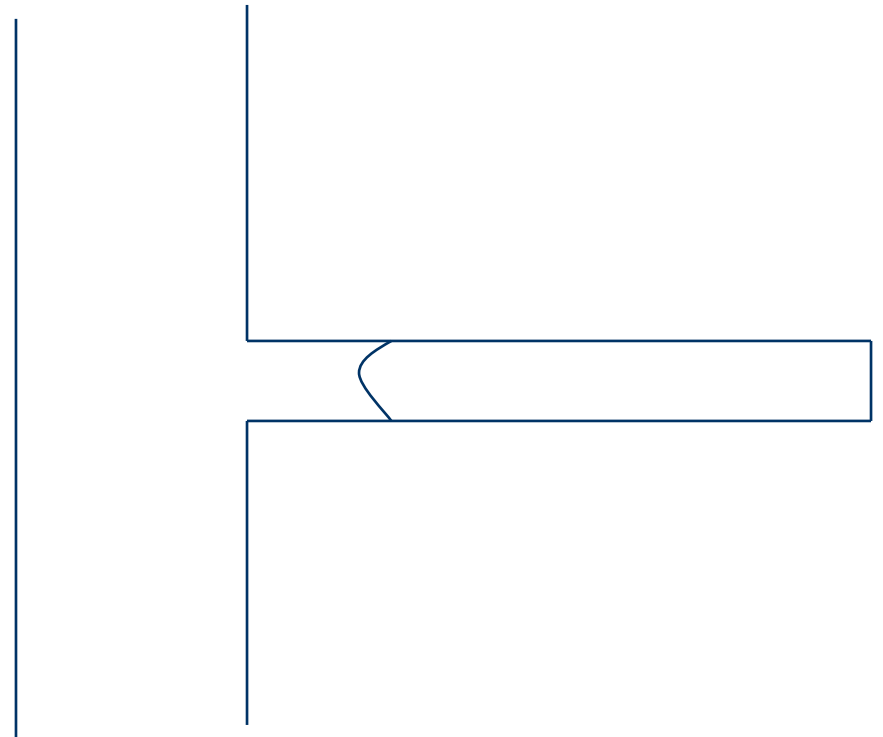
P absolute pressure

T absolute temperature

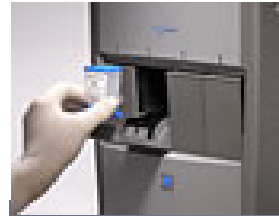
# Exercise:

## Fluid volume in capillary “dead-end”

- Where does the meniscus stop?
- What is the volume that is pulled into the narrow capillary?
- What happens when temperature is increased?



- Molecular diagnostics
  - Cancer
  - Infections (bacteria, virus, parasites)
  - Cardiovascular diseases
- Molecular markers
  - DNA, RNA
  - Proteins; antigens, enzymes, hormones
  - Low molecular compounds
- Sample preparation
  - filters, micro-pillars, magnetic beads, separation
- Washing
- (Amplification e.g. PCR)
- Reactions
  - Immunoreactions
  - Hybridization
- Detection
  - Labels (dye, fluorescent, radioactive)
  - Label-free (impedance, electrochemical, amperometric, cantilevers, evanescent fields)
- Choose methods for all steps:  
SENSITIVITY + SPECIFICITY



Cepheid  
GeneXpert  
technology

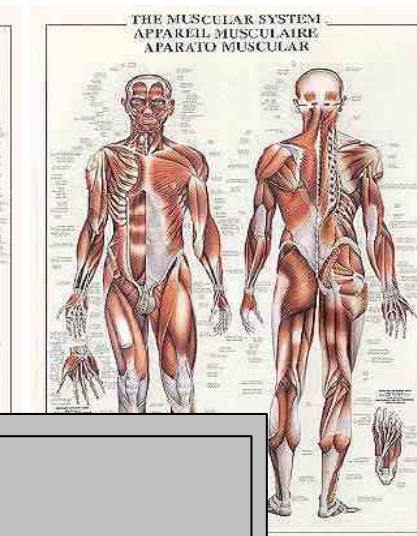


Roche AmpliChip  
Pharmacogenetic  
microarray based  
test

The main objective for The MicroActive project is to develop an instrument for molecular diagnostics intended for use in the doctor's office

- Human papilloma virus
- Cervical cancer
- mRNA
- Two microchips
- Instrument

- Cell sample in, diagnosis on 10 HPV viruses out



NORCHIP



# A sample preparation chip has been manufactured

- Purification of nucleic acids
- Start material (5 ml): liquid based cytology
- Output (20  $\mu$ l): mRNA suitable for NASBA amplification
- Functions
  - Lysis buffer, wash buffers, elution buffer stored on-chip
  - Cell filter
  - Nucleic acid capture filters
- Chip output has successfully been amplified by NASBA



Biomerieux, Qiagen



MicroActive chip (IMM)

# A NASBA amplification and fluorescent detection chip has been manufactured

Input: 20  $\mu\text{l}$  of purified nucleic acids

Split fluid volume into 10 droplets of 500 nl

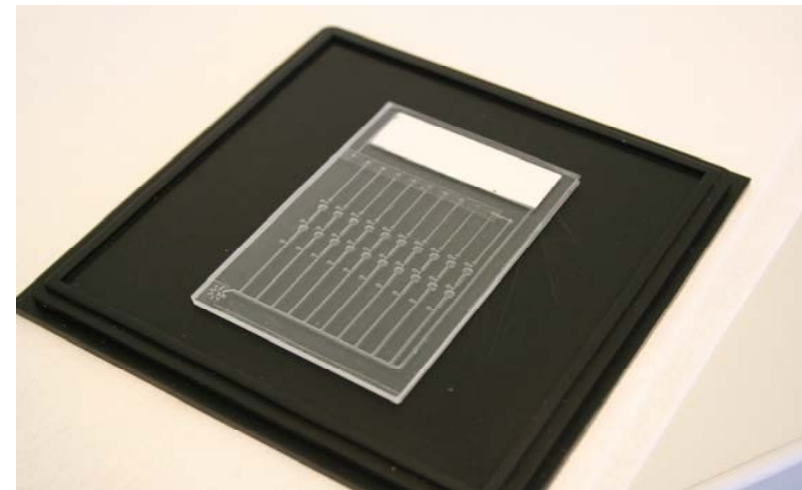
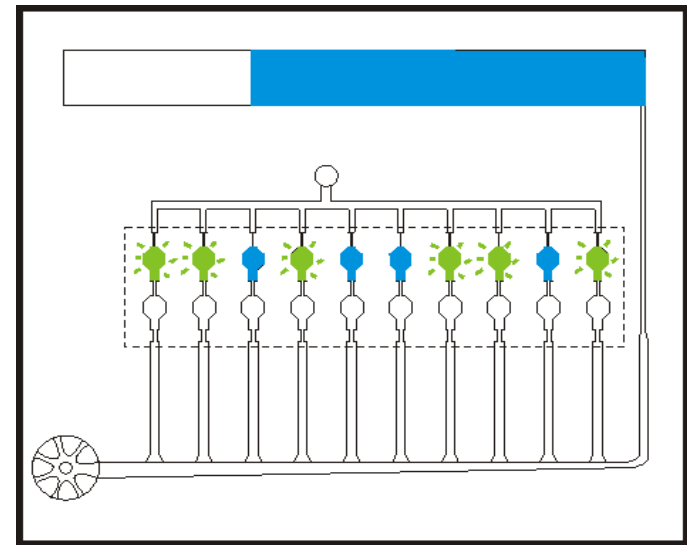
Primers for each HPV type and NASBA enzymes stored in dry state on chip

3 droplet stop positions controlled by hydrophobic patches in channels

Metering

Dissolution of master-mix reagents

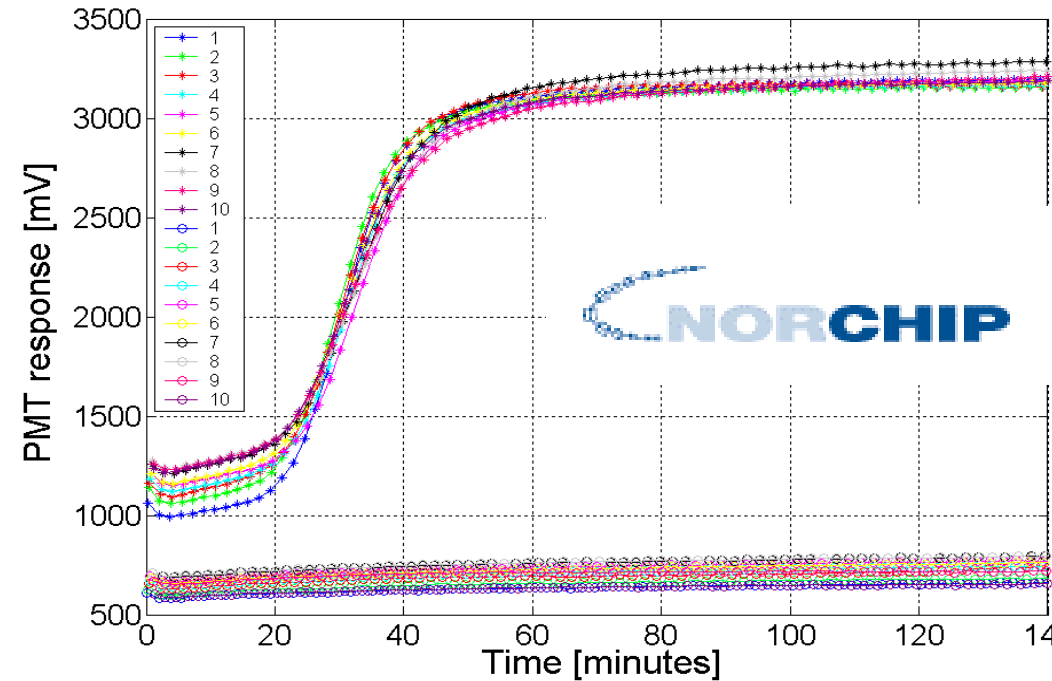
Dissolution of enzymes and detection



SINTEF injection molded chip

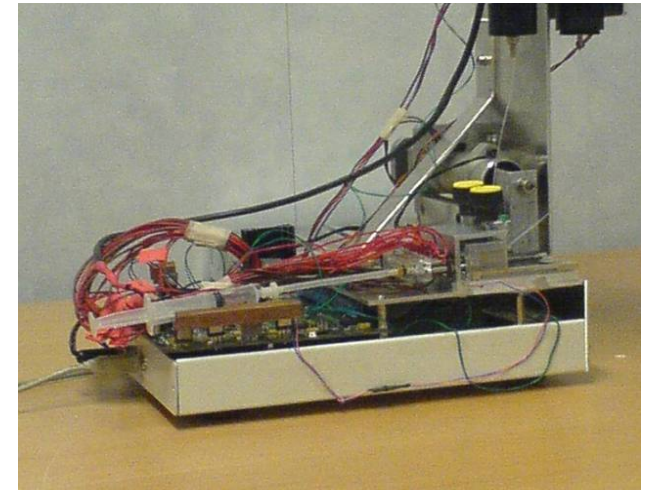
# NASBA of HPV-16 mRNA in 500 nl plugs in microchip

- Optimization of drying agents
- Wall roughness
- Wall coating
- Re-hydration of dried reagents
  
- NASBA amplification in sample plugs using dried enzymes

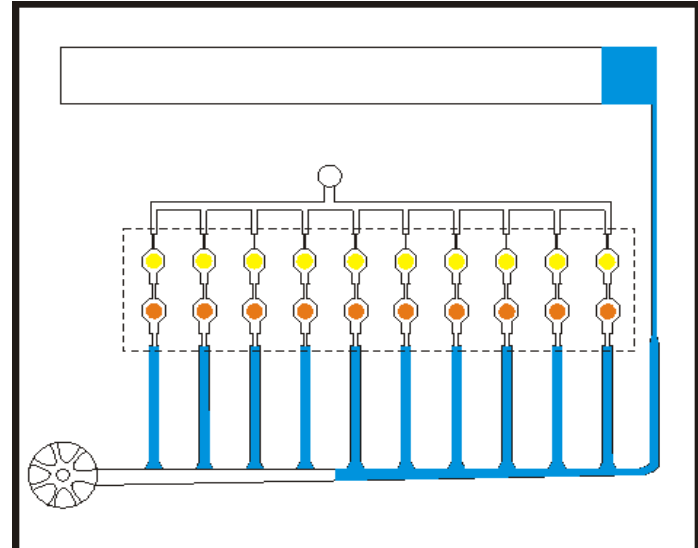
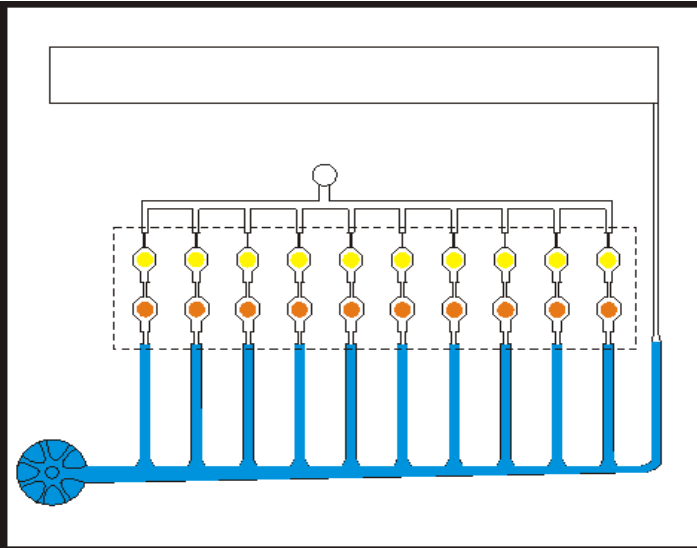


## Instrument

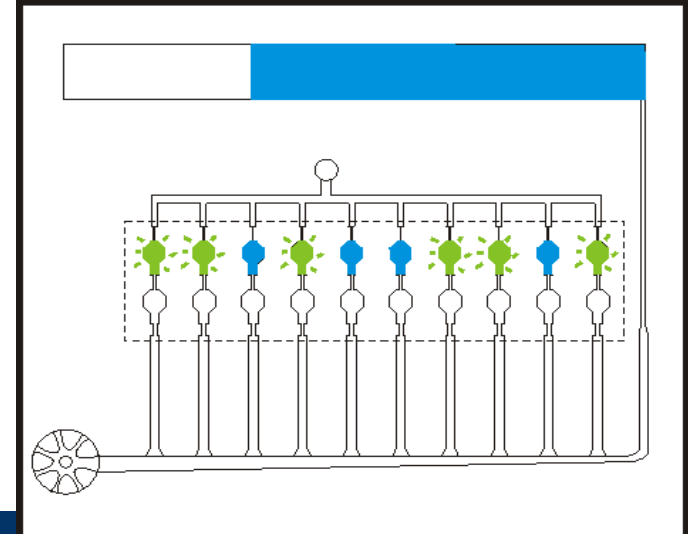
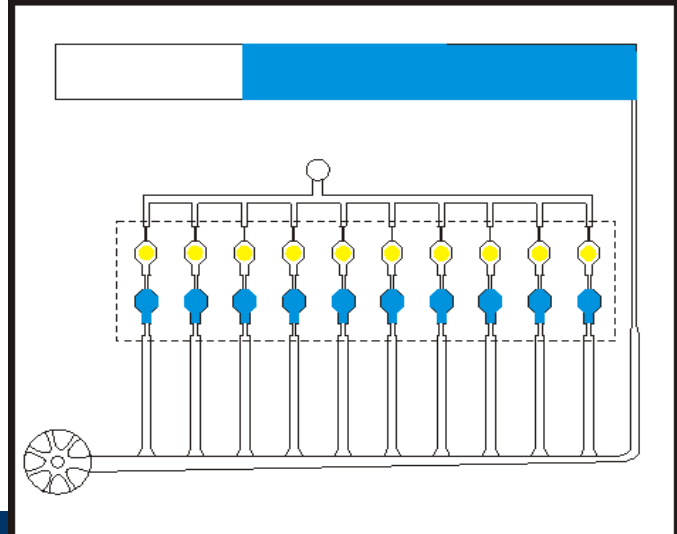
- Pump
- Heating (for amplification)
- Fluorescent readout of 10 chambers



# Cartoon of amplification chip function



- Sample insertion
- Splitting into channels
- Metering
- Mix reagents 1
- Mix reagents 2
- Amplification
- Detection



# Cepheid GeneXpert

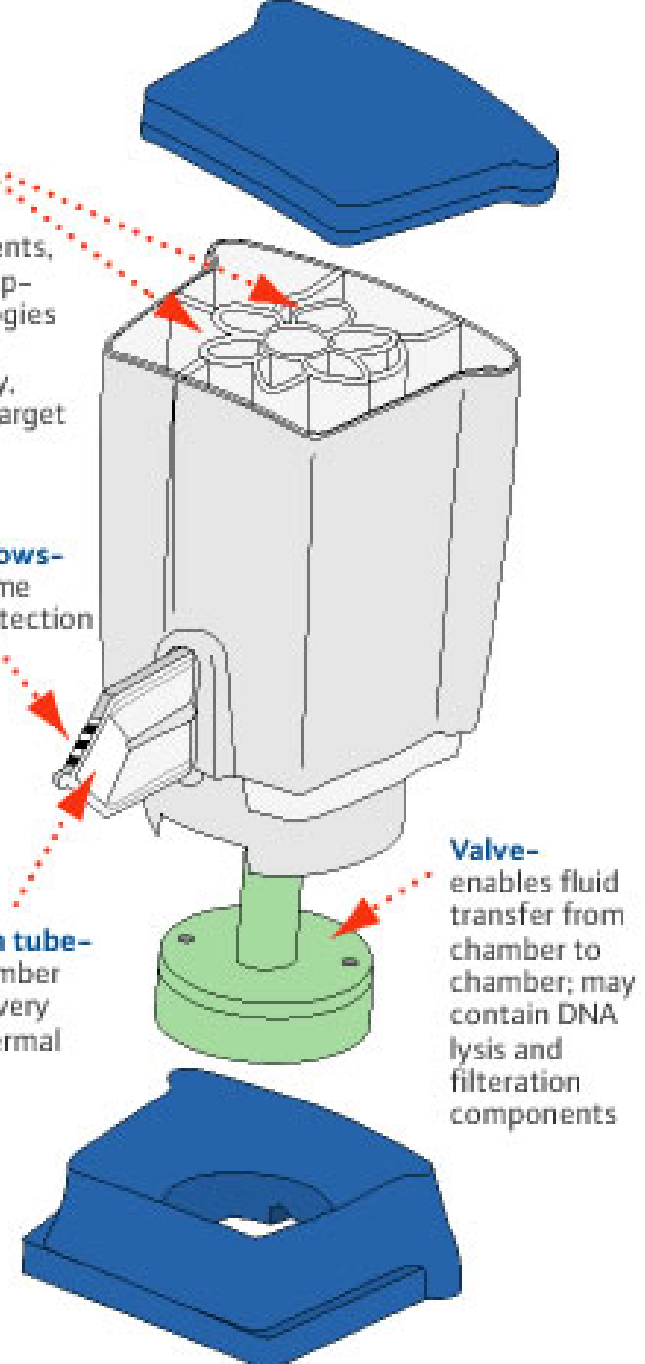


**Processing chambers-** contain reagents, filters, and capture technologies necessary to extract, purify, and amplify target DNA

**Optical windows-** enable real time four-color detection

**Reaction tube-** thin chamber enables very rapid thermal cycling

**Valve-** enables fluid transfer from chamber to chamber; may contain DNA lysis and filtration components



# Viscosity

## Senturia 13.2.1

- Deformation of fluids in the presence of shear forces
- The property of a fluid that resists the action of a shear force
- $\eta$  [ Pa s ]
- Newtonian fluid:

$$\tau = \eta \frac{U}{h}$$

$$\tau = \eta \frac{\partial U_x}{\partial y}$$

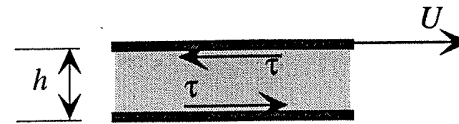
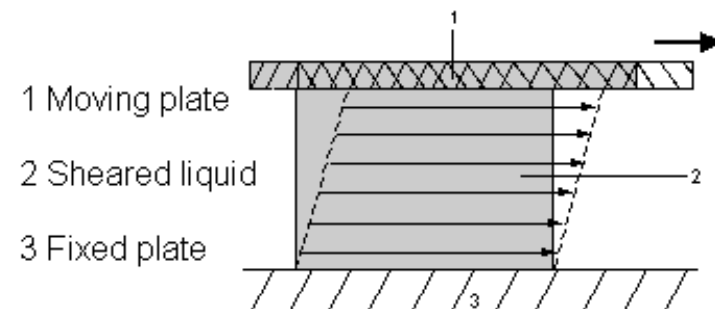


Figure 13.1. Fluid between two plates. The upper plate moves to the right with velocity  $U$ , setting up shear forces  $\tau$ .



# Navier-Stokes equation

- Conservation of mass

$$\frac{\partial \rho}{\partial t} + \nabla \cdot (\rho \vec{v}) = 0$$

- Newton's 2nd law for a fluid

$$\rho \left( \frac{\partial \vec{v}}{\partial t} + (\vec{v} \cdot \nabla) \vec{v} \right) = \nabla p + \eta \nabla^2 \vec{v}$$

- Incompressible flow

$$\rho \frac{\partial \vec{v}}{\partial t} = -\nabla P + \rho g + \eta \nabla^2 \vec{v}$$



# Reynolds number

$$\text{Re} = \frac{\rho UL}{\eta}$$

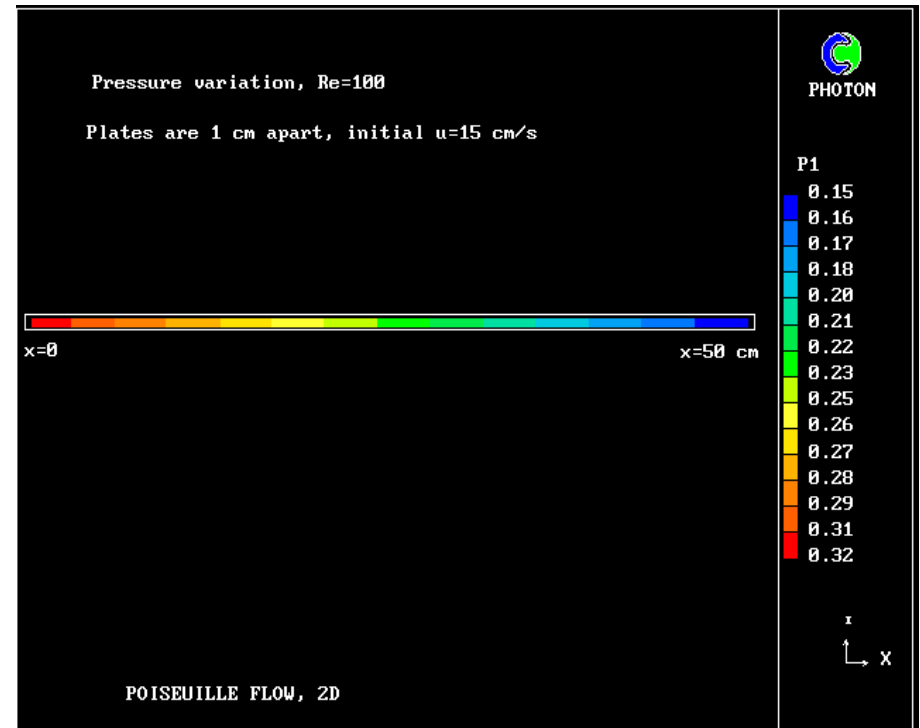
- Laminar or turbulent flow?
- Ratio of inertial forces to viscous forces
- Reynolds number:  
ratio of kinetic energy of a volume of fluid in the flow  
to  
the energy dissipated by the volume in the shear caused by interaction with its solid boundaries

- Microchannel:
  - 1 cm long
  - 1 mm wide
  - 100  $\mu\text{m}$  deep
- $L=50 \mu\text{m}$
- $\rho=1000 \text{ kg/m}^3$
- $\eta=0.001 \text{ kg/ms}$

Laminar for flow speeds less than 10m/s

# Poiseuille flow

- Pressure driven flow in channel
- Pressure drop along channel
- Steady flow
- Incompressible flow
- Flow in x-direction, only
- No-slip boundary equations



$$\eta \frac{\partial^2 U_x}{\partial y^2} + \frac{\Delta p}{L} = 0$$

Integrate twice:

$$U_x(y) = -\frac{1}{2\eta} \frac{\Delta p}{L} y^2 + c_1 y + c_2$$

No slip boundary condition gives:

$$U_x(y) = \frac{1}{2\eta} \frac{\Delta p}{L} \left[ \left( \frac{a}{2} \right)^2 - y^2 \right]$$

Flow rate:

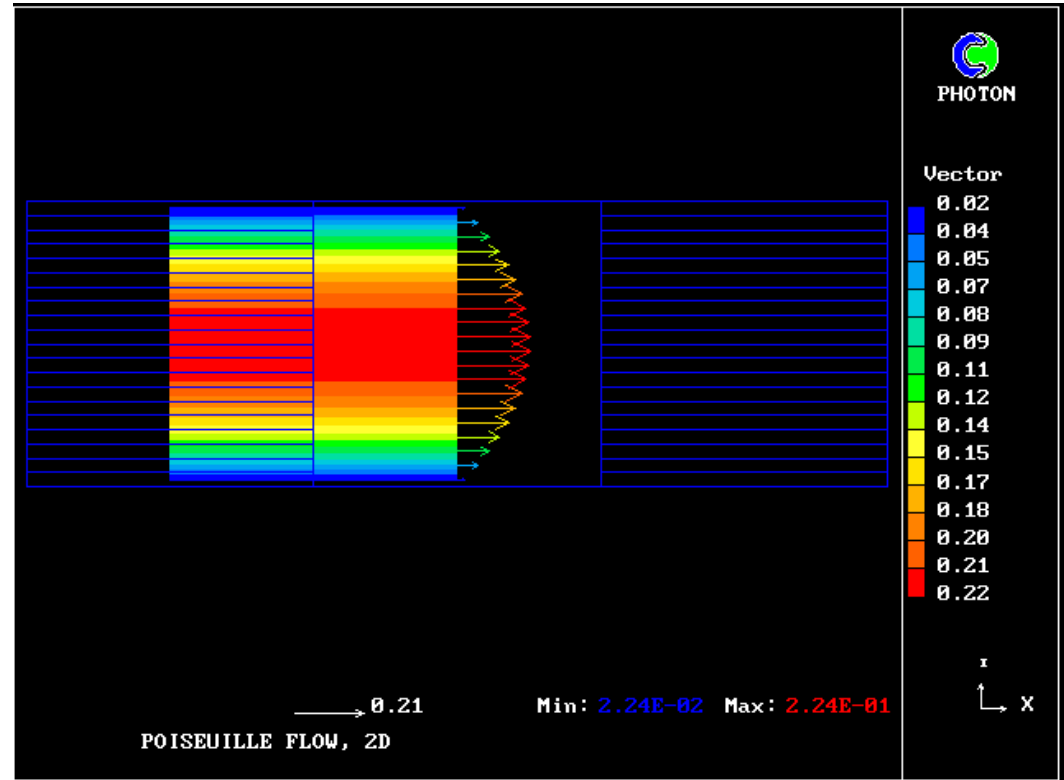
$$Q = \int_0^{l_z} dz \int_{-a/2}^{a/2} U_x(y) dy$$

$$Q = \frac{l_z a^3}{12\eta} \frac{\Delta p}{L}$$

Circular pipe:

$$Q = \frac{\pi a^4}{8\eta} \frac{\Delta p}{L}$$

# Poiseuille flow



# New Micro Flow Rate Sensor for Standardized Industrial Production

3  $\mu\text{m}$



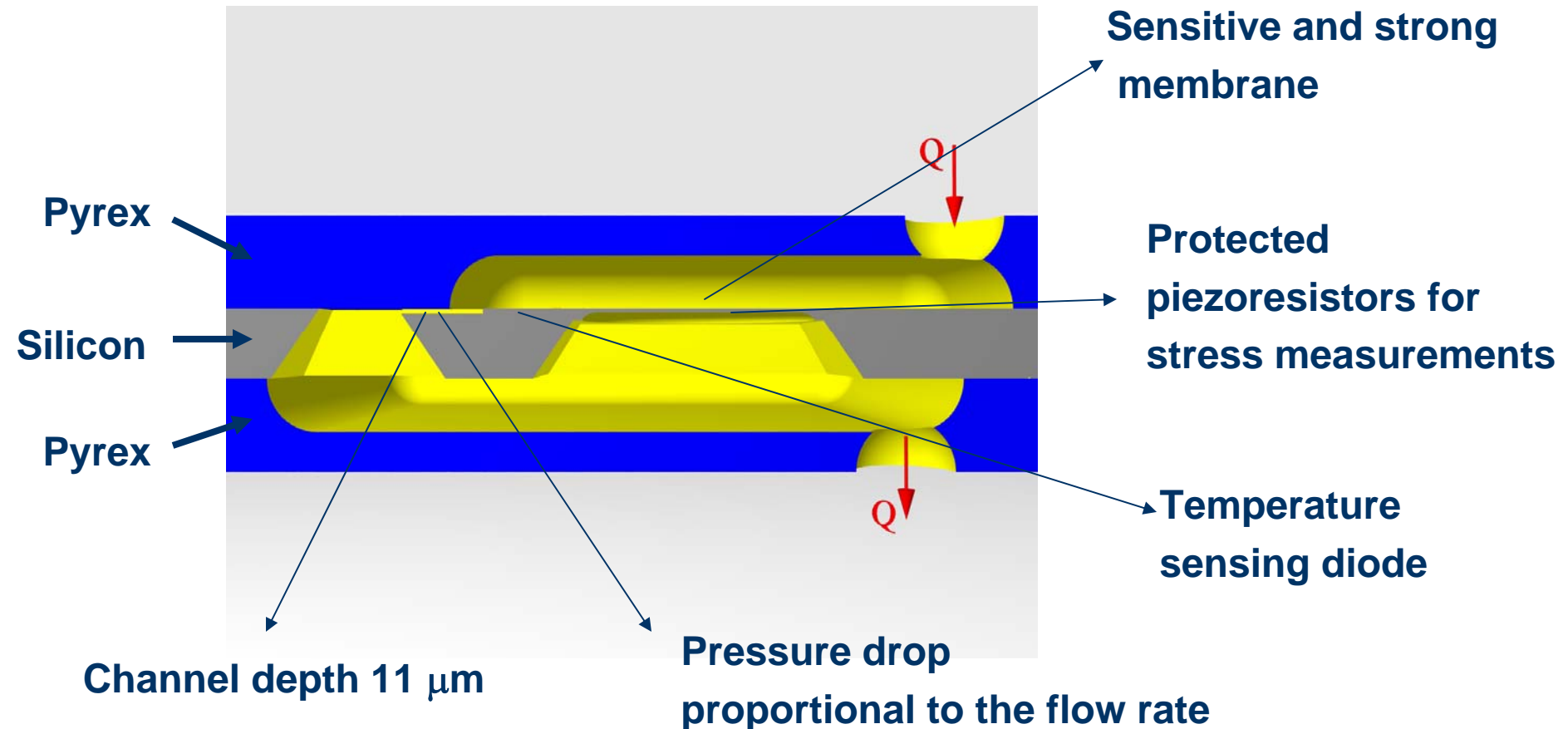
6 mm



Liv Furuberg  
Dag Wang  
Andreas Vogl

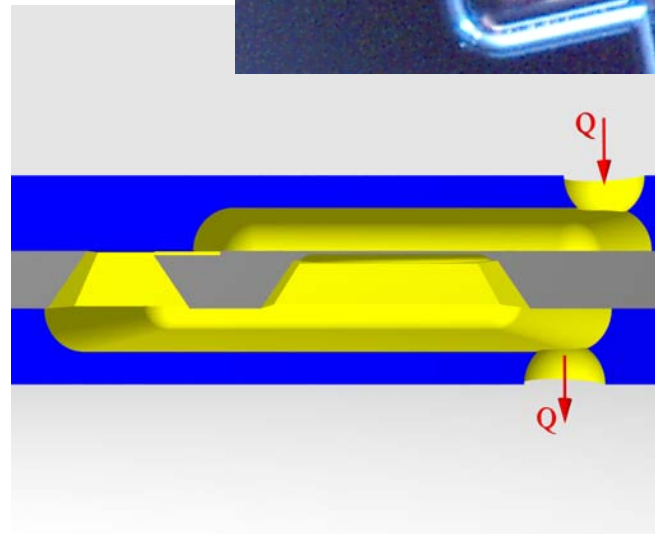
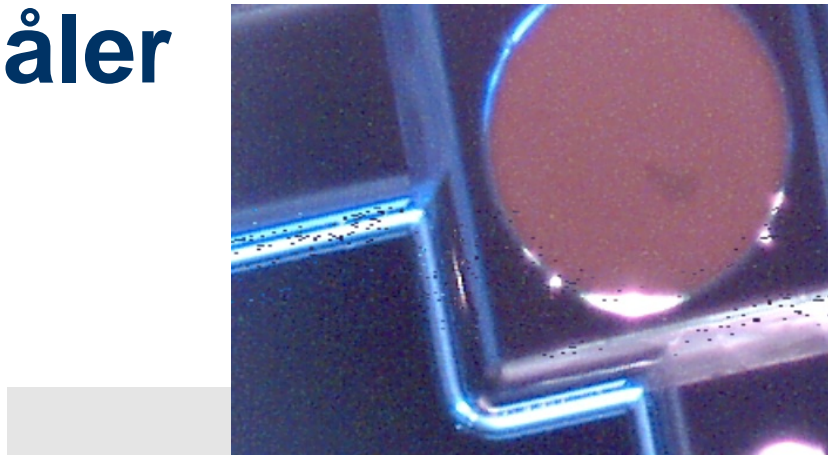
Microsystems and Nanotechnology  
SINTEF Information and Communication Technology

# The new design suggests a low-noise, mechanically robust flow sensor



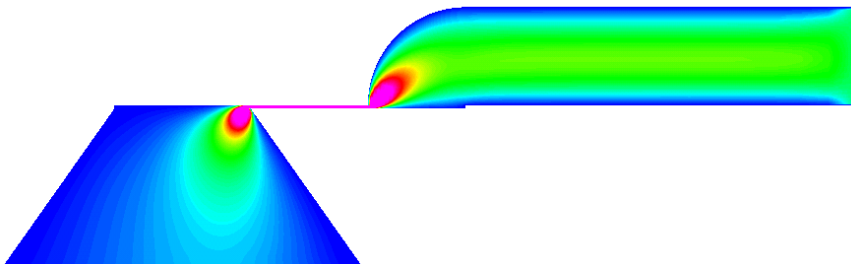
# Volum-strømningsmåler

- Applikasjoner: Dosering, tilføring av reagerer, måle flow gjennom analysesystem
- Væskestrøm gjennom brikken
- Glass-silisium-glass brikke
- Laminær strøm, lave Re tall
- Differensiell trykksensor (membran + piezomotstander)
- Trang kanal med trykkfall, Pouseille strøm
- Trykkfall ~ 100 -200 Pa
- Integrert temperaturmåler



- Kanal: 800x1500x10  $\mu\text{m}$
- Flow rate 2  $\mu\text{l}/\text{min}$

$$\Delta p = \frac{12 \cdot \eta \cdot l \cdot Q}{w \cdot h^3}$$



# Electroosmotic Flow

- Flow driven by electric field
- Voltage applied between electrodes immersed in electrolyte
- Force on fluid near the boundaries, excess of charged particles
- Debye screening layer, typically 10nm

- Disadvantages:

- Sensitivity to impurities
- Ohmic generation of heat
- Need for high voltages

- Advantage:

- Plug flow

Solving Navier Stokes

$$\Omega^0 = \frac{\mu}{\rho^m \epsilon^x \gamma^D}$$

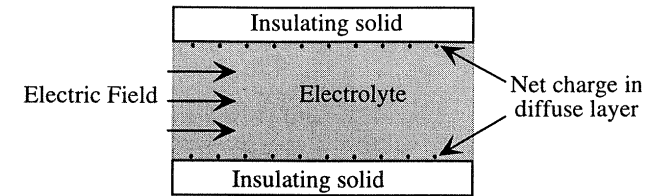


Figure 13.11. Illustrating electroosmotic flow

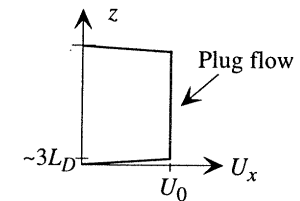


Figure 13.12. Electroosmotic flow profile.



# Poiseuille flow vs. electroosmotic flow

Advantage in 3D visualization/detection

Three pictures after creation of fluorescent molecule:

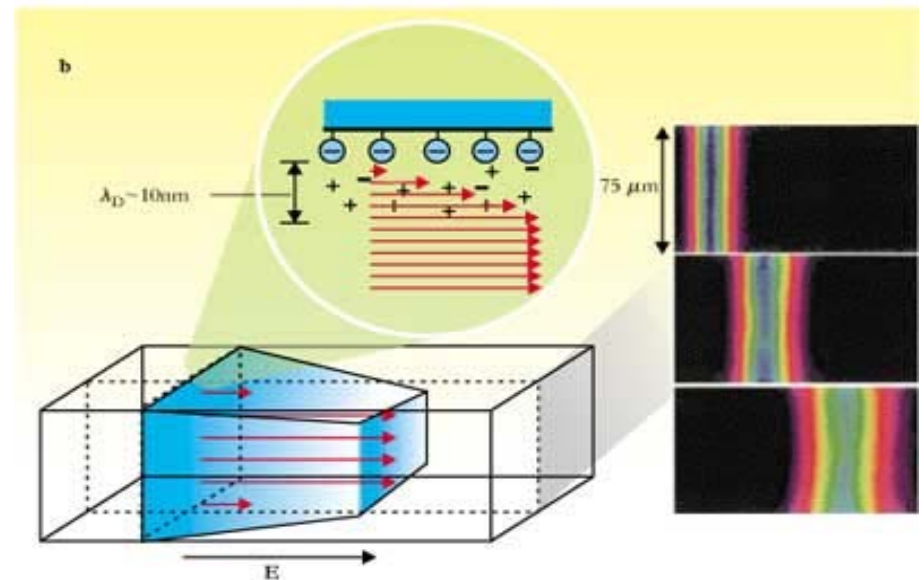
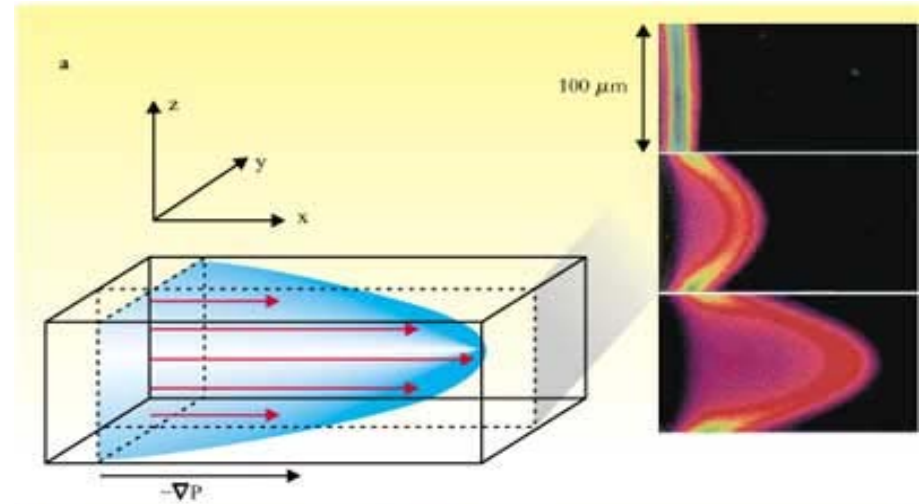
0s

66ms

165ms

Separation based on charge-to-size ratio of molecules.

Separated bands of species



# Electrophoresis

- Species carried along with electroosmotic flow
- Drift relative to the moving velocity:

$$v_{ep} = \mu_{ep} \mathcal{E}_x$$

- Electrophoretic mobility
- Apply voltages to channels
- Create controlled plug of species
- Separate molecules by charge and volume by electrophoresis

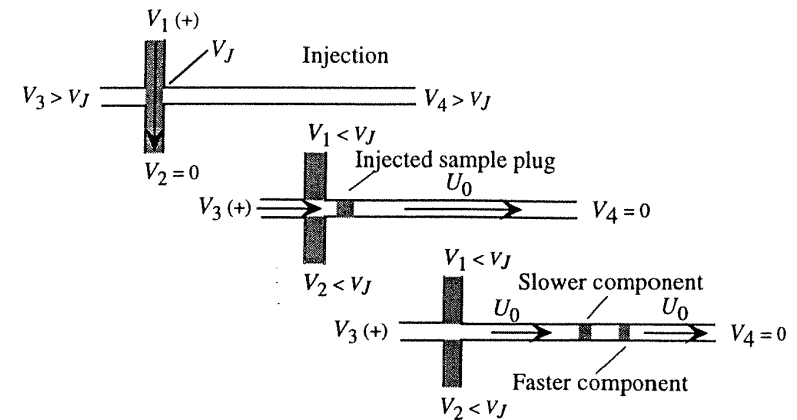


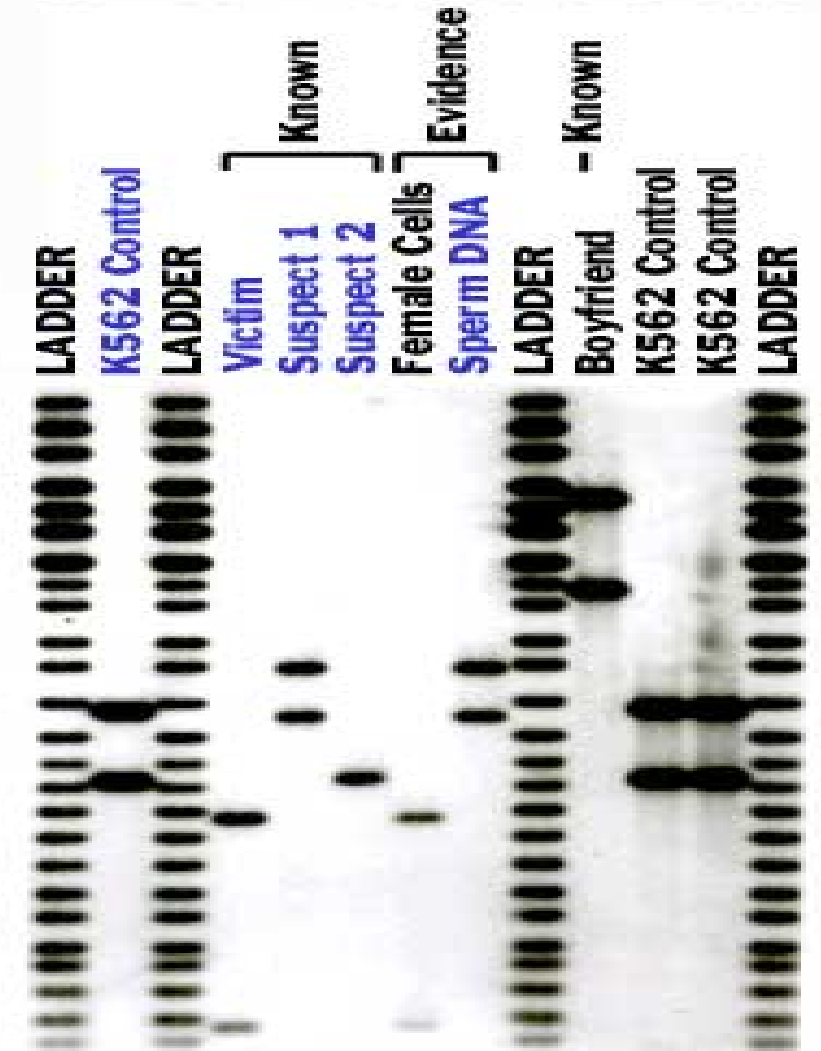
Figure 13.14. Illustrating electrophoretic separation with electroosmotic flow. The voltages used during the injection and separation sequence are described in the text.

# Suspect - DNA analysis

- DNA fingerprint



## Sexual Assault Case

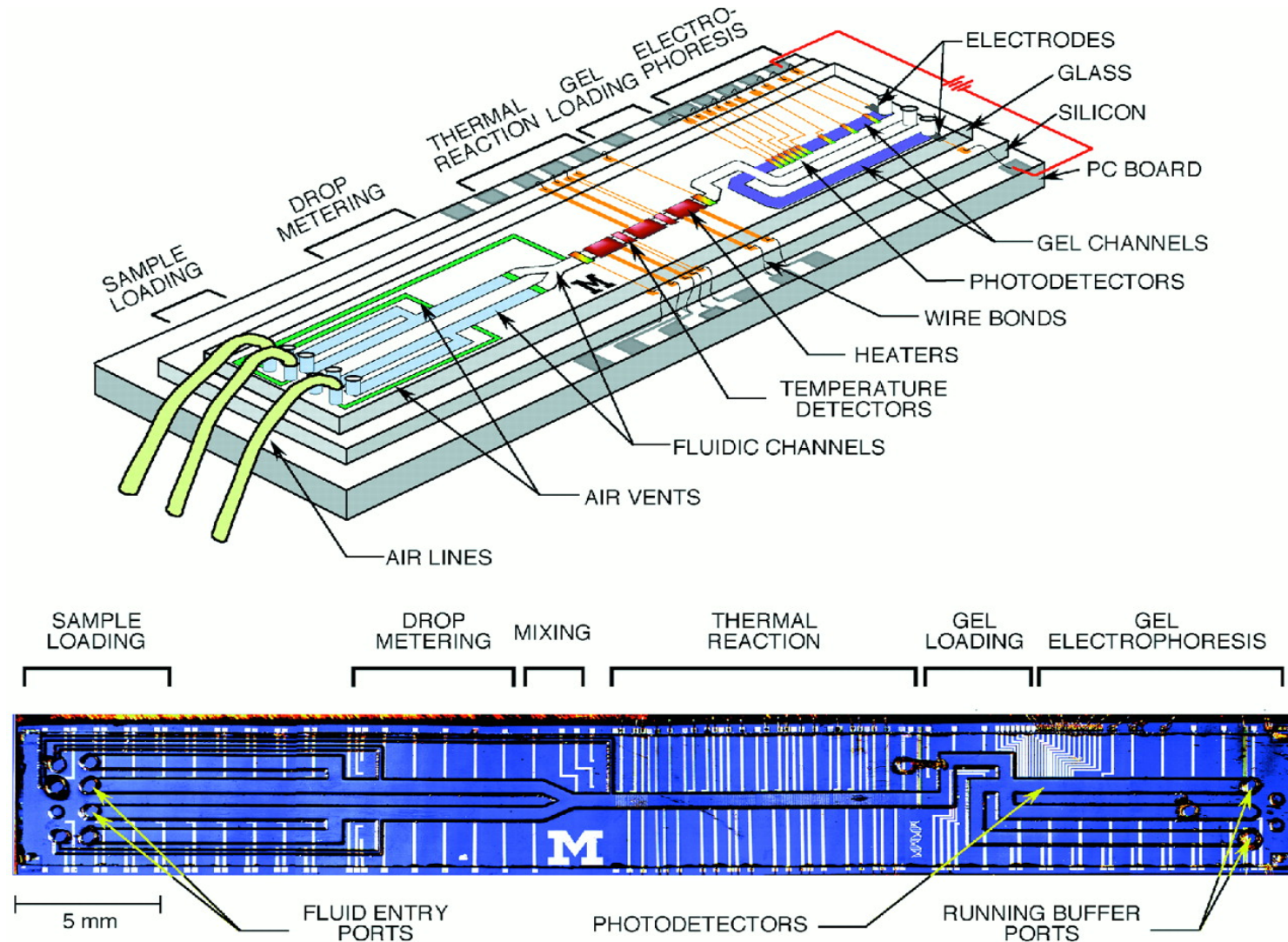


© 2001. How Stuff Works

# Burns et al., droplet based total analysis system

## “An Integrated Nanoliter DNA Analysis Device”

*Science* 16  
 October 1998:  
 Vol. 282. no.  
 5388, pp. 484 -  
 487



# Mixing

- Laminar flow
- Mixing by diffusion only

- Diffusion equation

$$\frac{\partial C(r,t)}{\partial t} = D\nabla^2 C(r,t)$$

- Average displacement of diffusing particle:

$$l = \sqrt{4Dt}$$

- Diffusion constant for water

$$D = 2.3 \cdot 10^{-9} \text{ m}^2 / \text{s}$$

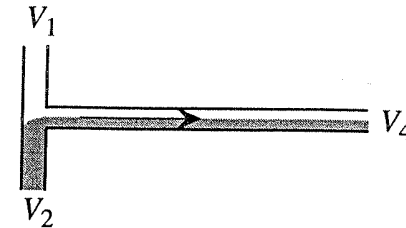


Figure 13.16. Illustrating laminar flow when two streams are combined. Mixing occurs only by diffusion.

- Water: Diffusion length after 1 s: 90 $\mu\text{m}$
- On the other hand:
- Characteristic lines become blurred...
- What about larger molecules?

# Diffusion of ink

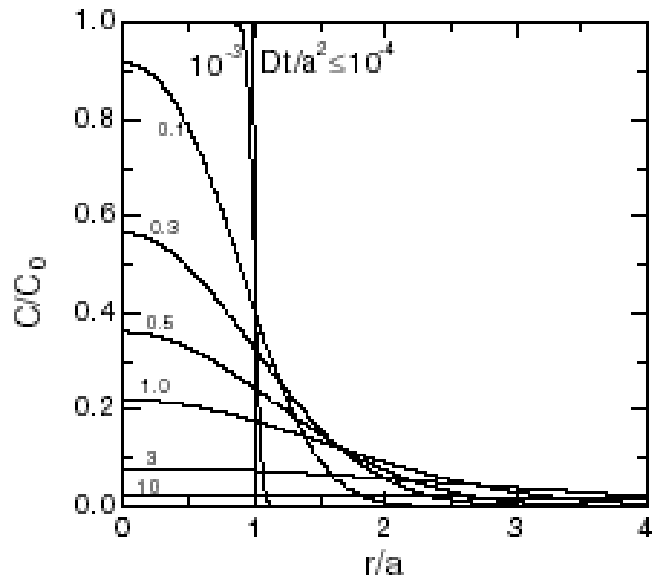


Figure 1. The concentration distribution at various times.

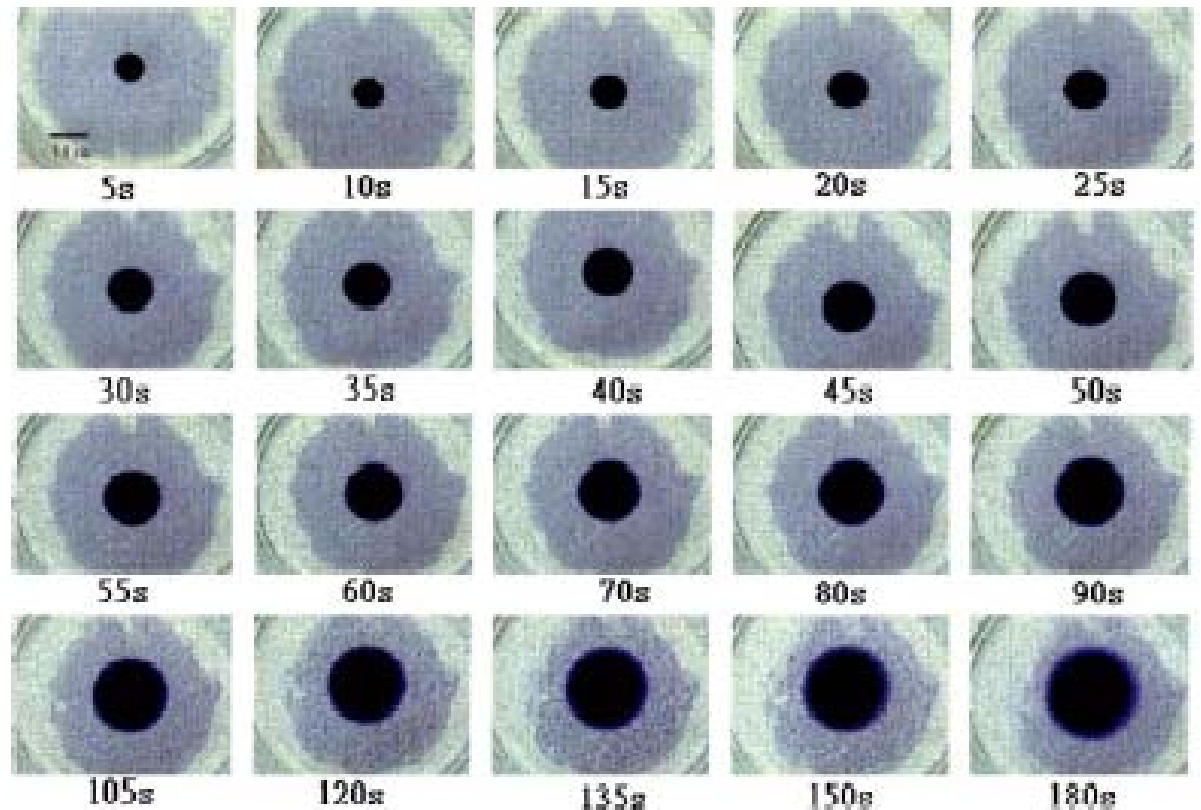


Figure 2. Sequential pictures of stamp ink diffusing in water.

# A simple diffusion-based filter, the H-filter

Micronics.net

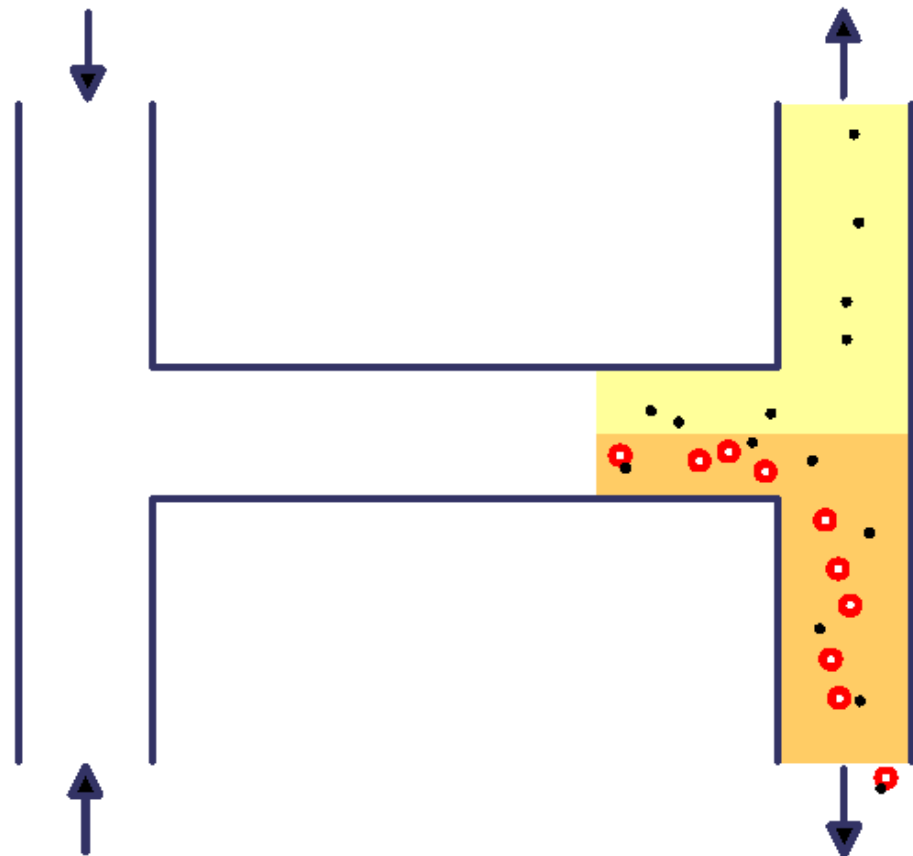
Large and small molecules in  
via lower channel

Clean solvent in via upper  
channel

Large molecules have a  
smaller average diffusion  
distance than small

More small molecules will go  
up

Repeat





# Mixing

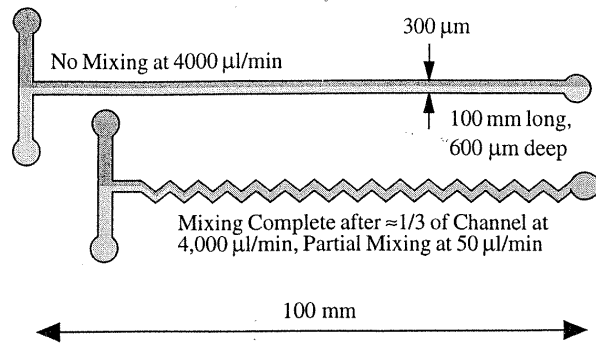


Illustration of miniature fluidic channels used to compare mixing in macroscopic and microscale fluidics. After Branebjerg, et al. (1994).

